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¹⁸O composition of CO₂ and H₂O ecosystem pools and fluxes in a tallgrass prairie: Simulations and comparisons to measurements

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Abstract

In this paper we describe measurements and modeling of ¹⁸O in CO₂ and H₂O pools and fluxes at a tallgrass prairie site in Oklahoma. We present measurements of the δ^{18} O value of leaf water, depth-resolved soil water, atmospheric water vapor, and Keeling plot δ^{18} O intercepts for net soil-surface CO₂ and ecosystem CO₂ and H₂O fluxes during three periods of the 2000 growing season. Daytime discrimination against C¹⁸OO, as calculated from measured above-canopy CO₂ and δ^{18} O gradients, is also presented. To interpret the isotope measurements, we applied an integrated land-surface and isotope model (ISOLSM) that simulates ecosystem H₂¹⁸O and C¹⁸OO stocks and fluxes. ISOLSM accurately predicted the measured isotopic composition of ecosystem water pools and the δ^{18} O value of net ecosystem CO₂ and H₂O fluxes. Simulations indicate that incomplete equilibration between CO₂ and H₂O within C₄ plant leaves can have a substantial impact on ecosystem discrimination. Diurnal variations in the δ^{18} O value of above-canopy vapor had a small impact on the predicted δ^{18} O value of ecosystem water pools, although sustained differences had a large impact. Diurnal variations in the δ^{18} O value of above-canopy CO₂ substantially affected the predicted ecosystem discrimination. Leaves dominate the ecosystem ¹⁸O-isoflux in CO₂ during the growing season, while the soil contribution is relatively small and less variable. However, interpreting daytime measurements of ecosystem C¹⁸OO fluxes requires accurate predictions of both soil and leaf ¹⁸O-isofluxes.

Nomenclature

 $C_i = \text{leaf internal CO}_2 \text{ concentration } (\mu \text{mol m}^{-3})$

 $C_{\rm H}$, $C_{\rm L}$ = CO₂ concentrations at the high and low sampling points (µmol m⁻³)

 F_{al} , $F_{la} = CO_2$ fluxes into and out of the leaf (μ mol m⁻²s⁻¹)

 F_{g_r} , F_{sm_r} , F_s = growth, stem, and net soil-surface CO₂ fluxes (µmol m⁻² s⁻¹) $I = \text{ecosystem}^{18} \text{O-isoflux} (\mu \text{mol } \text{m}^{-2} \text{s}^{-1} \text{\%})$

 R_{a} , R_{c} , $R_{eq} = CO_2^{-18}O$ isotopic ratios in the atmosphere, chloroplast, net ecosystem flux, and in equilibrium with leaf water

 $z_0 = e$ -folding depth for soil CO₂ source (m)

 δ_a , δ_e , δ_l , δ_{H} , δ_L , δ_{nl} , δ_{sw} , $\delta_s = \delta^{18}$ O values of atmospheric CO₂, net ecosystem CO₂ flux, leaf water; CO₂ at the high sampling point, CO₂ at the low sampling point, net leaf CO₂

- flux, stem respiration CO₂ flux, and soil CO₂ flux (‰)
- Δ_c = difference in δ^{18} O values of chloroplast and atmospheric CO₂ (‰)
- Δ_{eq} = difference in δ^{18} O values of equilibrium chloroplast and atmospheric CO₂ (‰)

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 Δ_L^{18} , Δ_s^{18} , Δ_e^{18} = net discrimination against C¹⁸OO from leaves, soil, and ecosystem (‰) Θ_{eq} = disequilibrium between CO₂ and leaf water

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Introduction

Terrestrial ecosystem carbon fluxes dominate largescale spatial and temporal variations in the δ^{18} O value of atmospheric CO₂, δ_a (‰). Ocean gas exchange, biomass and fossil fuel burning, and stratospheric reactions also impact δ_{a} , although their influence is much smaller (Francey & Tans, 1987; Farquhar et al., 1993; Ciais et al., 1997a; Peylin et al., 1999). At the global scale, a simple mass balance can be used to estimate the relative impact of these fluxes. The net ocean exchange, fossil fuel and biomass burning, and stratospheric fluxes are each less than 150 Pg C yr^{-1} %, while the photosynthetic and soil respiratory fluxes are each on the order of $1000 \text{ Pg C yr}^{-1}$ % (Tans, 1980; Farquhar et al., 1993). Using a global network of flask data, Francey & Tans (1987) inferred that terrestrial carbon fluxes interacting with soil and leaf water drive the large (~1.5‰) observed interhemispheric gradient in δ_a . Simultaneously, Friedli *et al.* (1987) showed that terrestrial fluxes strongly influence measured variations in δ_a above Switzerland.

Experimental and theoretical advances in our understanding of respiratory and photosynthetic influences on δ_a followed these studies. In both respiration and photosynthesis, the exchange of oxygen isotopes between water and dissolved CO₂ imparts the δ^{18} O value of water to the CO2 molecules. Hesterburg & Siegenthaler (1991) first investigated the interactions of soil CO₂ with soil water and developed a model to predict the δ^{18} O value of soil-respired CO₂. Tans (1998) expanded on this model to incorporate the influence of 'abiotic' or 'invasion' fluxes and developed analytical solutions for the δ^{18} O value of soil-gas CO₂ and surface CO₂ fluxes for a range of environmental conditions. Miller et al. (1999) performed a series of detailed measurements to quantify the effective kinetic fractionation of CO2 diffusion out of the soil and demonstrated the importance of the invasion flux in soil chamber measurements. By applying the analytical solutions of Tans (1998), Stern et al. (2001) estimated the importance of abiotic CO₂ exchange between the atmosphere and soil for a variety of ecosystems. To quantify leaf C¹⁸OO exchanges, Farquhar & Lloyd (1993) developed equations describing interactions between leaf CO₂ fluxes and ¹⁸O in leaf water. Leaf $C^{18}OO$ exchanges are dominated by the influx of CO_2 molecules into leaves, isotopic exchange with leaf water, and 'retrodiffusion' (i.e., molecules leaving the leaf without being fixed by photosynthesis, but having a different δ^{18} O value than when they entered).

Some of these results have been incorporated in global modeling studies of surface C¹⁸OO fluxes and δ_a . In the first such study, Farquhar et al. (1993) confirmed the importance of photosynthesis and respiration on δ_a . Ciais et al. (1997a, b) modeled net ecosystem C¹⁸OO fluxes and coupled them to a 3-D tracer-transport model to examine seasonal and latitudinal variations in δ_a . Using the same modeling framework, Peylin *et al.* (1999) explored the influence of specific geographic regions on temporal and latitudinal variations in δ_a . They argued that seasonal variations in northern hemisphere δ_a are dominated by carbon exchanges in the Siberian taiga region, where soil and leaf water are strongly depleted in ¹⁸O. Despite these modeling advances, a number of uncertainties remain, including the large interannual variability and apparent downward trend in global measurements of δ_a (Gillon & Yakir, 2001; Stern et al., 2001). Furthermore, the models of ecosystem C¹⁸OO fluxes used in these global-scale studies have not been adequately tested at the site level.

Several ecosystem-scale studies have reported using measurements of ¹⁸O in H₂O and CO₂ to better understand controls on the δ^{18} O value of CO₂ exchanged with the atmosphere or to partition measured net CO₂ fluxes into gross fluxes (for reviews, see Yakir & Sternberg, 2000; Dawson et al., 2002). Yakir & Wang (1996) used simultaneous measurements of ¹⁸O and ¹³C to partition net CO₂ fluxes between soil and plant components in several crop fields. Flanagan et al. (1995) described measurements of ¹⁸O in water pools and CO₂ at two heights in a pine forest canopy. They reported that predictions from a mixing model that included the various ecosystem CO₂ sources were consistent with observed diurnal variations in C¹⁸OO concentrations within the canopy. Using discrimination calculations based on measurements of ¹⁸O in soil, stem, and leaf water, Flanagan et al. (1997) estimated that mid-day discrimination against C18OO during photosynthesis was similar among the three boreal forest types, but that CO₂ released during soil respiration was less depleted than expected due to interactions with an extensive moss layer. In a tropical forest site, Harwood et al. (1998) reported leaf-level measurements of the

 δ^{18} O value of CO₂ fluxes and their relationship to vapor pressure deficits and stomatal conductance. Harwood *et al.* (1999) used Keeling plot analysis in a UK forest site to demonstrate that heterogeneity in ecosystem water pools likely accounted for variations in the δ^{18} O value of CO₂ fluxes. Keeling plots and measurements of ¹⁸O in ecosystem water pools have also been used to quantify soil and leaf contributions to the net ecosystem latent heat flux (Wang & Yakir, 2000).

The studies mentioned above were conducted in forest and agricultural ecosystems; much less information exists on $^{18}\!O$ exchanges in H_2O and CO_2 in grasslands, which cover close to 25% of the earth's land surface. Several recent studies indicate that C¹⁸OO exchanges may be more complicated than originally thought, particularly in C₄ grasslands. For example, incomplete isotopic equilibration (termed disequilibrium, Θ_{eq}) between CO₂ and leaf water due to low carbonic anhydrase (CA) enzyme activity may be common in most grasses (Gillon & Yakir, 2000, 2001). In this notation, Θ_{eq} varies between 0 and 1, and $\Theta_{eq} = 1$ indicates complete equilibration. Gillon & Yakir (2001) report an average leaf disequilibrium in C_4 plants, excluding outliers, of 0.38, while Helliker (2001) estimated Θ_{eq} to range from 0.6 to 0.7 in both leaf-level and whole-plant experiments with the C4 grass Andropogon gerardii. Additionally, large variations in leaf water isotopic composition along the leaf length were reported by Helliker & Ehleringer (2000), further complicating predictions of photosynthetic C¹⁸OO exchanges in grasslands.

In this paper, we describe a set of isotopic measurements and simulations in a C₄-dominated tallgrass prairie ecosystem located in north-central Oklahoma, USA. After describing the experimental methods and measurements, we apply a mechanistic land-surface and isotope model (ISOLSM; Riley *et al.*, 2002) to interpret the physiological and biophysical controls on measured δ^{18} O values of CO₂ and H₂O pools and fluxes. We also demonstrate the importance of incomplete equilibration between leaf water and CO₂ on ecosystem C¹⁸OO fluxes.

Materials and methods

Site description

Measurements were made in a tallgrass prairie pasture in Oklahoma, USA (36°56′N, 96°41′W) during three sampling campaigns in 2000: May 6–10, June 4–7, and July 18–21. The site is in a region with various land uses and covers, including crops, sparse trees, and other grasslands; it has not been grazed since 1996, but is burned every spring. The maximum leaf area index (LAI) is about 3.0 and the maximum net ecosystem exchange during the growing season is about $35 \,\mu\text{mol}\,\text{m}^{-2}\,\text{s}^{-1}$. The site and collection of meteorological forcing and flux data are described in detail in Suyker & Verma (2001).

 C_4 grasses constituted at least 78% of the species present in a vegetation cover survey conducted in 1997 (Suyker & Verma, 2001). However, this proportion varies seasonally. Still *et al.* (2003a) used night-time ¹³C measurements to estimate the fraction of ecosystem respiration resulting from C_4 -derived carbon. In 2000, these measurements indicated that the C_4 contribution increased from 67% in early spring to 77% in midsummer. Daytime measurements of above-canopy ¹³C gradients showed a similar seasonal increase in the fraction of net ecosystem exchange attributable to C_4 plants.

CO₂ measurements

Night-time CO₂ Keeling plots

We collected air samples at four heights above the surface (0.5, 1.5, 2.5, and 4.5 m) several times during the night when both plant and heterotrophic respiration were occurring. The 0.5 m sampling level was directly above the grassland canopy. Each air sample was collected over a 2-min interval, so that all four levels were sampled in less than about 10 min. Each set of four samples was used to create a single Keeling plot intercept. Further details of field sampling are described in Still et al. (2003a). Air samples were collected and stored in 100 ml glass flasks (Kontes Custom Glass Shop, Hayward, CA, USA) and returned to the Carnegie Institution of Washington for concentration and isotopic analysis. This analysis was conducted using a system that simultaneously measures CO₂ mixing and isotope ratios in small air samples (Ribas-Carbo et al., 2002). The measurement precision for CO₂ concentration with this system is 0.4-0.7 ppm, and for δ^{18} O values of CO₂ is 0.05‰.

Using the night-time data, we constructed mixing plots (Keeling plots) to estimate the isotopic composition of the net ecosystem CO₂ flux, δ_{e} (‰). This approach (Keeling, 1958; Keeling, 1961) assumes that the system can be represented as a simple mixture of atmospheric CO₂ and the CO₂ produced by ecosystem respiration. The intercept from a linear regression between the δ^{18} O value of the CO₂ concentration and the reciprocal of the CO₂ concentration provides an estimate of the isotopic composition of the respiration source. We used a geometric mean regression that incorporates errors in both the concentration and isotopic composition measurements to calculate the intercept (Friedli *et al.*, 1987; Sokal & Rohlf, 1995;

Flanagan *et al.*, 1996; Bowling *et al.*, 1999; Harwood *et al.*, 1999; Bowling *et al.*, 2001), while the standard error of the regression intercept was calculated with a Model I formula (Sokal & Rohlf, 1995; Laws, 1997). We report isotopic values in per mil (∞), with CO₂ isotopic concentration and flux ratios calculated relative to the standard Vienna Pee Dee belemnite (V–PDB-CO₂), and H₂O isotopic concentration and flux ratios calculated relative to Vienna-Standard Mean Ocean Water (V–SMOW).

Daytime discrimination

We sampled daytime CO₂ concentration and δ^{18} O values at the highest (4.5 m) and lowest (0.5 m) tower levels during the 2000 growing-season. The sample collection was the same as that used for night-time collections, except that only two levels were sampled for each measurement.

The average measured CO_2 concentrations and isotopic ratios sampled at each height during a one to three hour period were used for each discrimination calculation. On a typical day, fewer than 10 gradient pairs with sufficient concentration and isotope gradients could be sampled due to strong winds at the site. The net ecosystem discrimination against $C^{18}OO$, Δ_e^{18} (‰), was calculated as (derived in Still *et al.*, 2003a); see also Evans *et al.* (1986); Lloyd & Farquhar (1996):

$$\Delta_{\rm e}^{18} = \frac{-1000C_{\rm H}(\delta_{\rm H} - \delta_{\rm L})}{1000(C_{\rm H} - C_{\rm L}) + (\delta_{\rm H}C_{\rm H} - \delta_{\rm L}C_{\rm L})}, \qquad (1)$$

where $C_{\rm H}$ and $C_{\rm L}$ (µmol m⁻³) are the CO₂ concentrations at the high and low sampling points, respectively, and $\delta_{\rm H}$ and $\delta_{\rm L}$ (‰) are the δ^{18} O values of CO₂ at the high and low sampling points, respectively.

Soil-surface CO₂ flux

We measured the δ^{18} O value of the soil-surface CO₂ flux on two occasions in July 2000. The measurement protocol was designed to minimize pressure artifacts (Lund et al., 1999) and to average over the small-scale variation in C₃ and C₄ abundance. We used a modified Licor 6400 soil respiration system (Still et al., 2003a). A balloon was placed on the inside of the chamber over the pressure equalization port and a septum was attached so that air could be withdrawn from the chamber through a small column of magnesium perchlorate into pre-evacuated 100 mL flasks. When a sample was taken, atmospheric air flowed into the balloon, equalizing the pressure inside and outside the chamber. The chamber was placed on open patches of soil between grass tussocks. Before sampling, respired CO₂ was allowed to accumulate in the chamber for a time (the same for each replication) sufficient to reach a concentration of approximately 700 ppm. Seven flasks

from the chamber were taken, one each from seven separate soil patches. Two flasks were also filled with ambient air (one at the beginning and one at the end of the sampling sequence). The data were combined for Keeling plot analysis (geometric mean regression, nine points per curve) to obtain a flux-weighted average of the isotopic composition of CO_2 respired from the sampled surface area.

Using the method described in Miller *et al.* (1999), we calculated the impact of invasion (excess chamber CO_2 diffusing into the soil, equilibrating with soil water, and diffusing back into the chamber) on the soil-surface isoflux to be small (less than 0.5%), primarily because soil moisture in the top 15 cm was low at the time of this measurement (less than 25% water-filled pore space).

H₂O measurements

Atmospheric water vapor

Atmospheric water vapor samples were collected and analyzed for ¹⁸O composition. The water vapor mixing ratio was determined by averaging the micrometeorological measurements at each height over the sampling period. For ¹⁸O composition, water vapor was collected from four heights so that each Keeling plot consisted of four samples. The sampling methodology is described in detail in Helliker et al. (2002). Briefly, atmospheric air was pulled through a dry-ice-cooled glass condenser for 15 min. The samples were then sealed and shipped to the SIRFER laboratory at the University of Utah (http://ecophys.biology.utah.edu/ sirfer.html). Pure CO₂ was injected into condensers (to a final concentration of 10%) and allowed to equilibrate for 48 hours. The δ^{18} O value was determined by oncolumn injection through a continuous flow isotope ratio mass spectrometer. Keeling plots for water vapor were calculated analogously to those for CO₂.

Leaf and soil water

Soil water, bulk leaf water, and plant stem water were collected and analyzed for ¹⁸O composition. Soil water was sampled with a soil corer to 20 cm depth. We subsampled each core in four intervals: 0–2.5, 2.5–5, 5–10, and 10–20 cm. These soil plugs were placed in glass vials, sealed with parafilm, and stored for isotopic analysis. The leaf water isotopic composition was determined by collecting upper leaves from the dominant plants (C₃: *Solidago missouriensis*; C₄: *Andropogon gerardii, Panicum virgatum, and Sorghastrum nutens*) at mid-day and storing them in glass vials as with the soil samples.

The leaf and soil samples were analyzed at SIRFER at the University of Utah. Water was extracted from the sample with cryogenic distillation and its ¹⁸O composition was determined by the CO_2 -H₂O equilibration method of Socki *et al.* (1992) with modifications described in Helliker & Ehleringer (2000).

ISOLSM simulations

ISOLSM couples the land-surface model LSM1.0 (Bonan, 1996) with modules that predict the ¹⁸O composition of canopy water vapor, leaf water, and vertically resolved soil water; CO₂ oxygen isotope exchanges with soil and leaf water; leaf photosynthetic C¹⁸OO fluxes; soil CO₂ and C¹⁸OO diffusive fluxes (including abiotic soil exchange); and ecosystem exchange of H₂¹⁸O and C¹⁸OO with the atmosphere. A detailed description of the processes simulated in ISOLSM can be found in Riley et al. (2002). Briefly, the depth-resolved ¹⁸O isotopic ratio of soil water is solved by an explicit numerical solution to the H₂¹⁸O mass balance. The model accounts for equilibrium fractionation during evaporation and fractionation through the laminar layer at the soil surface. Rain and irrigation inputs are treated as boundary conditions on the soil water ¹⁸O balance. A semi-implicit numerical scheme is used to simulate the soil-gas diffusion and CO₂ interactions with ¹⁸O in the soil water. For the simulations shown here, the model calculates the energy balance, photosynthetic CO₂ flux, soil moisture and temperature, and the δ^{18} O value of soil moisture with a time step of 100 s, while the soil-gas diffusion model uses a time step of 20 s. Predictions of the δ^{18} O value of leaf water, δ_1 (‰), are based on the predicted stem water and canopy water vapor isotopic composition and the Craig-Gordon model (Craig & Gordon, 1965) with modifications for leaves as described by Flanagan et al. (1991). The canopy water vapor ¹⁸O composition is determined diagnostically based on fluxes from the atmosphere, leaves, and the soil surface. As mentioned earlier, incomplete equilibration between leaf water and CO₂ can occur in C₄ grass leaves as a result of low CA activity. Gillon & Yakir (2000) showed that a reasonable approximation for leaf disequilibrium is $\Theta_{eq} = \Delta_c / \Delta_{eq}$, where $\Delta_{\rm c} = (R_{\rm c}/R_{\rm a} - 1)1000$, $\Delta_{\rm eq} = (R_{\rm eq}/R_{\rm a} - 1)1000$, and R_{a} , R_{c} , and R_{eq} represent the isotopic ratios of the background atmosphere, chloroplast CO₂, and CO₂ in equilibrium with leaf water, respectively. We apply these relationships to calculate an equivalent δ^{18} O value of leaf internal CO2 that reflects the reduced leaf discrimination due to disequilibrium.

We used measurements from the site to specify soil characteristics and the vertical root distribution. ISOLSM is forced with measurements of air temperature, wind speed, CO_2 concentration, vapor pressure, downward short-wave and long-wave radiation, precipitation or irrigation amount and its isotopic ratio,

and the δ^{18} O value of above-canopy vapor and CO₂. We conducted the simulations presented here assuming a 100% C_4 canopy, as C_4 plants dominate the net ecosystem exchange during the growing season. Since we did not have continuous measurements of the isotopic composition of atmospheric vapor, we assumed a value 7‰ less than the predicted stem water isotopic composition. Actual values are impacted by many factors other than evapotranspiration (e.g., horizontal and vertical atmospheric advection), and show diurnal variations of up to 4‰ (Helliker *et al.*, 2002). We also lacked measurements of the isotopic composition of above-canopy CO₂ and assumed a constant value of 0‰ for the results shown here, although diurnal variations of up to 2‰ are common (Helliker et al., 2002). We test the sensitivity of model predictions to variations in the δ^{18} O values of atmospheric CO₂ and H₂O below.

Results and discussion

In this section, we present experimental results and discuss comparisons between measured and simulated latent heat, sensible heat, and CO_2 fluxes; the ¹⁸O composition of water in leaves and stems; vertically resolved soil water ¹⁸O composition; and H₂¹⁸O and C¹⁸OO exchanges with the atmosphere. We also apply ISOLSM to examine biological and physical controls on these exchanges.

Ecosystem CO₂ and latent and sensible heat fluxes

Model predictions of ecosystem CO_2 and latent and sensible heat fluxes impact predictions of ecosystem $H_2^{18}O$ and $C^{18}OO$ stocks and fluxes. For example, sustained inaccuracy in the predicted ecosystem latent heat flux can result in inaccurate predictions of nearsurface soil water isotopic composition. Such errors can propagate to impact stem and leaf water isotopic composition predictions and therefore the ecosystem exchanges of $C^{18}OO$. The model accurately simulated the magnitude and dynamics of CO_2 and latent and sensible heat fluxes over the growing season. For example, Fig 1(a) and (b) show comparisons between measured and modeled fluxes over a 10-day period beginning May 1, 2000.

Isotopic composition of ecosystem water pools

Figure 2 shows the predicted and measured depthresolved soil water isotopic composition over the 3month period. We initialized the modeled soil water isotopic composition using measurements taken at the beginning of May 2000. The model reasonably predicts

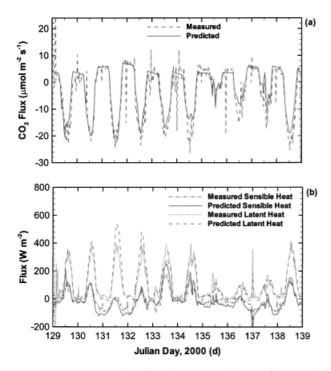


Fig. 1 Measured and predicted ecosystem (a) CO_2 fluxes and (b) latent and sensible heat fluxes for a 10-day period in May 2000. Spikes in the measured night-time CO_2 fluxes are likely due to transient perturbations in the boundary layer.

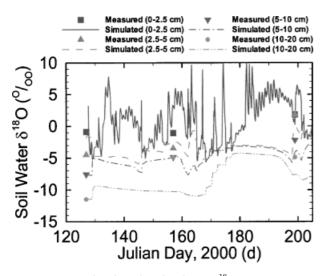


Fig. 2 Measured and predicted soil water ¹⁸O isotopic composition over four depth intervals. The top soil layer δ^{18} O value responds rapidly to surface evaporation and precipitation. δ^{18} O values are relative to the V-SMOW standard.

the δ^{18} O value of soil water measured in the subsequent periods. The 10–20 cm soil layer water isotopic composition is predicted to be ~5 and 3‰ too light in the June and July measurement periods, respectively, while the soil water isotopic composition between 2.5 and 5 cm is

predicted to be ~2‰ too light in the July measurement period. The predictions are dependent on the assumed rooting activity depth profile and the δ^{18} O value of atmospheric water vapor, as described in Riley *et al.* (2002). The impact of errors in the rooting activity is difficult to evaluate since the activity profile at this site is not known. Errors in the imposed atmospheric water vapor δ^{18} O value could easily result in an error in the predicted near-surface soil water isotopic composition of 2‰ after 3 months of simulation.

The temporal resolution of the measurements shown in Fig. 2 does not allow us to conclude that predicted diurnal variations in soil water isotopic composition between 0 and 2.5 cm, which are as large as 5‰, are accurate. However, we observed comparable diurnal variability in measurements of the δ^{18} O value of nearsurface soil water taken in the summer of 2001 in a sorghum field in Oklahoma (data not shown). These variations are driven by soil evaporation, and are largest when evaporation is large, e.g., following precipitation. Nonetheless, more diurnal measurements of soil water ¹⁸O composition are required to test these model predictions adequately. Accurately predicting the near-surface soil water isotopic composition is important since it influences the isotopic composition of the net soil-surface CO₂ and H₂O fluxes.

Predicted and measured leaf and stem water isotopic compositions for the dominant C₄ grasses are shown in Fig. 3 for the 3 days in May, June, and July for which we have measurements. δ_1 and the δ^{18} O value of stem water are accurately predicted in May and July, while δ_1 is predicted to be $\sim 3\%$ too heavy in the June sampling. The predicted δ_1 typically increases from early morning to mid-afternoon by between 10‰ and 20‰, and then relaxes back to a night-time steady value. We have observed similar patterns and magnitudes of diurnal variation in sorghum leaf water ¹⁸O composition in this area (data not shown). The leaf water isotopic composition is substantially more variable than that of soil water (particularly deeper than 5 cm; Fig. 2) over the afternoon, resulting in more variable δ^{18} O values of leaf CO₂ fluxes than of soil CO₂ fluxes.

Ecosystem H₂¹⁸O fluxes

The predicted and measured evapotranspiration (ET) flux isotopic composition is shown in Fig. 4 for the 3 days when H₂O Keeling plots were available. The predicted ecosystem water flux isotopic composition shows strong diurnal variation, consistent with the variable soil evaporation isotopic composition and fraction of total ET attributable to soil evaporation. The δ^{18} O value of the net ecosystem vapor flux will be more variable the larger the contribution of soil

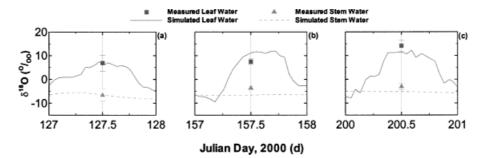
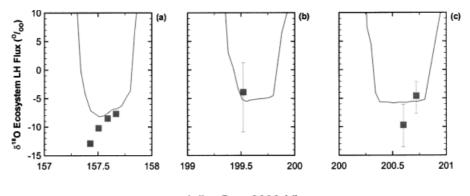


Fig. 3 Measured and predicted leaf and stem water ¹⁸O isotopic composition. The predicted stem water ¹⁸O composition represents a convolution over depth of rooting activity and soil water ¹⁸O composition. Leaf water δ^{18} O values depend on leaf temperature, δ^{18} O value of stem water, and canopy airspace vapor concentration and δ^{18} O value. δ^{18} O values are relative to the V-SMOW standard.



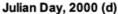


Fig. 4 Measured and predicted ¹⁸O isotopic composition of ecosystem latent heat flux (soil evaporation and transpiration). For figure clarity, we omitted the 95% confidence intervals on (a) which, from left to right are: (2.5, -27.2), (2.6, -22.7), (3.1, -19.9), and (0.5, -15.5). When transpiration dominates the latent heat flux, the ecosystem isotopic composition more closely matches the stem water ¹⁸O composition. As the soil evaporative flux becomes a larger fraction of the ecosystem latent heat flux, the ecosystem signal becomes more variable. δ^{18} O values are relative to the V-SMOW standard.

evaporation to the total ET flux. δ^{18} O values of canopy and above-canopy atmospheric vapor and soil-surface evaporation and transpiration are shown for a typical day in Fig. 5. The soil evaporation isotopic composition is typically lightest at about 11 in the morning and increases throughout the day. This increase is caused by the concurrently increasing near-surface soil water isotopic composition and soil-surface equilibrium-partitioning coefficient (due to increased soil temperature). The perturbations in canopy water vapor and soil evaporation δ^{18} O values at about day 180.6 occur because of a rapid change in measured downward short-wave radiation over this period, probably caused by clouds. At any particular time, the canopy water vapor isotopic composition depends on H218O and H_2O fluxes between the canopy air and (1) overlying atmosphere, (2) leaves, and (3) soil surface (see Eqn (11) of Riley et al. (2002)). Typically, the predicted canopy air space vapor isotopic composition in the morning is close to that of the background above-canopy vapor, increases until about mid-day as a result of the heavier transpiration flux, and then returns to the background value as the ecosystem latent heat flux diminishes in the late afternoon.

Soil respiration C¹⁸OO fluxes

The measured and predicted δ^{18} O values of soilrespired CO₂ are presented in Fig. 6. In these simulations, we imposed soil respiration profiles that decay exponentially with depth. We assume that the root respiration profile follows the root distribution profile with an *e*-folding depth, z_0 (m), of 0.2 m, and that the microbial respiration has a profile defined by $z_0 = 0.05$ m. Thus, about 75% of the predicted soil respiration occurs in the top 15 cm of soil. In reality, the depth distribution of respiration will depend on the distribution of root activity, soil moisture and

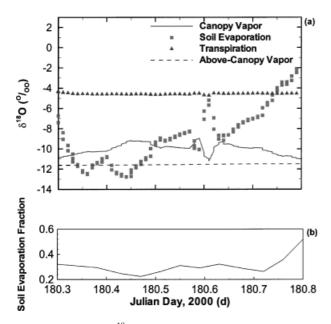


Fig. 5 (a) Predicted ¹⁸O isotopic composition of canopy vapor, soil evaporation, transpiration, and above-canopy vapor for a single daytime period. The canopy vapor isotopic composition increases during the day due to the heavier transpiration flux, and then returns to the above-canopy composition as the ecosystem latent heat flux diminishes. (b) Fraction of evapotranspiration due to soil evaporation. δ^{18} O values are relative to the V-SMOW standard.

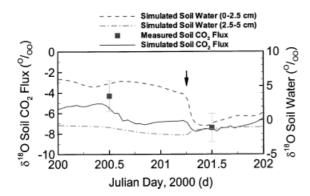


Fig. 6 Measured and predicted ¹⁸O isotopic composition of the net soil-surface CO₂ flux (left ordinate) and predicted δ^{18} O value of soil water between 0–2.5 cm and 2.5–5 cm (right ordinate). The arrow indicates a small precipitation event with a δ^{18} O value of – 2.6‰. Predicted and measured δ^{18} O values of the net soil-surface CO₂ flux reflect the relatively lighter soil water in the 0–2.5 cm layer after the precipitation event. Soil water δ^{18} O values are relative to the V-SMOW standard; CO₂ flux δ^{18} O values are relative to the V-PDB-CO₂ standard.

temperature, and microbial substrate availability, among others. The δ^{18} O value of the net soil-surface CO₂ flux is sensitive to the depth distribution of CO₂ production. For example, if a larger fraction of respiration occurs near the surface, the δ^{18} O value of the net soil-surface

CO₂ flux will become more enriched as a result of equilibration with more enriched near-surface soil water.

The decrease in the measured δ^{18} O value of soilrespired CO₂ between days 200 and 201 is caused by a small precipitation event with a δ^{18} O value of -2.6%(indicated by the arrow in Fig. 6). This precipitation input is reflected in the simulated δ^{18} O value of soil water as a decline in the top soil layer and as a small increase in the second soil layer. The small increase in the δ^{18} O value of soil water in the 2.5–5 cm layer results from advection of the heavier soil water (before the precipitation event) from 0-2.5 cm. As the CO₂ diffuses through the top soil layer, it interacts with this lighter soil water and thereby acquires a lighter isotopic composition. Note that this result contradicts the observation of Miller *et al.* (1999) that the δ^{18} O value of water in the top $5 \,\mathrm{cm}$ of soil does not substantially impact the δ^{18} O value of the soil-surface CO₂ flux. The decrease in the δ^{18} O value of soil-respired CO₂ resulting from the precipitation event is accurately captured by ISOLSM. In general, the impact of a precipitation event on the δ^{18} O value of soil-respired CO₂ depends on the precipitation flux, δ^{18} O value of precipitation and antecedent soil moisture, and the fraction of soil respiration occurring in the near-surface soil.

Ecosystem C¹⁸OO fluxes

Figures 7(a)–(c) show predicted and measured δ^{18} O values of the net ecosystem CO₂ flux (Keeling plot intercepts) for the three periods where measurements were made (predicted values are shown when the magnitude of the ecosystem CO₂ flux is greater than $2 \mu \text{mol m}^{-2} \text{s}^{-1}$). Three simulation scenarios are shown: the first assumes complete equilibration within the leaf ($\Theta_{\text{eq}} = 1.0$) and the second and third assume $\Theta_{\text{eq}} = 0.5$ and 0.35, respectively. In Fig. 7, only measurements with R^2 values greater than 0.6 and CO₂ gradients (between 0.5 and 4.5 m height) greater than 15 ppm are included (Table 1).

The δ^{18} O value of the predicted ecosystem CO₂ flux varied substantially during the day and between days due primarily to variations in δ_1 (also shown in Fig. 7) and leaf internal CO₂ concentration, C_i (µmol m⁻³). At night the predicted CO₂ flux isotopic composition remained relatively constant since the soil water isotopic composition and soil temperature are relatively constant. The sharp transitions in the δ^{18} O values of the net ecosystem CO₂ flux during the morning and evening occur when the system switches between being dominated by respiration and photosynthesis. During the day, photosynthetic uptake and retrodiffusion dominate ecosystem CO₂ and C¹⁸OO exchange. Lower

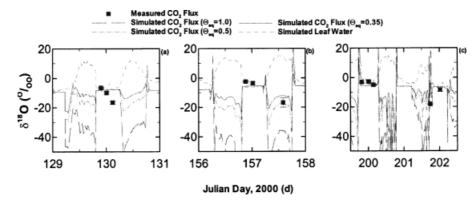


Fig. 7 Measured Keeling plot intercepts, predicted δ^{18} O value of the ecosystem CO₂ flux, and predicted δ_1 for five measurement periods. Simulated results are shown for three levels of leaf disequilibrium ($\Theta_{eq} = 1.0, 0.5, \text{ and } 0.35$). Note that a negative δ^{18} O value of the daytime ecosystem flux enriches the atmosphere. Leaf water δ^{18} O values are relative to the V-SMOW standard; CO₂ flux δ^{18} O values are relative to the V-PDB-CO₂ standard.

Table 1 R^2 values and differences in CO₂ concentrations between 0.5 and 4.5 m for the night-time CO₂ Keeling plot intercepts shown in Fig. 7

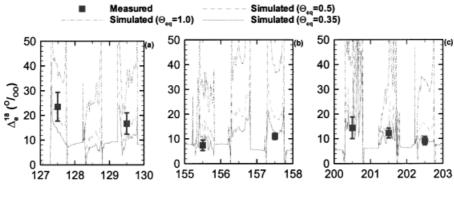
Date	Time (hours)	ΔCO_2 (ppm)	R^2
5/9/2000	21:37	65	0.99
5/10/2000	12:10	25	0.97
5/10/2000	2:50	22	0.99
6/5/2000	21:11	128	0.83
6/6/2000	12:23	118	0.62
7/18/2000	21:37	47	0.90
7/19/2000	12:20	118	0.98
7/19/2000	3:32	31	0.85
7/20/2000	21:47	16	0.72
7/21/2000	12:27	9	0.97

values of Θ_{eq} resulted in significantly more positive δ^{18} O values of daytime ecosystem CO₂ fluxes. Note that a negative δ^{18} O value of the daytime ecosystem CO₂ flux enriches the atmosphere.

Using a two-component mixing model (Keeling plot) to estimate the δ^{18} O value of respired CO₂ from measurements can be misleading, since (1) changes in the δ^{18} O value of ecosystem respiration can occur independently of changes in CO₂ concentration and (2) the water pools driving these isotopic changes can be spatially and temporally heterogeneous (Amundson *et al.*, 1998; Tans, 1998). As a result, extrapolation of the linear regression to infinite CO₂ concentration may not give an accurate estimate of the isotopic composition of respired CO₂. This decoupling of atmospheric CO₂ mole fraction and the isotopic composition of respired CO₂ can be caused by temporal changes in the leaf or soil water isotopic compositions over the measurement period. δ_1 is more likely to change over the night than

the δ^{18} O value of soil water (particularly below ~5 cm depth). It is unlikely that changes in leaf and soil water isotopic compositions affected our measurements at a particular time because each of our night-time estimates was based on measurements collected along a vertical spatial gradient over a short time interval. However, this effect can impact our interpretation of measurements throughout the night, as discussed below. The abiotic invasion of C¹⁸OO into soils and horizontal atmospheric advection can also impact the expected linear Keeling plot relationship. These potential confounders indicate that care must be taken when testing the accuracy of model predictions against δ^{18} O Keeling plot intercepts. Despite these concerns, the majority of our Keeling plots seem to follow the twocomponent model assumption, as demonstrated by the high R^2 values shown in Table 1.

The May 2000 ecosystem CO₂ flux isotopic compositions are notable since the Keeling plot intercepts tend toward more negative values throughout the night, contrary to model predictions. We believe that this measured trend is real. The CO₂ gradients in these Keeling plots (Table 1) are less than the threshold recommended by Pataki et al. (2003). However, that recommendation was based on the decrease in standard error of Keeling plot intercepts with increasing CO₂ gradients. The threshold they recommend (75 ppm) corresponds to a standard error of the intercept of 1‰; the standard errors of our three night-time intercepts are 0.2‰, 0.9‰, and 1.0‰, despite the relatively small CO_2 gradients. The inability of the model to capture this night-time trend may reflect (a) more rapid relaxation than is predicted by the model of leaf water isotopic composition back to the relatively depleted source water, thereby impacting leaf dark respiration; (b) an inaccurate prediction of night-time leaf stomatal



Julian Day, 2000 (d)

Fig. 8 Measured and predicted ecosystem discrimination for seven daytime periods. Predictions are shown for three levels of leaf disequilibrium ($\Theta_{eq} = 1.0, 0.5, \text{ and } 0.35$). Θ_{eq} values of 0.5 or 0.35 force model predictions closer to measured values (see text for caveats). From left to right, the number of flask pairs for each calculation is 5, 5, 5, 9, 3, 5, and 6. Discrimination values are relative to the V-PDB-CO₂ standard.

conductance; (c) changes in the isotopic composition of soil respiration not accurately predicted by the model; (d) incorrect prediction of invasion due to inaccurate estimates of the δ^{18} O value of net soil-surface CO₂ fluxes; or (e) advection (i.e., temporal heterogeneity in background atmospheric δ^{18} O values resulting from advection of air from adjacent areas). We do not have sufficient information to evaluate the relative impact of these mechanisms on the trend in these measurements.

Daytime discrimination

The measured (Eqn (1)) and predicted daytime ecosystem discrimination values (Δ_e^{18}) are shown in Figs 8(a)–(c). The predicted Δ_e^{18} is calculated as

$$\Delta_{\rm e}^{18} = \left(\frac{R_{\rm a}}{R_{\rm e}} - 1\right) 1000 = \frac{\delta_{\rm a} - \delta_{\rm e}}{1 + \delta_{\rm e}/1000},\tag{2}$$

where $R_{\rm e}$ (–) is the isotopic ratio of the net ecosystem CO₂ flux. Comparing predicted and measured ecosystem discrimination indicates that leaf disequilibrium may be substantial. In May, June, and July, Θ_{eq} values of 0.5, 0.35, and 0.35, respectively, force model predictions closer to measured values. Care must be taken in interpreting this observation since other modeled state variables (e.g., δ_1 and C_i) also have substantial impacts on Δ_{e}^{18} and may be inaccurately represented in the model. For example, our modeled δ_1 is higher than that observed in June, which will result in an overestimate of leaf discrimination. Further, our assumption of isotopic steady state in the leaf may result in an overestimate of δ_1 during transient environmental conditions. Errors in simulating C_i are potentially more troublesome due to the non-linear relationship between C_i and leaf discrimination against C¹⁸OO (Farquhar &

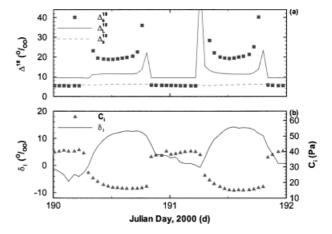


Fig. 9 Characteristic diurnal patterns of Δ_e^{18} . (a) Predicted ecosystem, soil, and leaf discrimination for $\Theta_{eq} = 0.5$. (b) Predicted δ_1 and C_i . Daytime ecosystem discrimination follows the pattern of leaf discrimination with a relatively constant offset due to soil discrimination. Discrimination values are relative to the V-PDB-CO₂ standard.

Lloyd, 1993). C₃ plants have higher C_i than C₄ plants and, as a result, larger leaf discrimination values. Therefore, the diminishing proportion of C₃ plants over the season may also cause the progression toward lower measured Δ_e^{18} ; this effect is not included in the modeling predictions shown here.

We examine more closely the factors contributing to the predicted Δ_e^{18} over the course of a single day in Figs 9(a) and (b). In Fig. 9(a), $\Theta_{eq} = 0.5$ and Δ_s^{18} and Δ_L^{18} (‰) represent the net discrimination of C¹⁸OO from soil and leaves, respectively, and are calculated analogously to Δ_e^{18} . The predicted δ_1 and C_i are shown in Fig. 9(b). The net ecosystem discrimination increases from ~5‰ before sunrise to about 20‰, and remains relatively constant until late in the afternoon. The abrupt shifts in the early morning and later afternoon occur as the system transitions between being dominated by photosynthesis and respiration; the net ecosystem isoflux is relatively small during these periods. Note that, although soil 'discrimination' is calculated to be positive, soil respiration typically makes the atmosphere lighter, in contrast to photosynthesis. Diurnal variations in Δ_s^{18} are small. With respect to impacts on Δ_L^{18} , the increase (decrease) in δ_l is opposed by the decrease (increase) in C_i during the morning (afternoon).

Ecosystem isoflux

The ecosystem ¹⁸O-isoflux, $I \pmod{m^2 s^{-1}}$, quantifies the net impact of ecosystem fluxes on the atmospheric C¹⁸OO content:

$$I = -(F_{\rm al} - F_{\rm la})\delta_{\rm nl} + (F_{\rm g} + F_{\rm sm})\delta_{\rm sw} + F_{\rm s}\delta_{\rm s}, \qquad (3)$$

where δ_{nl} , δ_{sw} , and δ_s (‰) are the predicted δ^{18} O values of the net leaf, stem respiration, and net soil-surface CO₂ fluxes, respectively; F_{al} and F_{la} (µmol m⁻²s⁻¹) are the predicted CO₂ fluxes into and out of the leaf, respectively; and F_g , F_{sm} , and F_s (µmol m⁻²s⁻¹) are the predicted growth respiration, stem respiration, and net soil-surface CO₂ fluxes, respectively. The three groups of terms on the right-hand side of Eqn (3) describe the net leaf isoflux, above-ground respiratory isoflux, and net soil-surface respiratory isoflux, respectively.

The net leaf isoflux dominates the ecosystem isoflux (Fig. 10), which varies substantially over the course of the day. The predicted photosynthetic CO_2 exchange and δ_1 each reach their maximum in the middle of the day, thereby reinforcing the net leaf isoflux magnitude. The net soil-surface isoflux is a small fraction of the

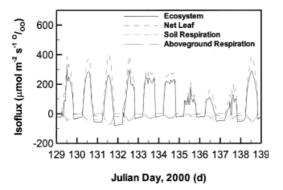


Fig. 10 Simulated net isotopic forcing (¹⁸O-isoflux) for $\Theta_{eq} = 0.5$. The daytime isoflux is dominated by the net leaf isoflux. δ^{18} O values are relative to the V-PDB-CO₂ standard.

ecosystem isoflux for several reasons. First, the net soilsurface CO₂ flux is a small fraction of the ecosystem CO₂ flux during this period. This tallgrass system maintains an approximately even carbon balance over the season through losses from soil microbial respiration and fire at other times of the year. In systems where diurnal respiratory and photosynthetic carbon fluxes are more balanced, the soil-surface isoflux will be a more significant contributor to the ecosystem isoflux. Second, the soil water isotopic composition at this site is not strongly depleted relative to the atmosphere (compared with, for example, boreal regions; Flanagan et al., 1997). Third, the mid-day leaf water is strongly enriched due to the low relative humidity and high δ^{18} O value of soil water, thus ensuring a large net leaf isoflux during the day. Finally, photosynthetic fluxes are much higher in productive grasslands than in many other ecosystems.

The relatively low soil moisture in this system leads to a lower effective diffusive fractionation and therefore a relatively enriched net soil-surface CO₂ flux. Concurrently, the lower soil moisture may cause a decrease in heterotrophic respiration, thereby reducing the soil isoflux. The net impact of these effects is difficult to discern without a model like ISOLSM that can explicitly simulate the impact of soil moisture and temperature on the CO₂ source strength and diffusivity as a function of depth. We will address these interactions and their impact on the δ^{18} O value of the soil-surface CO₂ flux in future work.

As mentioned earlier, C4 plants have lower discrimination against C¹⁸OO than C₃ plants. Thus, all else being equal, soil respiration has a relatively larger impact on the ecosystem isoflux in a C₄-dominated system. In a related analysis, we applied ISOLSM to investigate the impact of C₃ vs. C₄ plant physiology in this tallgrass system (Still et al., 2003b). In addition to the impact on leaf discrimination due to different CO₂ concentrations in the leaf stomatal pore, the C₃ and C₄ systems also have slightly different predicted soil moistures and leaf temperatures for the same meteorological forcing. Soil moisture and leaf temperature impact the soil-surface and leaf isofluxes. In these simulations, the soil isoflux was a larger proportion of the ecosystem isoflux in the C₄ system, due primarily to the reduced daytime leaf discrimination.

Diurnal variations in the δ^{18} O value of above-canopy CO_2

As part of our measurement campaigns in June and July, air samples from the 4.5 m high sampling port were collected in pre-dried and evacuated 100-mL glass flasks every 1.5–3 h and analyzed for their CO₂

concentration and oxygen isotope composition ($\delta_{\rm H}$). Several features are immediately apparent in the isotopic composition data (Figs 11(a) and (c)): an average diurnal cycle with an amplitude of ~1.5‰, an upward trend in the June period, and a downward trend in the July period. The diurnal cycle amplitude is large and roughly the same size as the interhemispheric $\delta_{\rm a}$ gradient observed by the NOAA-CMDL network (Farquhar *et al.*, 1993; Ciais *et al.*, 1997a, b). During both periods, diurnal variations in $\delta_{\rm H}$ are closely coupled to variations in CO₂ concentration. This tight coupling implies that changes in $\delta_{\rm H}$ are dominated by leaf and soil fluxes and not by changes in the δ^{18} O value of water pools independently of flux variations. There was no precipitation during these two periods.

An examination of modeled ecosystem ¹⁸O-isofluxes (Figs 11(b) and (d)) over these periods shows that the observed δ^{18} O diurnal cycle in both periods is strongly modulated by leaf photosynthetic ¹⁸O-isofluxes, with a small and relatively invariant contribution from soil ¹⁸O-isofluxes. The weak trends in observed $\delta_{\rm H}$ in June and July follow trends in the modeled net leaf ¹⁸O- isoflux. Unfortunately, a full comparison between modeled ¹⁸O-isofluxes and observed $\delta_{\rm H}$ variations is not possible, since $\delta_{\rm H}$ is also influenced by (1) advection of air parcels from adjacent regions, which may have different vegetation cover and soil water ¹⁸O composition, and (2) the atmospheric boundary layer height and turbulence conditions, which vary with surface heating and cooling. Coupling ISOLSM to a regional-scale meteorological model that simulates ¹⁸O exchanges with CO₂ and H₂O is in progress and will facilitate analyses that include these proceses.

Sensitivity to the $\delta^{18}O$ value of atmospheric CO_2 and H_2O

The simulations described above assume that the δ^{18} O value of atmospheric water vapor was 7‰ less than the predicted stem water isotopic composition, while the δ^{18} O value of atmospheric CO₂ was assumed to be constant at 0‰. Measurements at this site indicate that the δ^{18} O value of above-canopy H₂O and CO₂ can vary by 4‰ and 2‰, respectively, over the day (Helliker

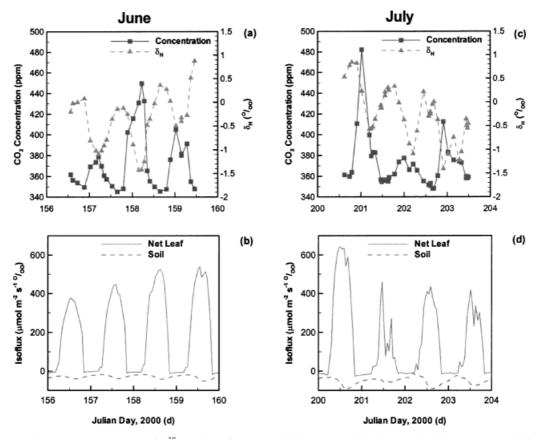


Fig. 11 Measured CO₂ concentration and δ^{18} O value of CO₂ sampled at 4.5 m (δ_H) during the (a) June 4–7 and (c) July 18–21 measurement campaigns. Simulated net leaf and soil ¹⁸O-isofluxes (µmol m⁻² s⁻¹) are also shown for these two periods (b, d). δ^{18} O values are relative to the V-PDB-CO₂ standard.

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et al., 2002). To test the impact of this variation, we imposed changes of these magnitudes while maintaining the average isotopic composition over the day unchanged. The ¹⁸O composition of atmospheric H₂O was forced to increase linearly with time between 8:00 and 18:00 hours, with a daily average 7‰ lighter than stem water at 8:00 hours. For the atmospheric isotopic composition of CO₂, we imposed a time-dependent sinusoidal profile with a maximum at 14:00 hours and a daily average of 0‰.

Imposing diurnal variation in the δ^{18} O value of atmospheric H₂O resulted in effectively no change in the soil water, stem water, and leaf water isotopic compositions, and therefore no change in the δ^{18} O value of the ecosystem CO₂ flux. Note that changes in the mean δ^{18} O value of atmospheric H₂O significantly impact the isotopic composition of ecosystem water pools. In contrast, imposing a varying δ^{18} O value of atmospheric CO2 resulted in increases of about 2‰ in the δ^{18} O value of the mid-day ecosystem CO₂ flux, consistent with the increase in the atmosphere-to-leaf $C^{18}OO$ fluxes. Note that an increase in the $\delta^{18}O$ value of the mid-day CO₂ flux will result in a lighter atmosphere. Night-time δ^{18} O values of the ecosystem CO₂ flux were relatively unaffected by the imposed variability in the δ^{18} O value of atmospheric CO₂.

Conclusions

In this paper, we present measurements and model predictions of CO₂ fluxes, latent and sensible heat fluxes, leaf and stem H₂¹⁸O composition, vertically resolved soil water H2¹⁸O composition, and H2¹⁸O and C¹⁸OO ecosystem fluxes in a tallgrass prairie in Oklahoma, USA. Generally, ISOLSM accurately predicted the dynamics and magnitude of these quantities over the 3-month simulation period. The paucity of measurements spanning the diurnal cycle makes it difficult to evaluate the short-term predictions of the model. However, the large predicted diurnal variations in the δ^{18} O values of ecosystem water pools and resultant C18OO fluxes argue that care must be taken when applying discrete measurements of these quantities to partition net ecosystem fluxes into component gross fluxes.

Continuous δ^{18} O values of atmospheric CO₂ and H₂O are required inputs to the model, and this information was unavailable for this site. Previous measurements in this system have shown that these quantities can vary substantially over the course of the day. Our simple sensitivity analysis indicated that diurnal variability in the δ^{18} O value of atmospheric H₂O had a small impact on the isotopic composition of ecosystem water pools, although sustained changes can have significant im-

pacts. Typical variations in the δ^{18} O value of atmospheric CO₂ increased the mid-day δ^{18} O value of the ecosystem CO₂ flux by up to 2‰; night-time values were relatively unaffected by the change.

Our simulation results indicate that incomplete equilibration between CO_2 and leaf water is common in this system. Further work is required to quantify accurately the extent of disequilibrium and its impact on net ecosystem $C^{18}OO$ fluxes. The ISOLSM simulations demonstrated that the ecosystem isoflux was dominated by leaf fluxes during the simulation period.

The development of ecosystem isotope models would benefit from further research on the biophysical and physiological controls on plant retro-diffused and soil-respired fluxes, as these fluxes account for the largest uncertainties in the atmospheric C¹⁸OO budget. Although leaf retro-diffused fluxes are theoretically well understood for C₃ plants (Farquhar & Lloyd, 1993; Farquhar et al., 1993; Ciais et al., 1997b), recent experimental evidence suggests a more complicated picture for C₄ plants (Gillon & Yakir, 2000; Helliker & Ehleringer, 2000; Gillon & Yakir, 2001). In particular, the impact of partial equilibration of CO₂ with leaf water and variations in the δ^{18} O value of water along the leaf need to be addressed. Recent experimental (Miller et al., 1999) and theoretical (Tans, 1998; Stern et al., 1999; Stern et al., 2001; Riley et al., 2002) work has advanced our understanding of soil-respired C¹⁸OO fluxes; further work is necessary to predict these fluxes across the range of soil types and environmental conditions found in real systems. These types of investigations will enhance our ability to use the δ^{18} O value of atmospheric CO₂ as a tracer of gross carbon exchanges and a constraint on land-surface biophysical processes.

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