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Abstract

Periodontal ligament derived stem cells (PDLSCs) are capable of differentiating into multiple cell types and inducing a promising immunomodulation for tissue regeneration and disease treatment. However, it is still challenging to develop a practical approach to activate endogenous stem cells for tissue self-healing and regeneration. In this study, transcriptome analysis reveals that resveratrol promotes PDLSC stemness through activation of stem cell, osteoprogenitor, and chondroprogenitor markers. Self-renewal and multipotent differentiation abilities are also improved in resveratrol-treated PDLSCs. In addition, immunomodulation of PDLSCs is dramatically increased after resveratrol treatment. Mechanistically, we show that resveratrol activates ERK/WNT crosstalk through elevation of olfactory and growth factor signaling pathways to upregulate the expression levels of RUNX2 and FASL for osteogenesis and immunomodulation, respectively. By using a periodontitis animal model, administration of resveratrol partially rescues bone loss through activation of endogenous somatic stem cells and inhibition of inflammatory T-cell infiltration. Taken together, our findings identify a novel pharmacological approach to achieve autotherapies for endogenous tissue regeneration.

Keywords: periodontal ligament derived stem cells (PDLSCs), regenerative medicine, resveratrol, immunomodulation, autotherapy, periodontitis

Introduction

Tissue regeneration is a dynamic remodeling process through the external stimulations and the internal processes. To maintain body homeostasis, continual tissue renewal and regeneration are necessary, which are mainly attributed to adult stem cells (Zhang et al. 2016). Mesenchymal stem cells (MSCs), originating from both the mesoderm and the neural crest, constitute a population of self-renewal stem cells that can give rise to multiple specialized cell types (Pittenger et al. 1999). Along with their extensive distribution in many adult tissues, MSCs are required for continuous tissue homeostasis maintenance within diverse organs, which have made them an attractive target for tissue engineering (Prockop 1997). MSCs derived from orofacial sources, such as periodontal ligament stem cells (PDLSCs), have superior capability for orofacial regeneration as they may be more committed to differentiating into craniofacial tissues (Moshaverinia et al. 2014). However, the craniofacial tissue regeneration often results in an unfavorable outcome due to the altered local microenvironment and rapid apoptosis of transplanted MSCs (Liu et al. 2011). Therefore, there is an urgent need to discover novel therapeutic avenues for activating endogenous stem cell-based tissue regeneration.

Autotherapies are a novel treatment strategy to induce the body's innate ability to heal and protect itself, which propose a minimally invasive approach to elevate tissue healing and regeneration. To achieve the endogenous tissue self-healing, host local microenvironment as a stem cell niche provides a unique tissue structure to activate somatic stem cells for tissue regeneration (Lumelsky et al. 2018; Ruddy and Morshead 2018). MSCs, as adult stem cells, can maintain tissue homeostasis and regeneration and interplay with immune cells for immunomodulation (Akiyama et al. 2012; Chen et al. 2017).

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A supplemental appendix to this article is available online.

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However, it is still largely unknown how and whether activation of endogenous MSCs can promote tissue regeneration.

Resveratrol is a natural phytoalexin that exhibits potentials to promote tissue regeneration in various tissues and organs (Baur et al. 2006; Tseng et al. 2011). At the cellular level, resveratrol improves MSC-based therapy for liver and cardiac regeneration through activation of stem cell function and improving the survival of transplanted MSCs (Pinarli et al. 2013; Okay et al. 2015). However, the role of resveratrol in autotherapy-based tissue regeneration is largely unknown. Specifically, we aimed to identify whether resveratrol can activate somatic stem cells and promote MSC immunomodulation for endogenous tissue regeneration. In this study, we showed that resveratrol elevates cell proliferation, increases multipotent differentiation abilities, and improves PDLSC immunomodulation. Administration of resveratrol in vivo significantly activates endogenous stem cells, inhibits inflammatory cell infiltration, and rescues periodontitis phenotypes in a mouse model. Collectively, our data provide a novel strategy using a pharmacological approach for autotherapy-based tissue regeneration.

Materials and Methods

Refer to the Appendix for detailed information on methods.

Animals

Twelve-week-old female C57BL/6J and severe combined immunodeficient (SCID) mice were purchased from the Jackson Laboratory. Aged-matched female mice were used as controls in the present study. All animal experiments were performed under institutionally approved protocols for the use of animal research (University of Pennsylvania Institutional Animal Care and Use Committee #806682). This study followed the Animal Research: Reporting of In Vivo Experiments (ARRIVE 2.0) guidelines.

Antibodies and Reagents

All antibodies and reagents used in this study are described in the Appendix.

Results

Resveratrol Enhances Stemness of PDLSCs In Vitro

To gain insight into the biological function of resveratrol in PDLSCs, we first performed RNA sequencing (RNA-seq) analysis to compare transcriptomic profiles with or without resveratrol treatment. We found 724 transcripts that significantly change their expression, \log_2 fold change (FC) >1 and FC <1 and P < 0.01, after resveratrol treatment in PDLSCs compared to vehicle-treated cells (Fig. 1A, B). Among these, 323 (44.6%) were downregulated and 401 (55.4%) upregulated upon resveratrol treatment (Fig. 1C). Enrichment analysis of Gene

Oncology (GO) terms over the 724 resveratrol targets showed that the most enriched were associated with olfactory-related G-protein coupled receptor (GPCR) signaling categories (Fig. 1D). We next asked whether resveratrol could stimulate stemness and stem cell properties of PDLSCs. We identified 39 MSC stemness-related genes significantly change their expression after resveratrol treatment. Among them, 8 (20.5%) were downregulated and 31 (79.5%) upregulated (Fig. 1E), indicating resveratrol highly activated the stem cell function of PDLSCs. These results were further confirmed by quantitative polymerase chain reaction (qPCR) to show resveratrol significantly elevated 1) MSC markers, including CD44, intercellular adhesion molecule 1 (ICAM1), and vascular cell adhesion molecule 1 (VCAM1); 2) osteoprogenitor markers, including osterix (SP7), secreted phosphoprotein 1 (SPP1), and GLI family zinc finger 1 (GLI1); and 3) chondroprogenitor markers, including SRY-box transcription factor 9 (SOX9), collagen type X alpha 1 (COL10A1), and activated leukocyte cell adhesion molecule (ALCAM) (Fig. 1F).

Resveratrol Promotes PDLSC Proliferation and Differentiation Capacities

Next, we examined whether resveratrol could increase PDLSC proliferation and differentiation capacities. We then performed GO analysis to focus on cell proliferation pathways and found resveratrol highly activated cell cycle categories, particularly cell cycle, DNA replication, chromosome segregation, and DNA repair/cell cycle checkpoint (Fig. 2A). These results were further confirmed by gene set enrichment analysis (GSEA) to determine the prior gene set is significantly different between 2 biological states. Our results showed that cell proliferation related gene sets, such as DNA replication, RNA polymerase, and biosynthesis of amino acids, were highly enriched after resveratrol treatment in PDLSCs (Fig. 2B). We next performed immunofluorescence (IF) staining using cell proliferation marker Ki67 to show that resveratrol greatly enhanced Ki67+ PDLSC percentage compared to vehicle control (Fig. 2C). In addition, we measured PDLSC metabolic activity by the MTT assay to show resveratrol largely activated PDLSC viability (Fig. 2D). To further explore the molecular regulation in cell cycle after resveratrol treatment, we performed qPCR to determine that cell cycle suppressor genes p21 and p53, but not p16, were significantly inhibited by resveratrol treatment (Fig. 2E), suggesting resveratrol elevated PDLSC proliferation through inhibition of cell cycle arrest/apoptosis.

As RNA-seq showed resveratrol can promote osteoprogenitor and chondroprogenitor markers (Fig. 1F), we then examined the capacities of osteogenesis and chondrogenesis in PDLSCs with resveratrol treatment. Under osteogenic inductions, resveratrol-treated PDLSCs showed superior osteogenesis, as indicated by increased mineralized nodule formation and expression of the osteogenic genes runt-related transcription factor 2 (RUNX2) and alkaline phosphatase (ALP), respectively (Appendix Fig. 1A, B). We next showed that resveratrol-treated PDLSCs generated more new bone than vehicle-treated

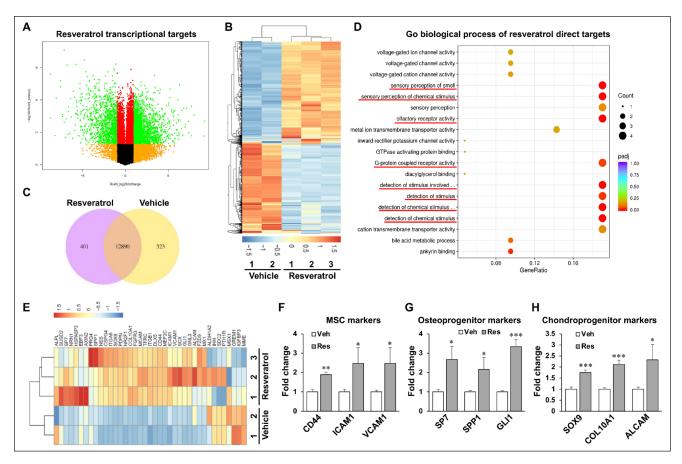


Figure 1. Transcriptome analysis revealed resveratrol promoted periodontal ligament stem cell (PDLSC) stemness. (A) Volcano plot with the log₂ fold changes in gene expression after resveratrol treatment on the *x*-axis and the statistical significance (*P* value) on the *y*-axis. (B) Heatmap of RNA sequencing (RNA-seq) expression data showing the genes that were differentially regulated following treatment with 5 μM resveratrol. Gene expression is shown in normalized log₂ counts per million. Differentially expressed genes were selected based on a 4-fold change. (C) Venn diagram of differentially expressed genes in resveratrol-treated PDLSCs with *P*<0.05. (D) Gene Ontology (GO) enrichment analysis for differentially regulated genes between control and resveratrol-treated group. Only top 19 false discovery rate (FDR) enrichments of GO terms from "biological process" category were listed. (E) Heatmap of RNA-seq expression data showing the PDLSC stemness genes that were differentially regulated following resveratrol treatment. (F–H) Quantitative polymerase chain reaction assay showed the significant increased levels of mesenchymal stem cell (MSC) markers, osteoprogenitor markers, and chondroprogenitor markers in resveratrol-treated PDLSCs. Error bars represent the standard deviation from the mean values. *P<0.05.***P<0.01.****P<0.005.

control at 8 wk postimplantation using an established in vivo MSC implantation assay, in which 4×10^6 PDLSCs with hydroxyapatite tricalcium phosphate (HA/TCP) particles as a carrier were subcutaneously implanted into immunocompromised mice (Appendix Fig. 1C). Parallel studies showed an increased capacity of resveratrol-treated PDLSCs to differentiate into chondrocytes under chondrogenic inductive conditions, associated with the elevated expression of aggrecan (ACAN) and SOX9 by IF staining (Appendix Fig. 1D). To further confirm the effect of resveratrol in chondrogenesis in vivo, we showed resveratrol-treated PDLSCs generated more cartilage-like structures with increased COL2+ chondrocytes than a control group at 8 wk postimplantation using an in vivo chondrogenic implantation assay, in which 4×10^6 PDLSCs with gelfoam-hydrogel as a carrier were subcutaneously implanted into immunocompromised mice (Appendix Fig. 1E). Taken together, these findings indicated that resveratrol promotes PDLSC stem cell properties.

Resveratrol Regulates ERK-WNT Crosstalk through Activation of Olfactory and Growth Factor Pathways

We next aimed to identify the molecular signaling targets of resveratrol in PDLSCs. To confirm our findings in Figure 1D, we performed GSEA analysis to show olfactory-related GPCR gene sets, such as olfactory pathways, olfactory receptor (OR) activity, and sensory receptor of smell, were highly enriched after resveratrol treatment (Fig. 3A). In addition to olfactory signaling, we also identified several growth factors, linked to platelet-derived growth factor (PDGF), vascular endothelial growth factor (VEGF), and epidermal growth factor (EGF) pathways, were highly activated in resveratrol-treated PDLSCs (Fig. 3B). We then examined the expression levels of ORs and growth factors by qPCR to show that ORs including OR52N4, OR10A3, OR2A7, OR2AG2, and OR2A42 and growth factors including PDGFD, EGF, PDGFRL, VEGFA, VEGFC, and PDGFC were highly activated by resveratrol treatment in

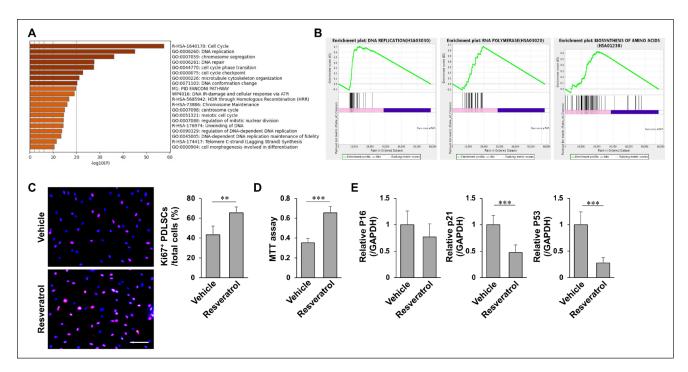


Figure 2. Resveratrol elevated periodontal ligament stem cell (PDLSC) proliferation. (A) Gene Ontology (GO) enrichment analysis showed resveratrol treatment significantly activated cell cycle and cell proliferation pathways in PDLSCs. (B) Next-generation RNA sequencing (RNA-seq) data of resveratrol-treated PDLSCs were interrogated by gene set enrichment analysis to identify enriched biologic pathways. Cell proliferation pathways, including DNA replication, RNA polymerase, and biosynthesis of amino acids, were significantly activated after resveratrol treatment in PDLSCs. (C) Ki67 immunofluorescence staining showed resveratrol treatment elevated Ki67⁺ cells compared to the vehicle-treated group. Scale bar: 25 μm. (D) Assessment of cell viability by the MTT assay showed that resveratrol promoted PDLSC viability. (E) Quantitative polymerase chain reaction assay showed the levels of cell cycle genes with or without resveratrol treatment in PDLSCs. Error bars represent the standard deviation from the mean values. **P<0.01. ****P<0.005.

PDLSCs (Fig. 3C). As the downstream target of both olfactory-GPCR (Mykytyn and Askwith 2017) and growth factor pathways (Bruggemann et al. 2021) is ERK mitogen-activated protein kinase signaling, we then showed that resveratrol significantly activated ERK signaling through phosphorylation of ERK (p-ERK) in PDLSCs (Fig. 3D). The crosstalk between ERK and WNT signaling has been shown to promote MSC lineage commitment toward osteogenesis through stimulation of RUNX2 activity (Cervenka et al. 2011). Our data further revealed that resveratrol was able to enhance the level of active β-catenin in PDLSCs (Fig. 3D). To examine whether WNT/βcatenin signaling acts as a downstream target of the ERK pathway, we used small interfering RNA (siRNA) to knock down ERK in resveratrol-treated PDLSCs. Western blot indicated that siERK treatment downregulated both p-ERK and active β-catenin levels, suggesting ERK signaling controlled WNT/βcatenin cascades in PDLSCs (Fig. 3D). To further explore the functional role of ERK/WNT crosstalk in PDLSCs, siERK treatment greatly blocked osteogenic capacity induced by resveratrol, as indicated by decreased mineralized nodule formation and reduced levels of RUNX2 and ALP (Fig. 3E, F). Collectively, these findings reveal that resveratrol-mediated olfactory and growth factor pathways activate downstream ERK/WNT crosstalk to promote PDLSC stemness.

Resveratrol Is Associated with PDLSC-Mediated Immunomodulation

Since immunomodulatory properties were recently identified as an important characteristic of PDLSCs, which has led to their systemic infusion to treat a variety of immune diseases, we next examined whether resveratrol regulates PDLSC-mediated immunomodulation. We first used a PDLSC/T-cell coculture system to show resveratrol-treated PDLSCs had significantly increased capacity to induce Annexin-V⁺ apoptotic T cells when compared to the vehicle-treated PDLSCs (Fig. 4A). In order to assess the therapeutic mechanism of resveratrol-treated PDLSCs, we used an inductive experimental colitis model (Alex et al. 2009) to evaluate the therapeutic effect of allogenic PDLSC transplantation (PDLSC-T) at day 3 after dextran sulfate sodium (DSS) induction. Treatment with 0.2×10^6 PDLSCs (positive control group) is considered a standard dosage to elicit a therapeutic response (Chen et al. 2014). Therefore, we infused 10% (0.1×) of that amount (0.02×10⁶ of resveratrol-treated PDLSCs) into colitis mice to examine whether resveratrol pretreatment could reduce the dosage of PDLSCs in immunotherapy (Fig. 4B). The body weight of mice with induced colitis was significantly reduced compared to control C57BL6 mice from day 8 to 12 post-DSS induction. After normal,

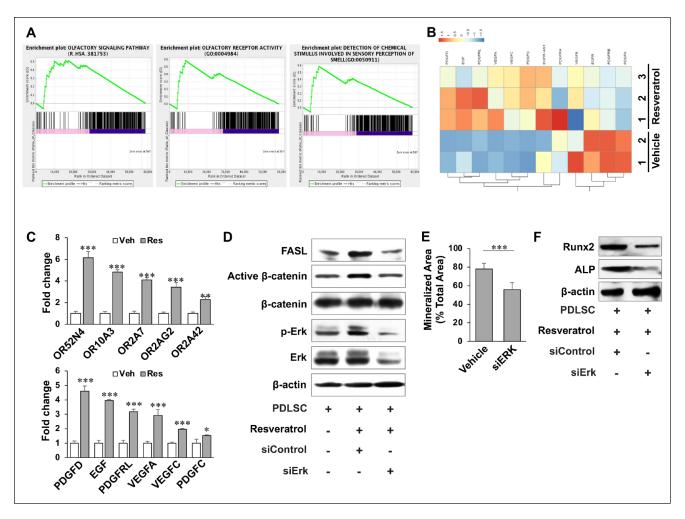


Figure 3. Resveratrol-activated ERK/WNT crosstalk through olfactory and growth factor pathways. (A) Gene set enrichment analysis assay showed resveratrol treatment elevated olfactory receptor signaling in periodontal ligament stem cells (PDLSCs). (B) Heatmap of RNA sequencing (RNA-seq) expression data showing the growth factor genes that were differentially regulated following resveratrol treatment in PDLSCs. (C) Quantitative polymerase chain reaction assay further confirmed that the expression levels of olfactory receptors (ORs) and growth factors were significantly increased after resveratrol treatment in PDLSCs. (D) Western blotting analysis showed the expression levels of FASL, active β-catenin, β-catenin, p-ERK, and ERK in PDLSCs with or without resveratrol treatment. ERK small interfering RNA (siRNA) transfection was also performed in resveratrol-treatmed PDLSCs to knock down ERK expression level. (E) Alizarin red staining showed the capacity to form mineralized nodules under osteoinductive conditions in siControl- and siERK-transfected PDLSCs with resveratrol treatment. (F) Western blotting analysis showed the expression levels of the osteogenic genes RUNX2 and ALP under osteoinductive conditions in siControl- and siERK-transfected PDLSCs with resveratrol treatment. β-Actin was used as a protein loading control. Error bars represent the standard deviation from the mean values. *P<0.05. **P<0.01. ***P<0.005.

resveratrol-treated or 0.1× resveratrol-treated PDLSC-T, the body weight was partially restored (Fig. 4C). The disease activity index (DAI), including body weight loss, diarrhea, and bleeding, was significantly elevated in the colitis mice. After normal, resveratrol-treated or 0.1× resveratrol-treated PDLSC-T, the DAI score was decreased (Fig. 4D). Furthermore, colon tissue from each group was analyzed. Both the absence of the epithelial layer and inflammatory cell infiltration were observed in the colon of induced colitis compared to the control group. Normal, resveratrol-treated or 0.1× resveratrol-treated PDLSC-T recovered epithelial structure and eliminated inflammatory cells in colitis mice (Fig. 4E). Histological activity index (Alex et al. 2009) confirmed that normal, resveratrol-treated or 0.1× resveratrol-treated PDLSC-T reduced the DAI

(Fig. 4G). In addition, IF staining showed significantly increased CD4⁺IL17⁺ T-helper 17 (Th17) infiltration in the colon of induced colitis. Transplantation of normal, resveratrol-treated or 0.1× resveratrol-treated PDLSCs dramatically reduced the infiltrated Th17 in colitis mice (Fig. 4F, H). Flow cytometry analysis further showed that elevated Th17 cells were observed in the colitis mice. Normal, resveratrol-treated or 0.1× resveratrol-treated PDLSC-T significantly downregulated Th17 (Fig. 4I). The data therefore suggest that the number of PDLSCs used for immunotherapy could be dramatically reduced to induce T-cell apoptosis and offer a potential treatment for colitis mice. FAS ligand (FASL) is a type II transmembrane protein that binds with FAS to form the death-inducing signaling complex in many cell types (Zhang et al. 2008). To

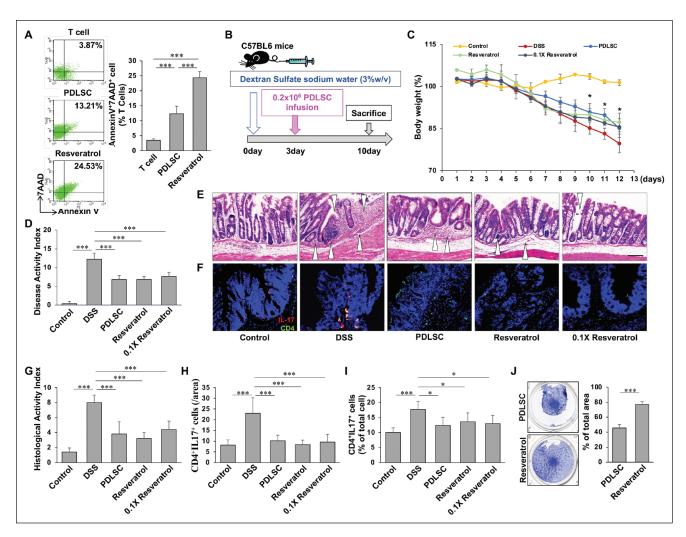


Figure 4. Resveratrol improved periodontal ligament stem cell (PDLSC) immunomodulation in vitro and in vivo in a DSS-induced experimental colitis mouse model. (A) PDLSCs induced annexinV+7AAD+ double-positive apoptotic CD3+ T cells in a PDLSC/T-cell direct coculture system in vitro. Resveratrol treatment further increased annexinV+7AAD+ double-positive apoptotic CD3+ T cells. (B) Schema showing PDLSC transplantation in dextran sulfate sodium (DSS)-induced experimental colitis mice. (C) Colitis mice showed significantly reduced body weight from 8 to 12d after DSS induction. The PDLSC transplantation, resveratrol-treated PDLSC transplantation, and 0.1× resveratrol-treated PDLSC transplantation groups showed inhibition of body weight loss compared to the colitis group at 12d after DSS induction (n=6 per group). (D) Disease activity index (DAI) was significantly increased in colitis mice compared to C57BL6 mice at 12d after DSS induction. PDLSC transplantation, resveratrol-treated PDLSC transplantation, and 0.1× resveratrol-treated PDLSC transplantation significantly reduced DAI score. (E, G) Hematoxylin and eosin staining showed the infiltration of inflammatory cells (white arrows) in colon with destruction of epithelial layer in colitis mice. PDLSC transplantation, resveratroltreated PDLSC transplantation, and 0.1× resveratrol-treated PDLSC transplantation rescued disease phenotype in colon and reduced histological activity index (G). Scale bar: 25 µm. (F, H) Immunofluorescence staining showed CD4⁺IL17⁺ Th17 cell infiltration in colitis. PDLSC transplantation, resveratrol-treated PDLSC transplantation, and 0.1× resveratrol-treated PDLSC transplantation dramatically reduced Th17 infiltration in colon. Scale bar: 25 µm. (I) Flow cytometry analysis further confirmed that Th17 cell level was significantly elevated in colitis mice compared to C57BL6 mice after DSS induction. PDLSC transplantation, resveratrol-treated PDLSC transplantation, and 0.1× resveratrol-treated PDLSC transplantation reduced the levels of Th17 cells in colitis mice. (J) Activated T cells were capable of inducing significant PDLSC death in a PDLSC/T-cell direct coculture system. Resveratrol treatment protected PDLSC apoptosis. Error bars represent the standard deviation from the mean values. *P<0.05. ***P<0.005.

examine whether resveratrol induces PDLSC immunomodulation via ERK-mediated FASL activation, Western blot showed resveratrol treatment significantly elevated the level of FASL in PDLSCs (Fig. 3D). Knockdown of *ERK* by siRNA dramatically reduced the level of FASL, indicating FASL is required for resveratrol-induced immunotherapy in PDLSCs through activation of ERK signaling (Fig. 3D). As immune components can also target MSCs through death pathways (Liu et al. 2011), we

next examined whether resveratrol treatment is able to protect PDLSC survival after cocultured with activated T cells in a PDLSC/T-cell coculture system. Toluidine blue staining showed activated T cells caused PDLSC, but not resveratrol-treated PDLSC, death (Fig. 4J). Collectively, our results revealed that resveratrol can promote PDLSC immunomodulation through activation of FASL-mediated T-cell apoptosis, as well as protection of PDLSC survival.

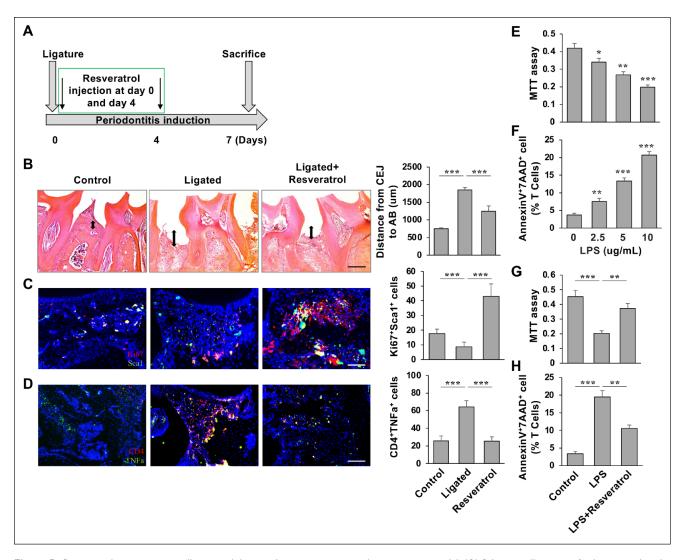


Figure 5. Resveratrol treatment partially rescued disease phenotypes in a periodontitis mouse model. (A) Schematic illustration for ligature-induced periodontitis model and local administration of resveratrol. A 5-0 silk ligature was tied around the maxillary second molar in C57BL/6 mice on day 0, and either placebo (PBS) or resveratrol was simultaneously injected as illustrated. (B) Periodontal bone resorption analysis. The distance from the cementoenamel junction (CEJ) to the pinnacle of the alveolar bone (AB) was determined to assess periodontal bone loss. (C) Immunofluorescence (IF) staining revealed Ki67+Sca1+ cells were reduced in periodontitis mice. Resveratrol treatment significantly elevated Ki67+Sca1+ cells. (D) IF staining showed increased CD4+TNF α + inflammatory T-cell infiltration in periodontitis mice. After resveratrol administration, CD4+TNF α + inflammatory T-cell infiltration was largely reduced. Scale bar: 25 μ m. (E, F) Cell viability and apoptosis were measured in periodontal ligament stem cells (PDLSCs) after treatment with various concentrations of lipopolysaccharide (LPS) for 12 h by using the MTT assay and flow cytometry analysis. Cell viability (G) and apoptosis (H) were measured in PDLSCs treated with resveratrol during 10 μ g/mL LPS stimulation. Error bars represent the standard deviation from the mean values. *P<0.01. ****P<0.01. ****P<0.005.

Resveratrol Ameliorates Periodontitis Phenotypes through Activating Stemness and Immunomodulation of Endogenous Stem Cells

Based on the recent advances in stem cell biology, immunology, and material sciences, autotherapies, a novel concept for tissue regenerative medicine, have been proposed to optimize endogenous tissue responses and microenvironment for somatic stem cell activation (Lumelsky et al. 2018; Yui et al. 2018). As we find that resveratrol can promote stemness and

immunomodulation of PDLSCs, we then asked whether resveratrol could activate endogenous stem cells for tissue regeneration through regulation of inflammatory microenvironment. To this end, we employed a ligature-induced periodontitis model, by which severe alveolar bone loss with an activated proinflammatory microenvironment was established (Marchesan et al. 2018). Local injection of resveratrol was performed at day 0 and day 4 after ligation (Fig. 5A). At day 7 postligation, severe alveolar bone loss was observed around the ligated second molar compared to the control group. In contrast, distance

from cementoenamel junction (CEJ) to alveolar bone (AB) crest in the resveratrol-treated group decreased to approximately half of the ligated mice with a clear statistical trend (Fig. 5B). These data prompted us to further examine whether resveratrol treatment activated endogenous stem cells and inhibited proinflammatory T cells in periodontitis mice. IF staining showed the number of costained Ki67 and the MSC marker SCA-1 was dramatically decreased in the ligated mice, while resveratrol treatment significantly elevated the number of Ki67+SCA-1+ cells in vivo (Fig. 5C). In addition, our results showed resveratrol treatment decreased the number of CD4⁺TNFα⁺ proinflammatory T cells when compared to ligated mice, in which CD4+TNF α + T cells were highly expanded after ligation (Fig. 5D). To investigate whether resveratrol treatment can rescue periodontal pathogenic bacteria such as Porphyromonas gingivalis (PS)-induced endogenous stem cell repression, we established an in vitro periodontitis model using PG-lipopolysaccharide (LPS)-treated PDLSCs to show that treatment with LPS significantly inhibited viability and increased cell apoptosis in a dose-dependent manner (Fig. 5E, F). Resveratrol treatment partially elevated PDLSC viability and decreased cell apoptosis under 10 µg/mL LPS stimulation (Fig. 5G, H). Overall, resveratrol treatment promotes endogenous stem cell activation and inhibits proinflammatory microenvironment for autotherapies in the periodontitis mice.

Discussion

MSC-based regenerative medicine is a promising approach for tissue reconstruction and disease management, by which MSCs can regulate tissue repair/homeostasis and interact with immune cells for immunomodulation (Prockop 1997; Akiyama et al. 2012; Chen et al. 2017). The increased research indicated that natural products could regulate the immune response with few adverse side effects, which offer new avenues for immunomodulation and can be promising agents in preventing chronic diseases (Kishore et al. 2019). Resveratrol is a promising pharmacological target in regulation of cell viability, proliferation, anti-inflammation, and osteogenesis of MSCs (Wang et al. 2014; Li et al. 2019). In this study, we reveal that resveratrol treatment significantly improves stemness of PDLSCs through activation of stem cell markers, elevation of self-renewal and multipotent differentiation abilities, and upregulation of immunomodulatory capabilities. Furthermore, activation of endogenous stem cells by a pharmacological approach, such as in vivo resveratrol treatment, can markedly improve tissue regeneration and rescue disease phenotypes in a periodontitis mouse model. This study provides experimental evidence that links resveratrol to PDLSC-mediated tissue regeneration and demonstrates the potential to improve autotherapies through activation of endogenous stem cells.

Profiling the transcriptional level of stem cells at a defined condition using RNA-seq is a promising analysis to identify and prioritize genetic variants in the altered expression levels (Schlieben et al. 2021). By using RNA-seq analysis, olfactory and growth factor pathways are identified as downstream targets of resveratrol in PDLSCs. In a recent study, olfactory receptors have been linked to bone metabolism in an osteoporotic condition, suggesting that ORs are expressed in MSC-osteoblast lineage cells and required to maintain skeletal homeostasis (Zhu et al. 2018). Our data reveal that several ORs are expressed in PDLSCs, and resveratrol treatment dramatically increases osteogenic capacity through elevating more than 20 ORs, implying ORs may play a critical role in PDLSC-mediated bone regeneration. Growth factors are well known to be key mediators in supporting MSC survival, proliferation, and differentiation, which are the drivers of regenerative medicine (Nie et al. 2020). Resveratrol significantly activates the expression levels of growth factors, further indicating growth factor and OR pathways may synergistically promote PDLSC stemness.

ERK signaling, the downstream target of both OR-GPCR and growth factor signaling, is required for bone formation during skeletal development and plays an important role in maintaining bone tissue homeostasis (Liu, Zhao et al. 2018). Several natural compounds act as activators of ERK signaling, which is the key driver in osteogenesis for defeating skeletal disorders (Liu, Zhao et al. 2018; Liu et al. 2019). WNT/βcatenin signaling has been shown as a downstream kinase of ERK and is also essential for the self-renewal and multipotent differentiation of MSCs and regulating bone tissue homeostasis (Cervenka et al. 2011; Lin et al. 2019). Our mechanism studies determine that ERK/WNT crosstalk mediates resveratrol-induced PDLSC activation, by which ERK activates downstream β-catenin through OR-GPCR and growth factor signaling. Since resveratrol has been shown to be involved in activation of the WNT/β-catenin cascades to promote osteogenic differentiation (Zhao et al. 2018), collectively, the present study explores the novel targets of resveratrol to activate ERK/WNT crosstalk in PDLSC-mediated tissue regeneration.

MSCs exhibit immunomodulatory properties by mediating the proliferation, migration, and differentiation of several major types of immune cells, and systemic infusion of MSCs or MSC-derived cellular components has been shown to yield therapeutic benefits for a variety of immune-related disorders (Le Blanc et al. 2004; Chen et al. 2015; Liu, Kou et al. 2018). Our previous studies showed that MSCs induce T-cell apoptosis via upregulation of the FASL-mediated FAS death pathway to achieve immune tolerance, by which β-catenin directly binds to the Fasl promoter to drive gene expression at the transcriptional level (Akiyama et al. 2012; Chen et al. 2014). Here, we showed that resveratrol improves PDLSC immunomodulation through FASL activation, which is regulated by ERK/ WNT crosstalk. Taken together, our data reveal an ERK/WNT/ FASL cascade in the regulation of PDLSC immunomodulation and explore the potential to improve PDLSC-based clinical therapies with reduced cell dosage.

The new paradigm of tissue regeneration shifts to manipulate/activate an endogenous tissue microenvironment to minimize invasive approaches for regenerative medicine, which is termed *autotherapies*. To achieve autotherapies, preconditioning

a regenerative microenvironment, creating a specific stem cell niche, and activating transcription factors for lineage reprogramming are the key components for enhancing endogenous tissue regeneration (Heber-Katz 2017; Lumelsky et al. 2018). Periodontitis is an inflammatory disease, characterized by inflammatory response and alveolar bone loss (Hajishengallis 2015). MSC-based therapy has been widely studied for the treatment of periodontal disease because of its effects on bone regeneration and immunomodulation (Shang et al. 2017). By using an in vivo periodontitis model, we showed resveratrol partially rescues disease phenotypes through activation of endogenous stem cells and inhibition of immune cell infiltration. In summary, this translational study substantially extends current knowledge about stem cell-based autotherapies. We also reveal an ERK/WNT crosstalk mechanism to boost PDLSC stemness and immunomodulation for tissue regeneration.

Author Contributions

W. Li, contributed to design, data acquisition, analysis and interpretation, drafted and critically revised the manuscript; X. Huang, contributed to design, data acquisition, analysis, and interpretation, drafted manuscript; W. Yu, contributed to design, data acquisition, and analysis, critically revised the manuscript; Y. Xu, R. Huang, J. Park, contributed to data acquisition and analysis, critically revised the manuscript; A. Moshaverinia, contributed to data analysis and interpretation, critically revised the manuscript; P. Arora, contributed to conception, data analysis, and interpretation, critically revised the manuscript; C. Chen, contributed to conception, design, data acquisition, analysis, and interpretation, drafted and critically revised the manuscript. All authors gave final approval and agree to be accountable for all aspects of the work.

Declaration of Conflicting Interests

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