

UCLA

UCLA Previously Published Works

Title

Steroid Hormone Vitamin D

Permalink

<https://escholarship.org/uc/item/3hz7z99m>

Journal

Circulation Research, 122(11)

ISSN

0009-7330

Authors

Demer, Linda L

Hsu, Jeffrey J

Tintut, Yin

Publication Date

2018-05-25

DOI

10.1161/circresaha.118.311585

Peer reviewed



Published in final edited form as:

Circ Res. 2018 May 25; 122(11): 1576–1585. doi:10.1161/CIRCRESAHA.118.311585.

Steroid hormone vitamin D: Implications for cardiovascular disease

Linda L. Demer^{1,2,3}, Jeffrey J. Hsu¹, and Yin Tintut^{1,2,4}

¹Departments of Medicine, University of California, Los Angeles, CA 90095-1679

²Physiology, University of California, Los Angeles, CA 90095-1679

³Bioengineering, University of California, Los Angeles, CA 90095-1679

⁴Orthopaedic Surgery, University of California, Los Angeles, CA 90095-1679

Abstract

Understanding of vitamin D physiology is important because about half of the population is being diagnosed with deficiency and treated with supplements. Clinical guidelines were developed based on observational studies showing an association between low serum levels and increased cardiovascular risk. However, new randomized-controlled trials have failed to confirm any cardiovascular benefit from supplementation in the general population. A major concern is that excess vitamin D is known to cause calcific vasculopathy and valvulopathy in animal models. For decades, administration of vitamin D has been used in rodents as a reliable experimental model of vascular calcification. Technically, vitamin D is a misnomer. It is not a true vitamin because it can be synthesized endogenously through ultraviolet exposure of the skin. It is a steroid hormone that comes in three forms that are sequential metabolites produced by hydroxylases. As a fat-soluble hormone, the vitamin D-hormone metabolites must have special mechanisms for delivery in the aqueous blood stream. Importantly, endogenously synthesized forms are carried by a binding protein, whereas dietary forms are carried within lipoprotein particles. This may result in distinct bio-distributions for sunlight-derived vs. supplement-derived vitamin D-hormones. Since the cardiovascular effects of vitamin D-hormones are not straightforward, both toxic and beneficial effects may result from current recommendations.

Keywords

Cholecalciferol; vitamin D; calcitriol; cardiovascular; calcification

On November 30, 2010, at the request of the Canadian and U.S. Governments, the Institute of Medicine provided a report addressing conflicting information on vitamin D.¹ This report took into consideration more than 1,000 studies and reports and considered testimony from scientists and “stakeholders.” Outcomes included bone and cardiovascular diseases as well

Corresponding author: Linda L. Demer, MD, PhD, Division of Cardiology, Center for Health Sciences A2-237, David Geffen School of Medicine at UCLA, Los Angeles, CA 90095-1679, Phone (310) 206-2677, LDemer@mednet.ucla.edu.

DISCLOSURES:

None.

as cancer, diabetes, inflammation, neuropsychological function, physical performance, pre-eclampsia and reproduction. The overall conclusion of this report was that “the majority of Americans and Canadians are receiving adequate amounts of both calcium and vitamin D,” and that “too much of these nutrients may be harmful.” It further noted that “information about health benefits beyond those for bone -- benefits often reported in the media -- were from studies with mixed and inconclusive results that could not be considered reliable.”² Even before this report, Towler had noted that the effects of vitamin D on cardiovascular health are complex and biphasic, with direct and indirect actions mediating its vasculotropic actions.³ There is not yet evidence from a randomized controlled trial showing cardiovascular benefit of vitamin D supplementation.⁴

Until recently, hormonal regulation of calcium-phosphate metabolism by vitamin D metabolites and the parathyroid gland were of little interest to cardiovascular scientists and clinicians. But with new clinical guidelines and media attention, awareness of vitamin D physiology is necessary, especially given that, despite the conclusions of the Institute of Medicine, routine vitamin D testing and supplementation are widely recommended by physicians. As commonly occurs with supplements, it is often used in doses far beyond those directed. Given its extensive actions in human metabolism - both beneficial and harmful - the biochemistry, physiology, and financial motivations surrounding vitamin D warrant attention.

Forms of vitamin D

From a technical standpoint, the term “vitamin D” is a misnomer. It is not a true vitamin because the human body has the capacity to synthesize its own cholecalciferol (D_3), except in rare instances of complete lack of ultraviolet radiation. It is more accurate to view it as a steroid hormone or an oxysterol. The International Union of Pure and Applied Chemistry’s Commission on the Nomenclature of Biological Chemistry defines vitamin D_3 as a steroid or secosteroid. Its chemical name is 9,10-secocholesta-5,7,10(19)-trien-3 β -ol. Six different steroid hormones go by the name “vitamin D,” with varying degrees of activity: the endogenous precursor, cholecalciferol (D_3), which is derived from cholesterol; its hydroxylated derivative, calcidiol [$25(OH)D_3$], which has partial activity; and its hydroxylated derivative, the “active” dihydroxy form, calcitriol [$1,25(OH)_2D_3$]. In addition, there is a plant-derived form, ergocalciferol (D_2), which also has the corresponding monohydroxy and dihydroxy metabolites. Since $25(OH)D_3$ is longer lasting, it is the level of this hormone -- not that of the more active $1,25(OH)_2D_3$ -- that is used to diagnose clinical deficiency. Notably, levels of $25(OH)D_3$ tend to vary inversely with the levels of the active form, possibly due to displacement of the active metabolite from D-binding protein (DBP).⁵ In this article, we will use the term “vitamin D-hormones” for all six types of steroid hormones and to emphasize their true physiological nature.

Sources of vitamin D-hormones and biodistribution

Sources of vitamin D-hormones include exposure to ultraviolet light, certain foods, and dietary supplements. As one of the four fat-soluble vitamins (A, D, E, and K), its lipophilicity requires special mechanisms to pass through the aqueous environment of blood

to reach tissues and cells. Separate mechanisms are used for the endogenous D₃ synthesized in sun-exposed skin vs. exogenous D₃ obtained from diet or supplements. This may result in distinct pharmacokinetic volumes and targets of distribution (Figure 1).

Endogenous vitamin D-hormone synthesis, transport and activation

Endogenous vitamin D-hormone synthesis occurs by ultraviolet light exposure of 7-dehydrocholesterol within the microvessels of the skin resulting in its conversion into cholecalciferol (D₃). But, as a fat-soluble oxysterol, D₃ must be carried in the blood by DBP, a liver-derived apoprotein and a member of the albumin gene family.⁶ For light-skinned individuals, sun exposure of the face and arms for just 15 minutes per week may produce tens of thousands of units of cholecalciferol. This endogenous production from sun exposure had been the major source for most humans for centuries. The fact that sun-derived D₃ is carried on DBP is a key difference from exogenous D₃, because of its potential influence on bio-distribution.

Exogenous vitamin D-hormone sources and delivery

Exogenous sources of vitamin D-hormones include diet (eggs, fish, liver, and marine mammal fat) and supplements. A cup of milk provides about 100 international units (IU) and a serving of salmon contains about 400 IU of D₃. While the dietary sources may be in the D₃ or D₂ form, supplements typically derive from the plant-derived hormone, ergocalciferol (D₂). A key feature of dietary or supplemental sources is that D₃ taken orally is absorbed from the intestinal tract via chylomicrons,⁷ which pass into the lymphatic circulation before returning to the central venous circulation via the thoracic duct. Eventually, about 35% of ingested D₃ is carried in lipoproteins,⁸ rather than DBP.

Activation by sequential hydroxylation of D₃

For both endogenous and exogenous sources, the D₃ carried in the bloodstream on either DBP or lipoproteins undergoes a two-step sequential hydroxylation to active metabolites. First, it is converted by 25-hydroxylase to the monohydroxy- derivative, 25(OH)D₃, the metabolite that is measured for “vitamin D levels.” This occurs primarily in the liver, but may take place in other tissues as well. Next, 25(OH)D₃ is further hydroxylated by 1-alpha hydroxylase to the active, dihydroxy- form, 1,25(OH)₂D₃.⁸ This occurs primarily in the capillaries surrounding the proximal convoluted tubules of kidney, but, importantly, the enzyme producing the active form is also found in vascular cells and monocytes among other tissues and cells (Table).

Guidelines for vitamin D-hormone assessment and supplementation

Different criteria for vitamin D deficiency have been proposed by the Endocrine Society, Osteoporosis Society, and Institute of Medicine. Normal reference values shown by individual clinical laboratories are not standardized. Conservative definitions define vitamin D deficiency as levels of 25(OH)D₃ < 20 ng/ml (< 50 nmol/L), and vitamin D insufficiency as 20 – 30 ng/ml (50 – 75 nmol/L).⁹ The Institute of Medicine chooses cut-off values of < 12 ng/ml and > 50 ng/ml as levels with increased risk of deficiency and excess, respectively.¹ The Institute of Medicine does not recommend specific doses, but, based on bone health

indicators, their analysis suggests that the daily use of vitamin D is 600 IU for individuals from 1 to 70 years of age, and 800 IU for individuals 71 and older, some or all of which may be achieved by ordinary sun exposure.¹ They further suggest a “safe upper limit” of dietary vitamin D intake as 4000 IU daily, a level at which risk for toxicity begins to increase. Yet, the Institute of Medicine emphasizes that this upper limit “should not be misunderstood as amounts people need or should strive to consume.”¹

The use of the terms “daily” and “per day” in these recommendations may give the false impression that a day without sunshine requires a dose of supplement. Even though adults may use a given amount of cholecalciferol each day, such daily use does not necessarily require daily replacement. 25(OH)D₃ has a half-life of 2 weeks¹⁰ to 3 months,¹¹ and is stored primarily in adipose tissue^{12, 13} and, to a lesser extent, in the liver.¹⁴ Presumably, this stored source of vitamin D is available for release back into the plasma, as indicated by a long-term study in Norwegians.¹⁵ Moreover, cholecalciferol recycles in the enterohepatic circulation.¹⁶ Thus, vitamin D-hormones may not require daily, weekly or even monthly replenishment. Summer sun exposure may provide enough for the winter.¹⁷ Major institutions have used dosing schedules as infrequent as once every 1–4 months.^{18, 19} It may be more correct to refer to a monthly requirement, and this requirement may vary depending on age (< 70 years or > 70 years) or season (i.e., summer vs. winter).¹⁷

Personalized approach to vitamin D-hormones

A personalized medicine approach is important in considering vitamin D-hormone supplementation because of the influence of differences in body composition, environmental factors, and genetic variations in D binding protein as well as variations in the intracellular vitamin D receptor. Although darkly pigmented individuals are believed to require more sun exposure to generate the same amount of vitamin D-hormones, they have genetic polymorphisms of the vitamin D binding protein,²⁰ which change the bioavailability of vitamin D, counteracting the decrease in synthesis.²¹ The half-life of 25(OH)D₃ in the bloodstream is influenced by genotype of the DBP. Based on in vitro studies, the intracellular vitamin D receptor (VDR), which binds and translocates 1,25(OH)₂D₃ to the nucleus, has approximately 7% readthrough efficiencies, producing VDR proteoforms that have reduced binding.²² This phenomenon may vary among individuals. High body fat content may decrease availability of fat-soluble 25(OH)D₃ due to sequestration in adipose tissue.²³ Conversely, high skeletal muscle content also modulates vitamin D-hormone availability; muscle cells internalize D-binding protein and expose it, allowing extensive intracellular uptake and retention of 25(OH)D₃.²⁴ The elderly may have lower levels due to less outdoor activity and sun exposure.¹⁷ Polymorphisms arising in the Inuit Eskimo background limit intestinal D₃ uptake protect against excess D₃ ingestion from the ancestral diet of whale blubber rich in vitamin D hormones. These genetic variations, phenotypic differences, and environmental influences underscore the importance of tailoring any recommendations for vitamin D supplementation to individualized needs.

Vitamin D-hormones and cardiovascular health

Evidence for cardiovascular protection by vitamin D-hormones was almost entirely inferred from observational studies. Confounders that may adversely affect results of these studies include obesity, which alters the storage of vitamin D, skeletal muscle content, physical exercise, which corresponds with time outdoors in the sun, and illness, which corresponds with time indoors.

Widespread supplementation became a guideline²⁵ based on observational studies without adequate randomized controlled trials.² Although it is widely cited as showing an inverse relation between 25(OH)D₃ levels and cardiovascular risk, a close look at data in the Offspring Cohort of the Framingham Heart Study²⁶ shows a U-shaped relationship between 25(OH)D₃ levels and cardiovascular risk (Figure 2²⁶). The apparent minimum risk occurs at a serum level of approximately 20 ng/ml, far below the level considered sufficient. A U-shaped curve was also found for the relationship between 25(OH)D₃ and all-cause mortality in the NHANES III population.²⁷ Preclinical studies support this relationship with both deficiency and excess of 25(OH)D₃ increasing atherosclerotic calcification.²⁸ Yet, public education campaigns continue to describe the relationship as inverse.

Although observational studies show associations between 25(OH)D₃ levels and cardiovascular risk, the few randomized controlled trials available have failed to confirm any cardiovascular benefit of supplementation²⁹ with the exception of patients with chronic kidney disease, where the kidney's 1 α -hydroxylase activity and capacity to produce active 1,25(OH)₂D₃ are greatly diminished. One study found no reduction in cardiovascular risk factors in patients randomized to supplementation.³⁰ Another study of over 5000 patients found no reduction in cardiovascular mortality in patients randomized to supplementation, even though the treatment increased 25(OH)D₃ levels by an average of 20 ng/ml.¹⁸ In contrast, patients with renal insufficiency and/or dialysis, where vitamin D-hormone deficiency is prevalent,³¹ vitamin D-hormone supplementation improved vascular function^{32–34} without affecting plasma levels of calcium and phosphate.^{35, 36} Overall, only the observational studies showed reductions in all-cause and cardiovascular mortality.^{37–39}

Accumulation and activation of LDL-associated vitamin D-hormones in the artery wall

Both mono- and dihydroxy- forms of vitamin D-hormones are delivered to cells either by DBP,⁴⁰ where entry is mediated by the endocytic receptors, cubilin/megalin,⁴¹ or by lipoproteins,⁸ where entry is mediated by the low-density lipoprotein (LDL) receptor.⁴² However, during pathogenesis of atherosclerosis, vitamin D-hormones that are consumed in the diet may accompany LDL into the subendothelial space of the artery wall where atherosclerotic lesions form.⁴³ In peripheral tissues that express lipoprotein lipase, the chylomicron metabolizing enzyme, a fraction of vitamin D-hormones can be taken up by the tissues. Since 1- α hydroxylase is present in tissues and cells, including vessel walls and monocyte-derived cells, the active form may be produced locally within the artery wall and, conceivably, within monocyte-laden atherosclerotic plaque.⁴⁴

Targets of vitamin D-hormones

Cellular and molecular effects of vitamin D-hormones are extensive. In addition to homodimerization, VDR heterodimerizes with the retinoid X receptor to activate transcription of a wide range of genes. As steroid hormones, they are related to estrogen, testosterone, mineralocorticoids, and glucocorticoids. Even our limited search of the literature (Table) reveals hundreds of diverse genomic and non-genomic targets of vitamin D-hormones, affecting a vast array of physiological functions. Adding further complexity, vitamin D-hormones have significant cross-talk with steroid and nuclear hormones and their receptors.⁴⁵ For instance, vitamin D₃ may affect actions of glucocorticoids.^{46,47} Conversely, steroid and xenobiotic receptors⁴⁸ as well as peroxisome proliferator-activated receptor gamma⁴⁹ inhibit VDR-mediated CYP24 (24-hydroxylase) promoter activity.

Effects of vitamin D-hormones in the vasculature

Given that diet-derived 25(OH)D₃ is carried in lipoproteins, and that lipoproteins accumulate in the subendothelial space of arteries leading to atherosclerotic lesions, it is likely that diet-derived 25(OH)D₃ also accumulates in the neointima artery wall and atherosclerotic plaque. Given that vascular smooth muscle cells and monocytes both produce 1-alpha hydroxylase, it follows that 1,25(OH)₂D₃ may also accumulate in artery walls and atherosclerosis. Potential effects of this accumulation remain to be determined. One possibility is acceleration of both atherosclerosis and cardiovascular calcification, based on studies showing that vitamin D-receptor deficiency significantly reduces calcific atherosclerosis in hyperlipidemic mice.⁵⁰ Vitamin D-hormones are known to stimulate smooth muscle cell proliferation⁵¹ and induce expression of fibroblast growth factor-23 (FGF-23), high levels of which are linked to adverse cardiovascular events.⁵² With respect to mineralization, effects of vitamin D-hormones are double-edged. Although there is convincing evidence that supplements increase bone density,⁵³ any benefit to bone may be at the cost of cardiovascular morbidity and mortality due to calcific vasculopathy and valvulopathy. Cardiovascular calcification has been shown to occur by many of the same cellular and molecular processes as bone mineralization,⁵⁴ including induction of osteogenic factors by vitamin D-hormones.⁵⁵ Indeed, high dose vitamin D supplements used for several decades as an experimental model reproducibly induces severe aortic calcification, acutely and chronically, over a wide range of conditions in a variety of species in the hands of many different investigators.^{56,57} The dramatic vascular calcification seen in patients with chronic kidney disease may be due in part to local induction of 1-alpha hydroxylase in the artery wall.⁵⁸ The extensive immunomodulatory effects of vitamin D have been reviewed elsewhere.⁵⁹

Vitamin D-hormone toxicity and benefits

Overuse of vitamin D-hormone supplements carries significant risks that have been known for decades, and these risks have traditionally been associated with those of the resulting hypercalcemia that can occur at 25(OH)D₃ plasma concentrations of > 150 ng/ml (> 375 nmol/L). Thus, the traditional clinical manifestations of vitamin D-hormone toxicity are those of hypercalcemia, which include generalized (fatigue, weakness), neurological (altered mental status, irritability, coma), gastrointestinal (nausea, vomiting, constipation), and

endocrinological (polyuria, polydipsia) symptoms. Additionally, renal injury as well as the development of kidney stones may occur. As such, studies evaluating the safety of various dosing regimens typically use measurements of serum and urinary calcium to monitor the safety of the administered doses.^{60, 61}

However, given the number of cell types and tissues that possess 25-hydroxylase, vitamin D-hormones may have effects on these systems without necessarily affecting the serum or urinary calcium levels, and all of the biological processes listed in the Table may be deranged by excess intake. A daily intake of 25(OH)D₃ up to 4,000 IU is deemed to be the upper limit of safety,⁶² as the risk of harm appears to increase above this level. Yet as discussed above, variations in vitamin D-hormone production and metabolism may depend significantly on individual genotype, phenotype, and environmental conditions; thus, a universal upper limit of safety and a universal lower limit of sufficiency for all patients may not necessarily be accurate. Additionally, excess vitamin D-hormone supplements also displace the active form from binding sites, making it more available even when not appropriate.⁵ Further, given the cross-talk with other steroid hormone receptors, vitamin D-hormones in excess may have physiological effects similar to those of glucocorticoids, estrogen, or even those of anabolic steroids.⁶³ Nonetheless, in general, it is difficult to categorize any one of the numerous effects of vitamin D as necessarily beneficial or toxic, given the dependence on location as well as physiological and pathological contexts. For instance, osteoblastogenesis may be beneficial in osteoporosis but hazardous in calcific vasculopathy and valvulopathy.

Concluding remarks

Widely used guidelines for monitoring and supplementing vitamin D₃ hormones resembles the ill-fated call, years ago, for widespread use of another steroid hormone, estradiol, for post-menopausal women based on observational studies. The impact of confounding environmental factors was not recognized. Even after the Women's Health Initiative showed increased cardiovascular risk in postmenopausal women randomized to hormone replacement therapy,¹⁹ recommendations were slow to change. Observational studies are not sufficient to recommend widespread hormonal supplementation, and the same applies to vitamin D-hormones. The ongoing randomized clinical trial, Vitamin D and Omega-3 Trial (VITAL), will be helpful in determining whether vitamin D-hormone supplementation provides any benefit in the primary prevention of cancer and cardiovascular disease.⁶⁴

Decades ago, the pioneering Johns Hopkins cardiologist, Dr. Helen Taussig, anticipated the need for a personalized approach to D₃ supplements: "As is so common, the popular belief was that 'if some is good, more is better.' The result was the overdosing with vitamin D and adding it to various foods. Then came the recognition of vitamin D intoxication... we are coming to appreciate that there exists an inborn variation in man's ability to metabolize vitamin D and that some individuals may be injured by doses of vitamin D which are safe for others."⁶⁵ For health reasons, many Americans pay extra for bread free of preservatives (such as antioxidants) and meats that are free of steroid hormones. In the next aisle of the store, they buy bottles of antioxidant preservatives and steroid hormones in pill form, labeled as nutritional supplements, including D₃ hormones. Scientists need to use their knowledge

of molecular, cellular, and integrative physiology to advocate for rational use of vitamin D-hormone supplements to prevent adverse consequences to cardiovascular health by overenthusiastic guidelines followed by well-meaning physicians.

Acknowledgements

Without express permission, the authors wish to acknowledge the late Hywel Davies MD FRCP (1924–2017), whose writing and input contributed to this review.

SOURCES OF FUNDING

This work was supported in part by funding from the National Institutes of Health (HL114709, HL121019, HL007895), the Claude D. Pepper Older American Independence Center (OAIC) at UCLA, and an award from the UCLA Specialty Training and Advanced Research (STAR) Program.

Nonstandard Abbreviations and Acronyms:

25(OH)D₃	Calcidiol
1,25(OH)₂D₃	Calcitriol
DBP	(Vitamin) D-binding protein
IU	International unit
D₂	Ergocalciferol
VDR	Vitamin D receptor
LDL	Low-density lipoprotein
VLDL	Very low-density lipoprotein CYP24 (24-hydroxylase)
FGF-23	Fibroblast growth factor-23
SMC	Smooth muscle cell
VSMC	Vascular smooth muscle cell
Hox-8	Muscle segment homeobox-containing gene Msx-2
UV	Ultraviolet
NFAT1	Nuclear factor of activated T cells 1
MAPK	Mitogen-activated protein kinase
c-myc	Cellular homolog of the oncogene of avian myelocytomatosis virus strain 29
Runx2	Runt related transcription factor-2

REFERENCES

1. Committee IoM. In: Ross AC, Taylor CL, Yaktine AL, Del Valle HB, eds. Dietary reference intakes for calcium and vitamin d Washington (DC); 2011.

2. Shapses SA, Manson JE. Vitamin d and prevention of cardiovascular disease and diabetes: Why the evidence falls short. *JAMA* 2011;305:2565–2566 [PubMed: 21693745]
3. Towler DA. Calcitropic hormones and arterial physiology: “D”-lightful insights. *J Am Soc Nephrol* 2007;18:369–373 [PubMed: 17259596]
4. Veloudi P, Jones G, Sharman JE. Effectiveness of vitamin d supplementation for cardiovascular health outcomes. *Pulse (Basel)* 2017;4:193–207 [PubMed: 28229054]
5. Fraser DR. Vitamin d. *Lancet* 1995;345:104–107 [PubMed: 7815853]
6. Constans J Group-specific component is not only a vitamin-d-binding protein. *Exp Clin Immunogenet* 1992;9:161–175 [PubMed: 1303095]
7. Bays HE, Neff D, Tomassini JE, Terhakovec AM. Ezetimibe: Cholesterol lowering and beyond. *Expert Rev Cardiovasc Ther* 2008;6:447–470 [PubMed: 18402536]
8. Haddad JG, Matsuoka LY, Hollis BW, Hu YZ, Wortsman J. Human plasma transport of vitamin d after its endogenous synthesis. *J Clin Invest* 1993;91:2552–2555 [PubMed: 8390483]
9. Holick MF. Vitamin d deficiency. *N Engl J Med* 2007;357:266–281 [PubMed: 17634462]
10. Jones KS, Assar S, Harnpanich D, Bouillon R, Lambrechts D, Prentice A, Schoenmakers I. 25(oh)d2 half-life is shorter than 25(oh)d3 half-life and is influenced by dbp concentration and genotype. *J Clin Endocrinol Metab* 2014;99:3373–3381 [PubMed: 24885631]
11. Oliveri B, Mastaglia SR, Brito GM, Seijo M, Keller GA, Somoza J, Diez RA, Di Girolamo G. Vitamin d3 seems more appropriate than d2 to sustain adequate levels of 25ohd: A pharmacokinetic approach. *Eur J Clin Nutr* 2015;69:697–702 [PubMed: 25782422]
12. Heaney RP, Recker RR, Grote J, Horst RL, Armas LA. Vitamin d(3) is more potent than vitamin d(2) in humans. *J Clin Endocrinol Metab* 2011;96:E447–452 [PubMed: 21177785]
13. Rosenstreich SJ, Rich C, Volwiler W. Deposition in and release of vitamin d3 from body fat: Evidence for a storage site in the rat. *J Clin Invest* 1971;50:679–687 [PubMed: 4322721]
14. Dueland S, Holmberg I, Berg T, Pedersen JI. Uptake and 25-hydroxylation of vitamin d3 by isolated rat liver cells. *J Biol Chem* 1981;256:10430–10434 [PubMed: 6270111]
15. Martinaityte I, Kamycheva E, Didriksen A, Jakobsen J, Jorde R. Vitamin d stored in fat tissue during a 5-year intervention affects serum 25-hydroxyvitamin d levels the following year. *J Clin Endocrinol Metab* 2017;102:3731–3738 [PubMed: 28973683]
16. Arnaud SB, Goldsmith RS, Lambert PW, Go VL. 25-hydroxyvitamin d3: Evidence of an enterohepatic circulation in man. *Proc Soc Exp Biol Med* 1975;149:570–572 [PubMed: 1153436]
17. Lawson DE, Paul AA, Black AE, Cole TJ, Mandal AR, Davie M. Relative contributions of diet and sunlight to vitamin d state in the elderly. *Br Med J* 1979;2:303–305 [PubMed: 476435]
18. Scragg R, Stewart AW, Waayer D, Lawes CMM, Toop L, Sluyter J, Murphy J, Khaw KT, Camargo CA, Jr. Effect of monthly high-dose vitamin d supplementation on cardiovascular disease in the vitamin d assessment study : A randomized clinical trial. *JAMA Cardiol* 2017;2:608–616 [PubMed: 28384800]
19. Manson JE, Hsia J, Johnson KC, Rossouw JE, Assaf AR, Lasser NL, Trevisan M, Black HR, Heckbert SR, Detrano R, Strickland OL, Wong ND, Crouse JR, Stein E, Cushman M, Women’s Health Initiative I. Estrogen plus progestin and the risk of coronary heart disease. *N Engl J Med* 2003;349:523–534 [PubMed: 12904517]
20. Kamboh MI, Ferrell RE. Ethnic variation in vitamin d-binding protein (gc): A review of isoelectric focusing studies in human populations. *Hum Genet* 1986;72:281–293 [PubMed: 3516862]
21. Sinotte M, Diorio C, Berube S, Pollak M, Brisson J. Genetic polymorphisms of the vitamin d binding protein and plasma concentrations of 25-hydroxyvitamin d in premenopausal women. *Am J Clin Nutr* 2009;89:634–640 [PubMed: 19116321]
22. Loughran G, Jungreis I, Tzani I, Power M, Dmitriev RI, Ivanov IP, Kellis M, Atkins JF. Stop codon readthrough generates a c-terminally extended variant of the human vitamin d receptor with reduced calcitriol response. *J Biol Chem* 2018
23. Gangloff A, Bergeron J, Lemieux I, Despres JP. Changes in circulating vitamin d levels with loss of adipose tissue. *Curr Opin Clin Nutr Metab Care* 2016;19:464–470 [PubMed: 27537278]
24. Abboud M, Rybchyn MS, Ning YJ, Brennan-Speranza TC, Giris CM, Gunton JE, Fraser DR, Mason RS. 1,25-dihydroxycholecalciferol (calcitriol) modifies uptake and release of 25-

- hydroxycholecalciferol in skeletal muscle cells in culture. *J Steroid Biochem Mol Biol* 2018;177:109–115 [PubMed: 29107178]
25. Pludowski P, Holick MF, Grant WB, Konstantynowicz J, Mascarenhas MR, Haq A, Povorozyuk V, Balatska N, Barbosa AP, Karonova T, Rudenka E, Misiorowski W, Zakharova I, Rudenka A, Lukaszkiwicz J, Marcinowska-Suchowierska E, Laszcz N, Abramowicz P, Bhattoa HP, Wimalawansa SJ. Vitamin d supplementation guidelines. *J Steroid Biochem Mol Biol* 2018;175:125–135 [PubMed: 28216084]
 26. Wang TJ, Pencina MJ, Booth SL, Jacques PF, Ingelsson E, Lanier K, Benjamin EJ, D'Agostino RB, Wolf M, Vasani RS. Vitamin d deficiency and risk of cardiovascular disease. *Circulation* 2008;117:503–511 [PubMed: 18180395]
 27. Melamed ML, Michos ED, Post W, Astor B. 25-hydroxyvitamin d levels and the risk of mortality in the general population. *Arch Intern Med* 2008;168:1629–1637 [PubMed: 18695076]
 28. Ellam T, Hameed A, ul Haque R, Muthana M, Wilkie M, Francis SE, Chico TJ. Vitamin d deficiency and exogenous vitamin d excess similarly increase diffuse atherosclerotic calcification in apolipoprotein e knockout mice. *PLoS One* 2014;9:e88767 [PubMed: 24586387]
 29. Pilz S, Verheyen N, Grubler MR, Tomaschitz A, Marz W. Vitamin d and cardiovascular disease prevention. *Nat Rev Cardiol* 2016;13:404–417 [PubMed: 27150190]
 30. Seibert E, Lehmann U, Riedel A, Ulrich C, Hirche F, Brandsch C, Dierkes J, Girndt M, Stangl GI. Vitamin d3 supplementation does not modify cardiovascular risk profile of adults with inadequate vitamin d status. *Eur J Nutr* 2017;56:621–634 [PubMed: 26621634]
 31. Cupisti A, Vigo V, Baronti ME, D'Alessandro C, Ghiadoni L, Egidi MF. Vitamin d status and cholecalciferol supplementation in chronic kidney disease patients: An italian cohort report. *Int J Nephrol Renovasc Dis* 2015;8:151–157 [PubMed: 26640388]
 32. Chitalia N, Ismail T, Tooth L, Boa F, Hampson G, Goldsmith D, Kaski JC, Banerjee D. Impact of vitamin d supplementation on arterial vasomotion, stiffness and endothelial biomarkers in chronic kidney disease patients. *PLoS One* 2014;9:e91363 [PubMed: 24646518]
 33. Kumar V, Yadav AK, Lal A, Kumar V, Singhal M, Billot L, Gupta KL, Banerjee D, Jha V. A randomized trial of vitamin d supplementation on vascular function in ckd. *J Am Soc Nephrol* 2017;28:3100–3108 [PubMed: 28667080]
 34. Levin A, Tang M, Perry T, Zalunardo N, Beaulieu M, Dubland JA, Zerr K, Djurdjev O. Randomized controlled trial for the effect of vitamin d supplementation on vascular stiffness in ckd. *Clin J Am Soc Nephrol* 2017;12:1447–1460 [PubMed: 28550081]
 35. Bhan I, Dobens D, Tamez H, Deferio JJ, Li YC, Warren HS, Ankers E, Wenger J, Tucker JK, Trottier C, Pathan F, Kalim S, Nigwekar SU, Thadhani R. Nutritional vitamin d supplementation in dialysis: A randomized trial. *Clin J Am Soc Nephrol* 2015;10:611–619 [PubMed: 25770176]
 36. Garcia-Lopes MG, Pillar R, Kamimura MA, Rocha LA, Canziani ME, Carvalho AB, Cuppari L. Cholecalciferol supplementation in chronic kidney disease: Restoration of vitamin d status and impact on parathyroid hormone. *Ann Nutr Metab* 2012;61:74–82 [PubMed: 22889840]
 37. Lu RJ, Zhu SM, Tang FL, Zhu XS, Fan ZD, Wang GL, Jiang YF, Zhang Y. Effects of vitamin d or its analogues on the mortality of patients with chronic kidney disease: An updated systematic review and meta-analysis. *Eur J Clin Nutr* 2017;71:683–693 [PubMed: 28488689]
 38. Mann MC, Hobbs AJ, Hemmelgarn BR, Roberts DJ, Ahmed SB, Rabi DM. Effect of oral vitamin d analogs on mortality and cardiovascular outcomes among adults with chronic kidney disease: A meta-analysis. *Clin Kidney J* 2015;8:41–48 [PubMed: 25713709]
 39. Zheng Z, Shi H, Jia J, Li D, Lin S. Vitamin d supplementation and mortality risk in chronic kidney disease: A meta-analysis of 20 observational studies. *BMC Nephrol* 2013;14:199 [PubMed: 24066946]
 40. Haddad JG. Plasma vitamin d-binding protein (gc-globulin): Multiple tasks. *J Steroid Biochem Mol Biol* 1995;53:579–582 [PubMed: 7626513]
 41. Christensen EI, Birn H. Megalin and cubilin: Multifunctional endocytic receptors. *Nat Rev Mol Cell Biol* 2002;3:256–266 [PubMed: 11994745]
 42. Teramoto T, Endo K, Ikeda K, Kubodera N, Kinoshita M, Yamanaka M, Ogata E. Binding of vitamin d to low-density-lipoprotein (ldl) and ldl receptor-mediated pathway into cells. *Biochem Biophys Res Commun* 1995;215:199–204 [PubMed: 7575591]

43. Navab M, Fogelman AM, Berliner JA, Territo MC, Demer LL, Frank JS, Watson AD, Edwards PA, Lusis AJ. Pathogenesis of atherosclerosis. *Am J Cardiol* 1995;76:18C–23C
44. Hsu JJ, Tintut Y, Demer LL. Vitamin d and osteogenic differentiation in the artery wall. *Clin J Am Soc Nephrol* 2008;3:1542–1547 [PubMed: 18562594]
45. Schwartz N, Verma A, Bivens CB, Schwartz Z, Boyan BD. Rapid steroid hormone actions via membrane receptors. *Biochim Biophys Acta* 2016;1863:2289–2298 [PubMed: 27288742]
46. Homme M, Schmitt CP, Himmele R, Hoffmann GF, Mehls O, Schaefer F. Vitamin d and dexamethasone inversely regulate parathyroid hormone-induced regulator of g protein signaling-2 expression in osteoblast-like cells. *Endocrinology* 2003;144:2496–2504 [PubMed: 12746312]
47. Obradovic D, Gronemeyer H, Lutz B, Rein T. Cross-talk of vitamin d and glucocorticoids in hippocampal cells. *J Neurochem* 2006;96:500–509 [PubMed: 16336217]
48. Zhou C, Assem M, Tay JC, Watkins PB, Blumberg B, Schuetz EG, Thummel KE. Steroid and xenobiotic receptor and vitamin d receptor crosstalk mediates cyp24 expression and drug-induced osteomalacia. *J Clin Invest* 2006;116:1703–1712 [PubMed: 16691293]
49. Alimirah F, Peng X, Yuan L, Mehta RR, von Knethen A, Choubey D, Mehta RG. Crosstalk between the peroxisome proliferator-activated receptor gamma (ppargamma) and the vitamin d receptor (vdr) in human breast cancer cells: Ppargamma binds to vdr and inhibits 1alpha,25-dihydroxyvitamin d3 mediated transactivation. *Exp Cell Res* 2012;318:2490–2497 [PubMed: 22884583]
50. Shamsuzzaman S, Onal M, St John HC, Jeffery JJ, Pike JW. Absence of the vitamin d receptor inhibits atherosclerotic plaque calcification in female hypercholesterolemic mice. *J Cell Biochem* 2017;118:1050–1064 [PubMed: 27567005]
51. Tukaj C, Trzonkowski P, Kubasik-Juraniec J, Mysliwski A. Quantifying division of aortal smooth muscle cells in culture stimulated by 1,25(oh)2d3. *J Steroid Biochem Mol Biol* 2007;103:525–528 [PubMed: 17368183]
52. Parker BD, Schurgers LJ, Brandenburg VM, Christenson RH, Vermeer C, Ketteler M, Shlipak MG, Whooley MA, Ix JH. The associations of fibroblast growth factor 23 and uncarboxylated matrix gla protein with mortality in coronary artery disease: The heart and soul study. *Ann Intern Med* 2010;152:640–648 [PubMed: 20479029]
53. Reid IR, Home AM, Mihov B, Gamble GD, Al-Abuwsfi F, Singh M, Taylor L, Fenwick S, Camargo CA, Stewart AW, Scragg R. Effect of monthly high-dose vitamin d on bone density in community-dwelling older adults substudy of a randomized controlled trial. *J Intern Med* 2017;282:452–460 [PubMed: 28692172]
54. Tintut Y, Parhami F, Bostrom K, Jackson SM, Demer LL. Camp stimulates osteoblast-like differentiation of calcifying vascular cells. Potential signaling pathway for vascular calcification. *J Biol Chem* 1998;273:7547–7553 [PubMed: 9516456]
55. Jono S, Nishizawa Y, Shioi A, Morii H. 1,25-dihydroxyvitamin d3 increases in vitro vascular calcification by modulating secretion of endogenous parathyroid hormone-related peptide. *Circulation* 1998;98:1302–1306 [PubMed: 9751679]
56. Price PA, Buckley JR, Williamson MK. The amino bisphosphonate ibandronate prevents vitamin d toxicity and inhibits vitamin d-induced calcification of arteries, cartilage, lungs and kidneys in rats. *J Nutr* 2001;131:2910–2915 [PubMed: 11694617]
57. Shi Y, Lu W, Hou Y, Fu K, Gan F, Liu J. Fibroblast growth factor 21 ameliorates vascular calcification by inhibiting osteogenic transition in vitamin d3 plus nicotine-treated rats. *Biochem Biophys Res Commun* 2018;495:2448–2455 [PubMed: 29273504]
58. Torremade N, Bozic M, Panizo S, Barrio-Vazquez S, Fernandez-Martin JL, Encinas M, Goltzman D, Arcidiacono MV, Fernandez E, Valdivielso JM. Vascular calcification induced by chronic kidney disease is mediated by an increase of 1alpha-hydroxylase expression in vascular smooth muscle cells. *J Bone Miner Res* 2016;31:1865–1876 [PubMed: 27074284]
59. Colotta F, Jansson B, Bonelli F. Modulation of inflammatory and immune responses by vitamin d. *J Autoimmun* 2017;85:78–97 [PubMed: 28733125]
60. Hollis BW, Johnson D, Hulsey TC, Ebeling M, Wagner CL. Vitamin d supplementation during pregnancy: Double-blind, randomized clinical trial of safety and effectiveness. *J Bone Miner Res* 2011;26:2341–2357 [PubMed: 21706518]

61. Hollis BW, Wagner CL. Clinical review: The role of the parent compound vitamin d with respect to metabolism and function: Why clinical dose intervals can affect clinical outcomes. *J Clin Endocrinol Metab* 2013;98:4619–4628 [PubMed: 24106283]
62. Committee IoM. Dietary reference intakes for calcium and vitamin d 2011
63. Davies H Coronary heart disease: The significance of coronary pathology in infancy and the role of mitogens such as vitamin d. *Med Hypotheses* 1989;30:179–185 [PubMed: 2689846]
64. Manson JE, Bassuk SS, Lee IM, Cook NR, Albert MA, Gordon D, Zaharris E, Macfadyen JG, Danielson E, Lin J, Zhang SM, Buring JE. The vitamin d and omega-3 trial (vital): Rationale and design of a large randomized controlled trial of vitamin d and marine omega-3 fatty acid supplements for the primary prevention of cancer and cardiovascular disease. *Contemp Clin Trials* 2012;33:159–171 [PubMed: 21986389]
65. Taussig HB. Possible injury to the cardiovascular system from vitamin d. *Ann Intern Med* 1966;65:1195–1200 [PubMed: 5333232]
66. Hosseinpour F, Ibranovic I, Tang W, Wikvall K. 25-hydroxylation of vitamin d3 in primary cultures of pig hepatocytes: Evidence for a role of both cyp2d25 and cyp27a1. *Biochem Biophys Res Commun* 2003;303:877–883 [PubMed: 12670492]
67. Chen ML, Boltz MA, Armbrecht HJ. Effects of 1,25-dihydroxyvitamin d3 and phorbol ester on 25-hydroxyvitamin d3 24-hydroxylase cytochrome p450 messenger ribonucleic acid levels in primary cultures of rat renal cells. *Endocrinology* 1993;132:1782–1788 [PubMed: 7681765]
68. Zehnder D, Bland R, Chana RS, Wheeler DC, Howie AJ, Williams MC, Stewart PM, Hewison M. Synthesis of 1,25-dihydroxyvitamin d(3) by human endothelial cells is regulated by inflammatory cytokines: A novel autocrine determinant of vascular cell adhesion. *J Am Soc Nephrol* 2002;13:621–629 [PubMed: 11856765]
69. Somjen D, Weisman Y, Kohen F, Gayer B, Limor R, Sharon O, Jaccard N, Knoll E, Stern N. 25-hydroxyvitamin d3-1alpha-hydroxylase is expressed in human vascular smooth muscle cells and is upregulated by parathyroid hormone and estrogenic compounds. *Circulation* 2005;111:1666–1671 [PubMed: 15795327]
70. Stoffels K, Overbergh L, Giulietti A, Verlinden L, Bouillon R, Mathieu C. Immune regulation of 25-hydroxyvitamin-d3-1alpha-hydroxylase in human monocytes. *J Bone Miner Res* 2006;21:37–47 [PubMed: 16355272]
71. Girgis CM, Clifton-Bligh RJ, Mokbel N, Cheng K, Gunton JE. Vitamin d signaling regulates proliferation, differentiation, and myotube size in c2c12 skeletal muscle cells. *Endocrinology* 2014;155:347–357 [PubMed: 24280059]
72. Prentice A Vitamin d deficiency: A global perspective. *Nutr Rev* 2008;66:S153–164 [PubMed: 18844843]
73. Long GG. Acute toxicosis in swine associated with excessive dietary intake of vitamin d. *J Am Vet Med Assoc* 1984;184:164–170 [PubMed: 6321415]
74. Szeto FL, Reardon CA, Yoon D, Wang Y, Wong KE, Chen Y, Kong J, Liu SQ, Thadhani R, Getz GS, Li YC. Vitamin d receptor signaling inhibits atherosclerosis in mice. *Mol Endocrinol* 2012;26:1091–1101 [PubMed: 22638071]
75. Chen S, Law CS, Grigsby CL, Olsen K, Hong TT, Zhang Y, Yeghiazarians Y, Gardner DG. Cardiomyocyte-specific deletion of the vitamin d receptor gene results in cardiac hypertrophy. *Circulation* 2011;124:1838–1847 [PubMed: 21947295]
76. Aihara K, Azuma H, Akaike M, Ikeda Y, Yamashita M, Sudo T, Hayashi H, Yamada Y, Endoh F, Fujimura M, Yoshida T, Yamaguchi H, Hashizume S, Kato M, Yoshimura K, Yamamoto Y, Kato S, Matsumoto T. Disruption of nuclear vitamin d receptor gene causes enhanced thrombogenicity in mice. *J Biol Chem* 2004;279:35798–35802 [PubMed: 15205460]
77. Suda T, Masuyama R, Bouillon R, Carmeliet G. Physiological functions of vitamin d: What we have learned from global and conditional vdr knockout mouse studies. *Curr Opin Pharmacol* 2015;22:87–99 [PubMed: 25938686]
78. Norman P, Moss I, Sian M, Gosling M, Powell J. Maternal and postnatal vitamin d ingestion influences rat aortic structure, function and elastin content. *Cardiovasc Res* 2002;55:369–374 [PubMed: 12123776]

79. Jiang W, Miyamoto T, Kakizawa T, Nishio SI, Oiwa A, Takeda T, Suzuki S, Hashizume K. Inhibition of Ixralpha signaling by vitamin d receptor: Possible role of vdr in bile acid synthesis. *Biochem Biophys Res Commun* 2006;351:176–184 [PubMed: 17054913]
80. Shimosawa T, Ando K, Fujita T. Enhancement of vasoconstrictor response by a noncalcemic analogue of vitamin d3. *Hypertension* 1993;21:253–258 [PubMed: 8428788]
81. Kang EJ, Lee JE, An SM, Lee JH, Kwon HS, Kim BC, Kim SJ, Kim JM, Hwang DY, Jung YJ, Yang SY, Kim SC, An BS. The effects of vitamin d3 on lipogenesis in the liver and adipose tissue of pregnant rats. *Int J Mol Med* 2015;36:1151–1158 [PubMed: 26239543]
82. Karadag C, Yoldemir T, Yavuz DG. Effects of vitamin d supplementation on insulin sensitivity and androgen levels in vitamin-d-deficient polycystic ovary syndrome patients. *J Obstet Gynaecol Res* 2018;44:270–277 [PubMed: 29094433]
83. Wong MS, Delansorne R, Man RY, Vanhoutte PM. Vitamin d derivatives acutely reduce endothelium-dependent contractions in the aorta of the spontaneously hypertensive rat. *Am J Physiol Heart Circ Physiol* 2008;295:H289–296 [PubMed: 18487433]
84. Sooy K, Sabbagh Y, Demay MB. Osteoblasts lacking the vitamin d receptor display enhanced osteogenic potential in vitro. *J Cell Biochem* 2005;94:81–87 [PubMed: 15517598]
85. Takasu H, Sugita A, Uchiyama Y, Katagiri N, Okazaki M, Ogata E, Ikeda K. C-fos protein as a target of anti-osteoclastogenic action of vitamin d, and synthesis of new analogs. *J Clin Invest* 2006;116:528–535 [PubMed: 16424941]
86. Tsonis PA. 1,25-dihydroxyvitamin d3 stimulates chondrogenesis of the chick limb bud mesenchymal cells. *Dev Biol* 1991;143:130–134 [PubMed: 1845863]
87. Wagatsuma A, Sakuma K. Vitamin d signaling in myogenesis: Potential for treatment of sarcopenia. *Biomed Res Int* 2014;2014:121254 [PubMed: 25197630]
88. Blumberg JM, Tzamelis I, Astapova I, Lam FS, Flier JS, Hollenberg AN. Complex role of the vitamin d receptor and its ligand in adipogenesis in 3t3-l1 cells. *J Biol Chem* 2006;281:11205–11213 [PubMed: 16467308]
89. Bunce CM, Brown G, Hewison M. Vitamin d and hematopoiesis. *Trends Endocrinol Metab* 1997;8:245–251 [PubMed: 18406812]
90. Chabas JF, Stephan D, Marqueste T, Garcia S, Lavaut MN, Nguyen C, Legre R, Khrestchatsky M, Decherchi P, Feron F. Cholecalciferol (vitamin d(3)) improves myelination and recovery after nerve injury. *PLoS One* 2013;8:e65034 [PubMed: 23741446]
91. Wu-Wong JR, Nakane M, Ma J, Ruan X, Kroeger PE. Effects of vitamin d analogs on gene expression profiling in human coronary artery smooth muscle cells. *Atherosclerosis* 2006;186:20–28 [PubMed: 16095599]
92. Rebsamen MC, Sun J, Norman AW, Liao JK. 1alpha,25-dihydroxyvitamin d3 induces vascular smooth muscle cell migration via activation of phosphatidylinositol 3-kinase. *Circ Res* 2002;91:17–24 [PubMed: 12114317]
93. Bukoski RD, DeWan P, McCarron DA. 1,25 (oh)2 vitamin d3 modifies growth and contractile function of vascular smooth muscle of spontaneously hypertensive rats. *Am J Hypertens* 1989;2:553–556 [PubMed: 2757811]
94. Shan NL, Wahler J, Lee HJ, Bak MJ, Gupta SD, Maehr H, Suh N. Vitamin d compounds inhibit cancer stem-like cells and induce differentiation in triple negative breast cancer. *J Steroid Biochem Mol Biol* 2017;173:122–129 [PubMed: 27923595]
95. Oh J, Weng S, Felton SK, Bhandare S, Riek A, Butler B, Proctor BM, Petty M, Chen Z, Schechtman KB, Bernal-Mizrachi L, Bernal-Mizrachi C. 1,25(oh)2 vitamin d inhibits foam cell formation and suppresses macrophage cholesterol uptake in patients with type 2 diabetes mellitus. *Circulation* 2009;120:687–698 [PubMed: 19667238]
96. Correale J, Ysraelit MC, Gaitan MI. Vitamin d-mediated immune regulation in multiple sclerosis. *J Neurol Sci* 2011;311:23–31 [PubMed: 21723567]
97. Wagner KD, Wagner N, Sukhatme VP, Scholz H. Activation of vitamin d receptor by the wilms' tumor gene product mediates apoptosis of renal cells. *J Am Soc Nephrol* 2001;12:1188–1196 [PubMed: 11373341]

98. Marie PJ, Connes D, Hott M, Miravet L. Comparative effects of a novel vitamin d analogue mc-903 and 1,25-dihydroxyvitamin d3 on alkaline phosphatase activity, osteocalcin and DNA synthesis by human osteoblastic cells in culture. *Bone* 1990;11:171–179 [PubMed: 2390375]
99. Schwartz Z, Swain LD, Ramirez V, Boyan BD. Regulation of arachidonic acid turnover by 1,25-(oh)2d3 and 24,25-(oh)2d3 in growth zone and resting zone chondrocyte cultures. *Biochim Biophys Acta* 1990;1027:278–286 [PubMed: 2397237]
100. Grosse B, Bourdeau A, Lieberherr M. Oscillations in inositol 1,4,5-trisphosphate and diacylglycerol induced by vitamin d3 metabolites in confluent mouse osteoblasts. *J Bone Miner Res* 1993;8:1059–1069 [PubMed: 8237475]
101. Liu X, Nelson A, Wang X, Farid M, Gunji Y, Ikari J, Iwasawa S, Basma H, Feghali-Bostwick C, Rennard SI. Vitamin d modulates prostaglandin e2 synthesis and degradation in human lung fibroblasts. *Am J Respir Cell Mol Biol* 2014;50:40–50 [PubMed: 23941558]
102. Schwartz Z, Swain LD, Kelly DW, Brooks B, Boyan BD. Regulation of prostaglandin e2 production by vitamin d metabolites in growth zone and resting zone chondrocyte cultures is dependent on cell maturation. *Bone* 1992;13:395–401 [PubMed: 1419381]
103. Polidoro L, Properzi G, Marampon F, Gravina GL, Festuccia C, Di Cesare E, Scarsella L, Ciccarelli C, Zani BM, Ferri C. Vitamin d protects human endothelial cells from h(2)o(2) oxidant injury through the mek/erk-sirt1 axis activation. *J Cardiovasc Transl Res* 2013;6:221–231 [PubMed: 23247634]
104. Andrukhova O, Slavic S, Zeitz U, Riesen SC, Heppelmann MS, Ambrisko TD, Markovic M, Kuebler WM, Erben RG. Vitamin d is a regulator of endothelial nitric oxide synthase and arterial stiffness in mice. *Mol Endocrinol* 2014;28:53–64 [PubMed: 24284821]
105. Bissonnette M, Tien XY, Niedziela SM, Hartmann SC, Frawley BP, Jr., Roy HK, Sitrin MD, Perlman RL, Brasitus TA. 1,25(oh)2 vitamin d3 activates pkc-alpha in caco-2 cells: A mechanism to limit secosteroid-induced rise in [ca2+]i. *Am J Physiol* 1994;267:G465–475 [PubMed: 7943245]
106. Berg JP, Haug E. Vitamin d: A hormonal regulator of the camp signaling pathway. *Crit Rev Biochem Mol Biol* 1999;34:315–323 [PubMed: 10565677]
107. Pardo VG, Boland R, de Boland AR. 1alpha,25(oh)(2)-vitamin d(3) stimulates intestinal cell p38 mapk activity and increases c-fos expression. *Int J Biochem Cell Biol* 2006;38:1181–1190 [PubMed: 16483831]
108. Piek E, Sleumer LS, van Someren EP, Heuvel L, de Haan JR, de Grijs I, Gilissen C, Hendriks JM, van Ravestein-van Os RI, Bauerschmidt S, Dechering KJ, van Zoelen EJ. Osteo-transcriptomics of human mesenchymal stem cells: Accelerated gene expression and osteoblast differentiation induced by vitamin d reveals c-myc as an enhancer of bmp2-induced osteogenesis. *Bone* 2010;46:613–627 [PubMed: 19857615]
109. Towers TL, Staeva TP, Freedman LP. A two-hit mechanism for vitamin d3-mediated transcriptional repression of the granulocyte-macrophage colony-stimulating factor gene: Vitamin d receptor competes for DNA binding with nfat1 and stabilizes c-jun. *Mol Cell Biol* 1999;19:4191–4199 [PubMed: 10330159]
110. Fretz JA, Zella LA, Kim S, Shevde NK, Pike JW. 1,25-dihydroxyvitamin d3 regulates the expression of low-density lipoprotein receptor-related protein 5 via deoxyribonucleic acid sequence elements located downstream of the start site of transcription. *Mol Endocrinol* 2006;20:2215–2230 [PubMed: 16613987]
111. Chen Y, Zhang J, Ge X, Du J, Deb DK, Li YC. Vitamin d receptor inhibits nuclear factor kappa b activation by interacting with ikappa b kinase beta protein. *J Biol Chem* 2013;288:19450–19458 [PubMed: 23671281]
112. Okamura M, Takano Y, Saito Y, Yao J, Kitamura M. Induction of nephrin gene expression by selective cooperation of the retinoic acid receptor and the vitamin d receptor. *Nephrol Dial Transplant* 2009;24:3006–3012 [PubMed: 19474283]
113. Zou A, Elgort MG, Allegretto EA. Retinoid x receptor (rxr) ligands activate the human 25-hydroxyvitamin d3–24-hydroxylase promoter via rxr heterodimer binding to two vitamin d-responsive elements and elicit additive effects with 1,25-dihydroxyvitamin d3. *J Biol Chem* 1997;272:19027–19034 [PubMed: 9228086]

114. Zhang Y, Leung DY, Goleva E. Vitamin d enhances glucocorticoid action in human monocytes: Involvement of granulocyte-macrophage colony-stimulating factor and mediator complex subunit 14. *J Biol Chem* 2013;288:14544–14553 [PubMed: 23572530]
115. Paredes R, Arriagada G, Cruzat F, Olate J, Van Wijnen A, Lian J, Stein G, Stein J, Montecino M. The runx2 transcription factor plays a key role in the 1 α ,25-dihydroxy vitamin d₃-dependent upregulation of the rat osteocalcin (oc) gene expression in osteoblastic cells. *J Steroid Biochem Mol Biol* 2004;89-90:269–271 [PubMed: 15225783]
116. Rohe B, Safford SE, Nemere I, Farach-Carson MC. Regulation of expression of 1,25d₃-marrs/erp57/pdia3 in rat iec-6 cells by tgf beta and 1,25(oh)2d₃. *Steroids* 2007;72:144–150 [PubMed: 17188725]
117. Schedlich LJ, Muthukaruppan A, O'Han MK, Baxter RC. Insulin-like growth factor binding protein-5 interacts with the vitamin d receptor and modulates the vitamin d response in osteoblasts. *Mol Endocrinol* 2007;21:2378–2390 [PubMed: 17595320]
118. Miyake N, Hoshi K, Sano Y, Kikuchi K, Tadano K, Koshihara Y. 1,25-dihydroxyvitamin d₃ promotes vitamin k₂ metabolism in human osteoblasts. *Osteoporos Int* 2001;12:680–687 [PubMed: 11580082]
119. Cheema C, Grant BF, Marcus R. Effects of estrogen on circulating “free” and total 1,25-dihydroxyvitamin d and on the parathyroid-vitamin d axis in postmenopausal women. *J Clin Invest* 1989;83:537–542 [PubMed: 2492309]
120. Ponda MP, Dowd K, Finkielstein D, Holt PR, Breslow JL. The short-term effects of vitamin d repletion on cholesterol: A randomized, placebo-controlled trial. *Arterioscler Thromb Vasc Biol* 2012;32:2510–2515 [PubMed: 22947589]
121. Fu B, Wang H, Wang J, Barouhas I, Liu W, Shuboy A, Bushinsky DA, Zhou D, Favus MJ. Epigenetic regulation of bmp2 by 1,25-dihydroxyvitamin d₃ through DNA methylation and histone modification. *PLoS One* 2013;8:e61423 [PubMed: 23620751]
122. Kim HS, Zheng M, Kim DK, Lee WP, Yu SJ, Kim BO. Effects of 1,25-dihydroxyvitamin d₃ on the differentiation of mc3t3-e1 osteoblast-like cells. *J Periodontal Implant Sci* 2018;48:34–46 [PubMed: 29535889]
123. Sodek J, Li JJ, Kim RH, Ogata Y, Yamauchi M. Characterization of the bone sialoprotein (bsp) gene promoter. *Connect Tissue Res* 1996;35:23–31 [PubMed: 9084640]
124. Kuroki Y, Shiozawa S, Kano J, Chihara K. Competition between c-fos and 1,25(oh)₂ vitamin d₃ in the transcriptional control of type i collagen synthesis in mc3t3-e1 osteoblastic cells. *J Cell Physiol* 1995;164:459–464 [PubMed: 7650055]
125. Merchiers P, Bulens F, Stockmans I, De Vriese A, Convents R, Bouillon R, Collen D, Belayew A, Carmeliet G. 1,25-dihydroxyvitamin d(3) induction of the tissue-type plasminogen activator gene is mediated through its multihormone-responsive enhancer. *FEBS Lett* 1999;460:289–296 [PubMed: 10544252]
126. Liu SM, Koszewski N, Lupez M, Malluche HH, Olivera A, Russell J. Characterization of a response element in the 5'-flanking region of the avian (chicken) pth gene that mediates negative regulation of gene transcription by 1,25-dihydroxyvitamin d₃ and binds the vitamin d₃ receptor. *Mol Endocrinol* 1996;10:206–215 [PubMed: 8825560]
127. Tang WJ, Wang LF, Xu XY, Zhou Y, Jin WF, Wang HF, Gao J. Autocrine/paracrine action of vitamin d on fgf23 expression in cultured rat osteoblasts. *Calcif Tissue Int* 2010;86:404–410 [PubMed: 20354682]
128. Kim S, Yamazaki M, Zella LA, Meyer MB, Fretz JA, Shevde NK, Pike JW. Multiple enhancer regions located at significant distances upstream of the transcriptional start site mediate rankl gene expression in response to 1,25-dihydroxyvitamin d₃. *J Steroid Biochem Mol Biol* 2007;103:430–434 [PubMed: 17197168]
129. Hodgkinson JE, Davidson CL, Beresford J, Sharpe PT. Expression of a human homeobox-containing gene is regulated by 1,25(oh)₂d₃ in bone cells. *Biochim Biophys Acta* 1993;1174:11–16 [PubMed: 8101453]
130. Lin YM, Sun HY, Chiu WT, Su HC, Chien YC, Chong LW, Chang HC, Bai CH, Young KC, Tsao CW. Calcitriol inhibits hcv infection via blockade of activation of ppar and interference with endoplasmic reticulum-associated degradation. *Viruses* 2018;10

131. Wietrzyk J, Filip B, Milczarek M, Klopotoska D, Maciejewska M, Dabrowska K, Kurzepa A, Dzimira S, Madej J, Kutner A. The influence of 1,25-dihydroxyvitamin d3 and 1,24-dihydroxyvitamin d3 on alphavbeta3 integrin expression in cancer cell lines. *Oncol Rep* 2008;20:941–952 [PubMed: 18813838]
132. Polly P, Carlberg C, Eisman JA, Morrison NA. Identification of a vitamin d3 response element in the fibronectin gene that is bound by a vitamin d3 receptor homodimer. *J Cell Biochem* 1996;60:322–333 [PubMed: 8867808]
133. Kuhne H, Schutkowski A, Weinholz S, Cordes C, Schierhorn A, Schulz K, Konig B, Stangl GI. Vitamin d receptor regulates intestinal proteins involved in cell proliferation, migration and stress response. *Lipids Health Dis* 2014;13:51 [PubMed: 24641763]
134. Lacraz S, Dayer JM, Nicod L, Welgus HG. 1,25-dihydroxyvitamin d3 dissociates production of interstitial collagenase and 92-kda gelatinase in human mononuclear phagocytes. *J Biol Chem* 1994;269:6485–6490 [PubMed: 7509804]
135. Miyashita M, Koga K, Izumi G, Sue F, Makabe T, Taguchi A, Nagai M, Urata Y, Takamura M, Harada M, Hirata T, Hirota Y, Wada-Hiraike O, Fujii T, Osuga Y. Effects of 1,25-dihydroxy vitamin d3 on endometriosis. *J Clin Endocrinol Metab* 2016;101:2371–2379 [PubMed: 27035829]
136. Song L, Papaioannou G, Zhao H, Luderer HF, Miller C, Dall’Osso C, Nazarian RM, Wagers AJ, Demay MB. The vitamin d receptor regulates tissue resident macrophage response to injury. *Endocrinology* 2016;157:4066–4075 [PubMed: 27526034]
137. Hakim I, Bar-Shavit Z. Modulation of tnf-alpha expression in bone marrow macrophages: Involvement of vitamin d response element. *J Cell Biochem* 2003;88:986–998 [PubMed: 12616536]
138. Tang X, Pan Y, Zhao Y. Vitamin d inhibits the expression of interleukin-8 in human periodontal ligament cells stimulated with porphyromonas gingivalis. *Arch Oral Biol* 2013;58:397–407 [PubMed: 23083515]
139. Maestro B, Davila N, Carranza MC, Calle C. Identification of a vitamin d response element in the human insulin receptor gene promoter. *J Steroid Biochem Mol Biol* 2003;84:223–230 [PubMed: 12711007]
140. Anisiewicz A, Pawlik A, Filip-Psurska B, Turlej E, Dzimira S, Milczarek M, Gdesz K, Papiernik D, Jarosz J, Klopotoska D, Kutner A, Mazur A, Wietrzyk J. Unfavorable effect of calcitriol and its low-calcemic analogs on metastasis of 4t1 mouse mammary gland cancer. *Int J Oncol* 2018;52:103–126 [PubMed: 29115583]
141. Cornet A, Baudet C, Neveu I, Baron-Van Evercooren A, Brachet P, Naveilhan P. 1,25-dihydroxyvitamin d3 regulates the expression of vdr and ngf gene in schwann cells in vitro. *J Neurosci Res* 1998;53:742–746 [PubMed: 9753201]
142. Chen S, Ni XP, Humphreys MH, Gardner DG. 1,25 dihydroxyvitamin d amplifies type a natriuretic peptide receptor expression and activity in target cells. *J Am Soc Nephrol* 2005;16:329–339 [PubMed: 15590756]
143. Krishnan AV, Swami S, Peng L, Wang J, Moreno J, Feldman D. Tissue-selective regulation of aromatase expression by calcitriol: Implications for breast cancer therapy. *Endocrinology* 2010;151:32–42 [PubMed: 19906814]
144. Quach HP, Noh K, Hoi SY, Bruinsma A, Groothuis GMM, Li AP, Chow ECY, Pang KS. Alterations in gene expression in vitamin d-deficiency: Down-regulation of liver cyp7a1 and renal oat3 in mice. *Biopharm Drug Dispos* 2018;39:99–115 [PubMed: 29243851]
145. Meyer MB, Goetsch PD, Pike JW. A downstream intergenic cluster of regulatory enhancers contributes to the induction of cyp24a1 expression by 1alpha,25-dihydroxyvitamin d3. *J Biol Chem* 2010;285:15599–15610 [PubMed: 20236932]
146. Hahn CN, Kerry DM, Omdahl JL, May BK. Identification of a vitamin d responsive element in the promoter of the rat cytochrome p450(24) gene. *Nucleic Acids Res* 1994;22:2410–2416 [PubMed: 8036172]
147. Malloy PJ, Peng L, Wang J, Feldman D. Interaction of the vitamin d receptor with a vitamin d response element in the mullerian-inhibiting substance (mis) promoter: Regulation of mis

expression by calcitriol in prostate cancer cells. *Endocrinology* 2009;150:1580–1587 [PubMed: 19056816]

Author Manuscript

Author Manuscript

Author Manuscript

Author Manuscript

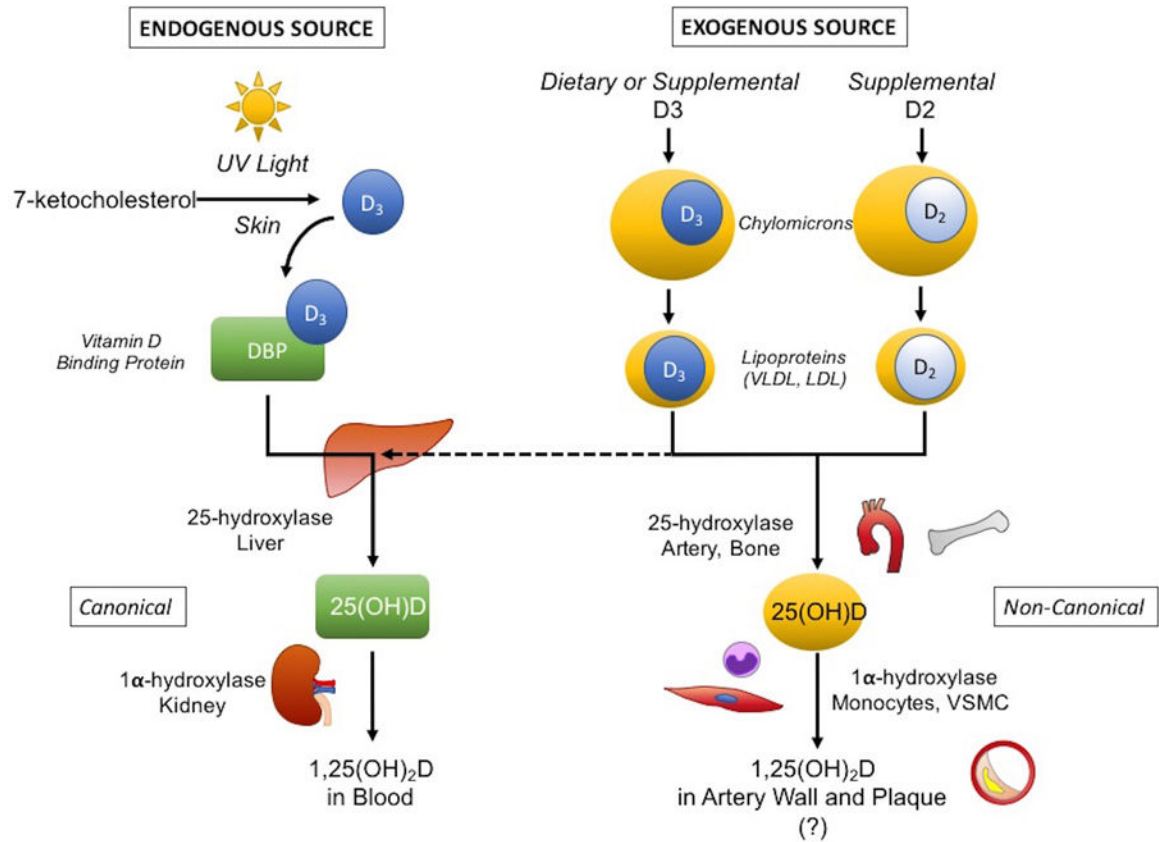


Figure 1. Vitamin D-hormone metabolism, carriers and distribution.

Due to its lipophilic nature, endogenously produced D₃ is carried in the aqueous environment of blood by D-binding protein (DBP), whereas exogenous (dietary and supplemental) D₃ and D₂, absorbed from the intestines, are transported within chylomicrons, which are further processed to lipoproteins (e.g., VLDL and LDL), many of which continue to carry the exogenous D. The conventional sites of vitamin D-hormone metabolism are the liver and proximal tubules of the kidney, where hydroxylases convert it to its active form. But hydroxylase activity is also found in parenchymal and immune cells, including VSMC and monocytes, in other tissues. This raises the potential for accumulation of vitamin D-hormone within LDL in the subendothelial space, where it may undergo activation in atherosclerotic plaque and possibly influence ectopic differentiation and calcification. Abbreviations: LDL, low-density lipoprotein; VLDL, very low-density lipoprotein; VSMC, vascular smooth muscle cells; UV, ultraviolet.

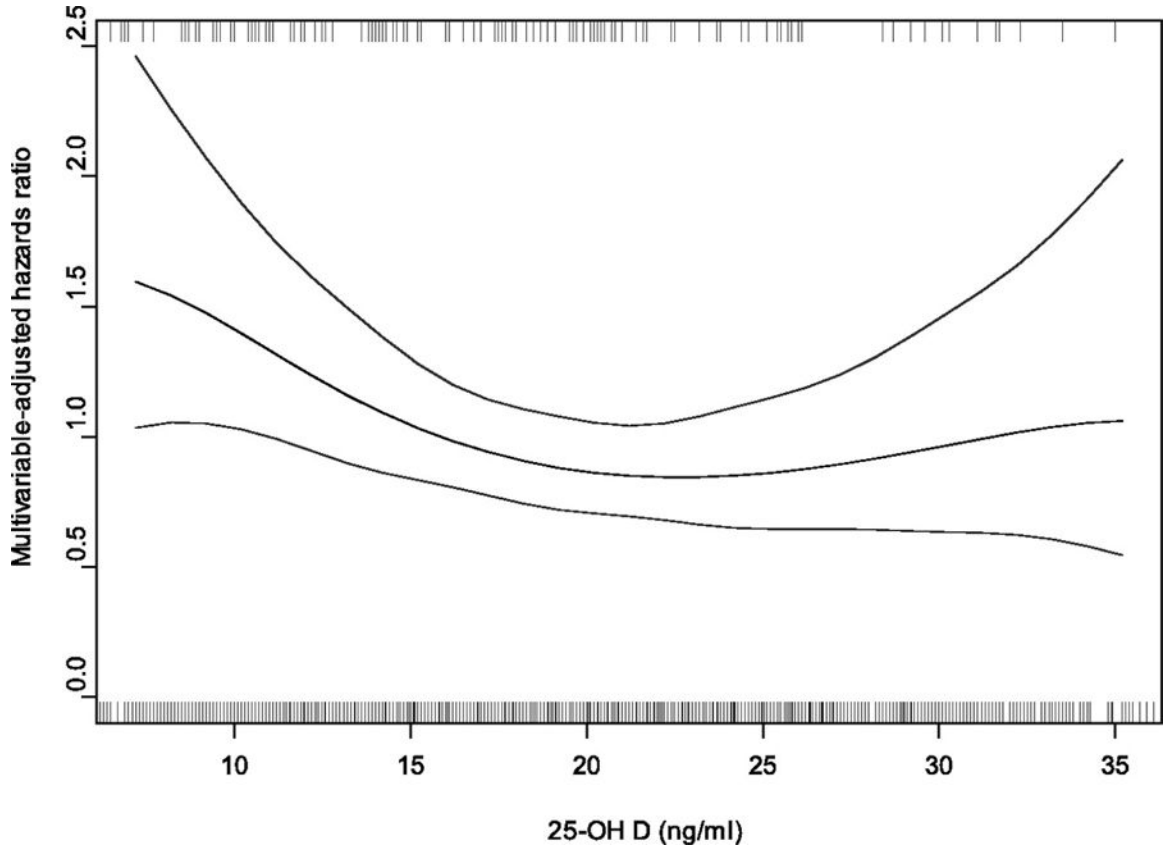


Figure 2. U-shape of the multivariable-adjusted relation between baseline serum 25(OH)D₃ levels and incident cardiovascular events reported by Wang et al. (from *Circulation* 117:503–511, with permission [pending]). Solid lines show the estimated relation of adjusted hazard ratios (with 95% confidence limits) and 25(OH)D₃ levels when time to cardiovascular event is modeled as a function of penalized regression splines of 25(OH)D₃ levels with adjustment for all other covariates. Hatched lines on the horizontal axis represent cardiovascular events (top axis) and individuals (bottom axis). This relationship suggests increased cardiovascular risk at both low and high levels.

Table.

Partial literature summary of vitamin D-hormone actions

Vitamin D-hormone effects	Targets
Cells that metabolize D3	• Hepatocytes, ⁶⁶ renal cells, ⁶⁷ endothelial cells, ⁶⁸ smooth muscle cells, ⁶⁹ monocytes, ⁷⁰ skeletal muscle cells ⁷¹
Pathophysiology affected	• Osteomalacia, ⁷² vascular calcification, ^{56, 57, 73, 28} renal and myocardial calcification, ⁷³ atherosclerosis, ⁷⁴ heart failure, ⁷⁵ thrombosis ⁷⁶
Tissue content and function affected	• Calcium balance, ⁷⁷ aortic elastin, ⁷⁸ bile acid synthesis, ⁷⁹ vasoconstrictor response to norepinephrine, ⁸⁰ lipogenesis, ⁸¹ insulin sensitivity, ⁸² endothelial-dependent contraction ⁸³
Cellular functions	• Osteoblastogenesis, ⁸⁴ osteoclastogenesis, ⁸⁵ chondrogenesis ⁸⁶ • Myogenesis, ⁸⁷ adipogenesis, ⁸⁸ hematopoiesis, ⁸⁹ nerve growth, ⁹⁰ • Smooth muscle cell (SMC) dedifferentiation, ⁹¹ SMC migration, ⁹² SMC contraction, ⁹³ • Tumor cell differentiation ⁹⁴ • Macrophage cholesterol uptake, ⁹⁵ T-lymphocytes ⁹⁶ • Apoptosis, ⁹⁷ DNA synthesis, ⁹⁸ arachidonic acid turnover ⁹⁹ • Oscillation of inositol phosphate 3 and diacylglycerol production ¹⁰⁰ • Prostaglandin production, ^{101, 102} superoxide anion production ¹⁰³ • Nitric oxide synthase ¹⁰⁴
Signaling pathways	• Protein kinase C-alpha, ¹⁰⁵ cyclic AMP, ¹⁰⁶ p38 MAPK, ¹⁰⁷ c-myc, ¹⁰⁸ c-fos, ⁸⁵ NFAT1, ¹⁰⁹ Wnt, ¹¹⁰ nuclear factor-kappa B ¹¹¹
Known interactions	• Retinoic acid receptors, ¹¹² retinoid X receptors, ¹¹³ glucocorticoid, ¹¹⁴ • Runx2, ¹¹⁵ transforming growth factor-beta, ¹¹⁶ insulin-like growth factor binding protein-5, ¹¹⁷ vitamin K, ¹¹⁸ estrogen, ¹¹⁹
Protein synthesis and serum levels regulated	• Low-density lipoprotein (LDL), ¹²⁰ prostaglandin synthesis, ¹⁰¹ endothelin receptor ⁹¹
Gene expression	• Bone morphogenetic protein -2, ¹²¹ alkaline phosphatase, ¹²² bone sialoprotein, ¹²³ collagen I, ¹²⁴ osteopontin, ¹²⁴ osteocalcin ¹¹⁵ tissue plasminogen activator, ¹²⁵ parathyroid hormone, ¹²⁶ fibroblast growth factor-23 ¹²⁷ • Receptor activator of nuclear factor kappa B ligand ¹²⁸ • Muscle segment homeobox-containing gene Msx-2 (Hox-8), ¹²⁹ • Peroxisome proliferator activated receptor-gamma ¹³⁰ • Integrins, ¹³¹ fibronectin, ¹³² laminin receptor ¹³³ • Collagenase, ¹³⁴ matrix metalloproteinase ¹³⁵ • Granulocyte-macrophage colony stimulating factor, ^{109, 114} macrophage-colony stimulating factor ¹³⁶ tumor necrosis factor-alpha, ¹³⁷ interleukin 8 ¹³⁸ • Insulin receptor ¹³⁹ • Vascular endothelial growth factor, ¹⁴⁰ nerve growth factor, ¹⁴¹ • Type A natriuretic peptide receptor ¹⁴² • Aromatase ¹⁴³ • Nephren ¹¹² • CYP7A1, ¹⁴⁴ CYP19A1, ¹⁴³ CYP24A1 ¹⁴⁵ • p450 cytochrome, ¹⁴⁶ Mullerian-inhibiting substance ¹⁴⁷