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Whole genome sequence analysis of apparent treatment resistant hypertension status in participants from the Trans-Omics for Precision Medicine program

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Introduction: Apparent treatment-resistant hypertension (aTRH) is characterized by the use of four or more antihypertensive (AHT) classes to achieve blood pressure (BP) control. In the current study, we conducted single-variant and gene-based analyses of aTRH among individuals from 12 Trans-Omics for Precision Medicine cohorts with whole-genome sequencing data.

Methods: Cases were defined as individuals treated for hypertension (HTN) taking three different AHT classes, with average systolic BP \geq 140 or diastolic BP \geq 90 mmHg, or four or more medications regardless of BP (n = 1,705). A normotensive control group was defined as individuals with BP < 140/90 mmHg (n = 22,079), not on AHT medication. A second control group comprised individuals who were treatment responsive on one AHT medication with BP < 140/ 90 mmHg (n = 5,424). Logistic regression with kinship adjustment using the Scalable and Accurate Implementation of Generalized mixed models (SAIGE) was performed, adjusting for age, sex, and genetic ancestry. We assessed variants using SKAT-O in rare-variant analyses. Single-variant and gene-based tests were conducted in a pooled multi-ethnicity stratum, as well as self-reported ethnic/racial strata (European and African American).

Results: One variant in the known HTN locus, *KCNK3*, was a top finding in the multiethnic analysis (p = 8.23E-07) for the normotensive control group [rs12476527, odds ratio (95% confidence interval) = 0.80 (0.74–0.88)]. This variant was replicated in the Vanderbilt University Medical Center's DNA repository data. Aggregate genebased signals included the genes *AGTPBP*, *MYL4*, *PDCD4*, *BBS9*, *ERG*, and *IER3*.

Discussion: Additional work validating these loci in larger, more diverse populations, is warranted to determine whether these regions influence the pathobiology of aTRH.

KEYWORDS

blood pressure, antihypertensive response, whole genome sequencing, TOPMed, treatment resistant hypertension

1 Introduction

Hypertension (HTN) is a leading risk factor for cardiovascular and renal disease (Global Burden of Metabolic Risk Factors for Chronic Diseases Collaboration, 2014). While awareness and treatment of HTN have improved, many patients experience suboptimal blood pressure (BP) control despite treatment (NCD Risk Factor Collaboration, 2019; Wijkman et al., 2021). Resistant HTN is defined as BP above target while using \geq 3 antihypertensive (AHT) classes or the need to use \geq 4 drug classes to achieve the BP goal (Carey et al., 2018; Akinyelure et al., 2020). Apparent treatment-resistant hypertension (aTRH) is used commonly in observational studies, where pseudo-resistant HTN cannot be excluded (e.g., caused by sub-optimal medication adherence, improper BP measurement, or white coat effect) (Judd and Calhoun, 2014; Akinyelure et al., 2020).

While the exact prevalence of aTRH is unknown, the estimated prevalence in population-based studies is between 12% and 15% among adults with HTN (Carey et al., 2018; Irvin et al., 2019) and even higher in clinical studies, which suggests aTRH affects about 15%–30% of patients with HTN (Hyman and Pavlik, 2001; Carey et al., 2018; Takahashi et al., 2021). Individuals with aTRH are at a

higher risk of cardiovascular morbidity and mortality compared to individuals with controlled hypertension. Globally, an estimated 7.1 million deaths per year, as well as 62% of cerebrovascular disease and 49% of ischemic heart disease, are attributable to suboptimal BP control (systolic BP (SBP) > 115 mmHg) (Sarafidis and Bakris, 2008). The increased cardiovascular risk among patients with aTRH is likely due to the increased prevalence of concomitant co-morbidities such as diabetes, renal diseases, and obesity among persons with aTRH (Sarafidis and Bakris, 2008; Gupta et al., 2011) and is not exclusively due to high BP.

The etiology of aTRH is unknown, but is likely multifactorial (Vongpatanasin, 2014), with contributions of both behavioral/ lifestyle and genetic factors suspected. While several large studies have identified genetic variants associated with HTN (Levy et al., 2009; Newton-Cheh et al., 2009; Franceschini et al., 2011; Evangelou et al., 2018; Giri et al., 2019), there are far fewer genetic studies of aTRH with limited findings (Lynch et al., 2013; Fontana et al., 2014; Dumitrescu et al., 2017; El Rouby et al., 2019; Irvin et al., 2019; Takahashi et al., 2021). We hypothesize that identifying the genetic contributors to aTRH may provide further insight into the etiology of the disease. The present study comprises 12 studies from the NHLBI Trans-Omics for Precision Medicine (TOPMed) program for a case-control single-variant and rare-variant, gene-based analyses of aTRH that utilizes TOPMed's whole genome sequencing (WGS) data on over 29,000 participants (1,705 aTRH cases, 22,079 normotensive controls, and 5,424 controls who were treatment responsive). We attempted to validate our top variants in >10,000 participants from the International Consortium for Antihypertensive Pharmacogenomics Studies (ICAPS) and in >35,000 participants from Vanderbilt University Medical Center's DNA repository, BioVU.

2 Methods

2.1 Discovery study population

For the current study, we conducted genomic analyses of aTRH among 29,208 participants from 12 TOPMed WGS studies (Supplementary Tables S1–S3). WGS and phenotype data were pooled across the studies for analysis. Additional information about the design for each study and the sampling of participants for WGS is available in the Supplementary Material. Informed consent was obtained from all participants, and the protocols for each study were approved by the institutional review board of the participating institutions.

2.2 Phenotype definition

AHT treatment data for each study was extracted by medication inventory or self-report, and medication classes that counted toward the total count in the present study are listed in Supplementary Table S4. Participants with conditions that may lead to secondary forms of HTN were excluded from analysis, including those with a body mass index (BMI) > 40 kg/m² and/or an estimated glomerular filtration rate (eGFR) < 30 mL/min/1.73 m², which defines advanced chronic kidney disease (Stages G4 and G5). Cases were defined as individuals taking three different AHT classes for HTN treatment who had an average systolic BP \geq 140 mmHg or diastolic BP \geq 90 mmHg or taking four or more medication classes regardless of BP level (total n = 1,705). A normotensive control group (n = 22,079) included individuals with systolic BP < 140 mmHg and diastolic BP < 90 mmHg and not reporting use of AHT medication. A second control group comprised of individuals who had BP controlled (BP < 140 mmHg systolic and <90 mmHg diastolic) on one AHT medication class (n = 5,424). The Seventh Report of the Joint National Committee on Prevention, Detection, Evaluation, and Treatment of High Blood Pressure (JNC 7) blood pressure guideline definitions were used in the current study given these guidelines were used for treatment thresholds during the time the data was collected for the majority of studies included in the analysis (Chobanian et al., 2003). Phenotypes were harmonized across studies using a protocol developed by the TOPMed Blood Pressure Working Group and have been previously described (Kelly et al., 2022).

2.3 Whole genome sequencing (WGS) and quality control (QC)

A detailed description of the TOPMed WGS methods has been reported previously (Taliun et al., 2021). Briefly, WGS was performed at an average depth of >30x by six sequencing centers (Broad Genomics, Northwest Genome Center, Illumina, New York Genome Center, Baylor, and McDonnell Genome Institute) with Illumina HiSeq X10 technology (Mikhaylova et al., 2021). Genotype calling was conducted jointly across the TOPMed studies using the GotCloud pipeline (Jun et al., 2015). Variants were removed from analysis with excess heterozygosity or Mendelian inconsistencies, with overall missingness greater than 5%, or with overlapping centromeric regions. Genotypes with average read depth less than 10x were set to missing. Samples that were duplicated, had sex discrepancies, were misidentified or had consent issues, or had poor concordance across WGS and array data were excluded, as previously described (Kelly et al., 2022).

2.4 Statistical analysis

Multi-ethnic and race/ethnicity-specific (self-reported African American [AA] and self-reported European American [EA]) analyses were conducted and baseline demographic information is presented in Table 1. In single variant analyses, single-nucleotide variants (SNVs) and insertion-deletion (indel) variants with a minor allele frequency (MAF) ≥0.5% were retained. This threshold was chosen to leverage rare variants observed in WGS (a lower filter than generally considered by GWAS) but we did not include rarer variants due to limited case counts, especially in the self-reported race strata. For the SNV association analysis, we used a logistic mixed model in Scalable and Accurate Implementation of Generalized mixed models (SAIGE) software (Zhou et al., 2018). We adjusted for familial relationships by constructing a genetic relationship matrix as well as age, sex, and ancestry principal components. Ancestry principal components were generated by the TOPMed Data Coordinating Center in PC-AiR (Conomos et al., 2015), which is available in the GENESIS R package A saddle-point approximation was conducted to account for case-control imbalance in the analyses. A Bonferroni correction

TABLE 1 Baseline characteristics of study participants.

	aTRH cases	Treatment-responsive controls	Normotensive controls
N	1,705	5,424	22,079
Age, years	64.67 ± 10.44	60.80 ± 10.75	54.79 ± 13.33
Self-reported race/ethnicity, n (%)			
African	768 (45.04%)	1,924 (35.47%)	5,032 (22.79%)
Asian	51 (2.99%)	236 (4.35%)	2,478 (11.22%)
European	395 (23.17%)	2,730 (50.33%)	12,483 (56.54%)
Hispanic	448 (26.28%)	477 (8.79%)	1,844 (8.35%)
Other/Unknown	43 (2.52%)	57 (1.05%)	242 (1.10%)
Female, n (%)	1,039 (60.94%)	3,855 (71.07%)	14,141 (64.05%)
BMI, kg/m ²	29.99 ± 4.75	28.48 ± 4.77	26.55 ± 4.59
eGFR, mL/min/1.73m ²	71.21 ± 21.74	86.81 ± 21.24	92.00 ± 20.21
SBP, mmHg	164.27 ± 23.29	135.80 ± 12.31	116.33 ± 12.50
DBP, mmHg	88.98 ± 13.01	81.32 ± 9.27	70.39 ± 8.88
AHT Medication Use, n (%)			
Angiotensin-converting enzyme inhibitor (ACE-I)	959 (56.25%)	877 (16.17%)	0 (0%)
Aldosterone antagonist (MRA)	255 (14.96%)	46 (0.85%)	0 (0%)
Alpha-blocker (AB)	487 (28.56%)	75 (1.38%)	0 (0%)
Angiotensin receptor blocker (ARB)	385 (22.58%)	202 (3.72%)	0 (0%)
Beta-blocker (BB)	1,153 (67.62%)	1,097 (20.22%)	0 (0%)
Calcium channel blocker (CCB)	1,024 (60.06%)	1,052 (19.40%)	0 (0%)
Central acting agent (CAA)	225 (13.20%)	92 (1.70%)	0 (0%)
Diuretic	1,389 (81.47%)	1,801 (33.20%)	0 (0%)
Direct vasodilator (DV)	205 (12.02%)	319 (5.88%)	0 (0%)

Abbreviations: aTRH- apparent treatment-resistant hypertension; BMI- body mass index; eGFR-estimated glomerular filtration rate; SBP- systolic blood pressure; DBP- diastolic blood pressure; AHT-antihypertensive.

was applied to correct for multiple testing. The statistical significance threshold was set at p < 5.00E-08. As a secondary analysis, we implemented inverse variance-weighted, fixed effects meta-analysis on the summary statistics from the self-reported AA and EA strata for (Global Burden of Metabolic Risk Factors for Chronic Diseases Collaboration, 2014) aTRH versus normotensive controls and (NCD Risk Factor Collaboration, 2019) aTRH versus treatment-responsive controls, using METAL software (Willer et al., 2010). Statistical heterogeneity was evaluated using Cochran's chi-square test in METAL.

Combined burden and variance-component tests (i.e., omnibus sequence kernel association tests, SKAT-O) were applied to examine the association between aTRH status and aggregate rare variants in Efficient and Parallelizable Association Container Toolbox (EPACTS) software (EPACTS, 2019). Variants were functionally annotated using the Whole Genome Sequence Annotator (WGSA) (Liu et al., 2016) and classified as high or moderate impact (Supplementary Table S5). Generalized linear mixed models parallel to the single variant association tests were conducted, adjusting for age, sex, and ancestry principal components. Aggregate results were retained if there were i) 2-4 SNVs, none of which were singletons, or ii) \geq 5 SNVs comprising the gene region. The gene-

based signals with p < 2.50E-06 (=0.05/20,000 gene regions) and p < 1.00E-04 achieved statistical or suggestive significance, respectively.

2.5 Look-up validation in BioVU and ICAPS

We sought replication using existing data in 5,729 AA and 31,805 EA participants from Vanderbilt University Medical Center's DNA Repository (BioVU), as well as 10,801 participants comprising the International Consortium of Antihypertensive Pharmacogenomic Studies (ICAPS) aTRH-specific study (2,190 AA, 8,074 EA, and 537 Hispanic). Briefly in BioVU, aTRH status was defined through the presence of an HTN ICD-9 or ICD-10 code, treatment with an AHT, or having two outpatient, non-emergency department SBP >140 mmHg and/or two diastolic BP (DBP) measures >90 mmHg. Patients with aTRH were identified based on failure to achieve controlled BP on three AHTs, including a thiazide diuretic, or prescribed four or more medications regardless of achieving control. The comparison group of hypertensive patients who achieved BP control on one or two medications, excluding participants with chronic kidney disease (stage

							1	
rsID	CHR:BP	REF/EA	EAF	Or (95% Cl)	<i>p</i> -value ^a	Consequence	Gene(s)	
Multi-ethnicity: aTRH (n = 1,705) versus treated controls (n = $5,424$)								
rs1175395756	chr6:124515703	TTCGCTTTCTCTCCGTC/T	0.2586	0.35 (0.25, 0.51)	1.38E-08	intronic	NKAIN2	
rs6086545	chr20:935220	C/A	0.1712	1.35 (1.20, 1.52)	4.86E-07	intergenic	ANGPT4;RSPO4	
rs189036218	chr6:8077088	C/T	0.0054	3.73 (2.23, 6.25)	5.33E-07	ncRNA_intronic	EEF1E1-BLOC1S5	
rs538121738	chr3:84401497	G/A	0.0104	2.51 (1.73, 3.65)	1.25E-06	intergenic	LINC02008;LINC00971	
rs116250062	chr5:108665213	T/A	0.0065	4.14 (2.33, 7.35)	1.29E-06	intergenic	FBXL17;LINC01023	
rs138450763	chr6:8129403	G/A	0.0052	3.64 (2.16, 6.14)	1.34E-06	intergenic	EEF1E1-BLOC1S5;SLC35B3	
rs12335619	chr9:3669438	G/C	0.0497	0.64 (0.54, 0.77)	1.48E-06	ncRNA_intronic	RFX3-AS1	
rs140823205	chr12:103789678	T/TA	0.0270	0.57 (0.45, 0.72)	1.59E-06	intronic	NT5DC3	
rs1546149	chr8:15475599	G/A	0.2864	1.26 (1.14, 1.38)	1.64E-06	intergenic	SGCZ;TUSC3	
rs761571	chr20:932022	T/C	0.1880	1.31 (1.17, 1.47)	1.89E-06	intergenic	ANGPT4;RSPO4	
Multi-ethnicity: aTRH (n = 1,705) versus normotensive controls (n = 22,079)								
rs1175395756	chr6:124515703	TTCGCTTTCTCTCCGTC/T	0.3455	0.32 (0.23, 0.43)	3.83E-13	intronic	NKAIN2	
rs142718434	chr18:69043215	C/T	0.0065	2.56 (1.78, 3.68)	3.98E-07	intronic	CCDC102B	
rs74574593	chr21:36426260	G/A	0.0183	1.74 (1.40, 2.17)	6.77E-07	intergenic	CHAF1B;CLDN14	
rs61312821	chr5:160927044	G/A	0.1129	1.42 (1.23, 1.63)	7.51E-07	intergenic	ATP10B;LINC02159	
rs12476527	chr2:26692756	G/T	0.4896	0.80 (0.74, 0.88)	8.23E-07	UTR5	KCNK3	
rs79890489	chr3:27831750	C/A	0.0059	2.47 (1.72, 3.55)	1.05E-06	ncRNA_exonic	LINC01981	
rs115299786	chr3:27830202	T/G	0.0062	2.43 (1.70, 3.48)	1.18E-06	ncRNA_intronic	LINC01980	
rs191754301	chr5:26402669	G/A	0.0065	2.40 (1.68, 3.41)	1.20E-06	intergenic	LINC02211;CDH9	
rs899934113	chr3:27845147	TA/T	0.0062	2.42 (1.69, 3.47)	1.26E-06	intronic	LINC01980	
rs76818748	chr15:54329458	T/C	0.0586	0.63 (0.52, 0.76)	1.46E-06	intronic	UNC13C	
AFR: aTRH (n	= 768) versus tre	ated controls (n = $1,924$)						
rs1546149	chr8:15475599	G/A	0.3398	1.45 (1.27, 1.65)	2.73E-08	intergenic	SGCZ;TUSC3	
rs183165107	chr5:38365438	G/A	0.0100	5.48 (2.90, 10.36)	1.59E-07	intronic	EGFLAM	
rs80116392	chr11:133314740	G/C	0.0721	1.87 (1.48, 2.37)	2.06E-07	intronic	OPCML	
rs566745771	chr12:132295343	C/T	0.0050	10.27 (4.20, 25.12)	3.28E-07	intronic	GALNT9	
rs74179759	chr20:15137285	G/A	0.0063	7.83 (3.51, 17.45)	4.78E-07	intronic	MACROD2	
rs6595479	chr5:123821788	G/A	0.0520	2.02 (1.53, 2.66)	7.60E-07	intergenic	CSNK1G3;LINC01170	
rs76368370	chr18:71114951	C/G	0.0396	2.21 (1.62, 3.03)	7.68E-07	intergenic	GTSCR1;LINC01541	
rs116645995	chr1:239428421	C/T	0.0137	3.78 (2.23, 6.40)	7.87E-07	intronic	CHRM3	
rs17126926	chr10:109992693	T/C	0.0555	1.98 (1.51, 2.60)	9.83E-07	ncRNA_intronic	ADD3-AS1	
rs115632010	chr4:93349186	A/T	0.0165	3.48 (2.11, 5.74)	9.99E-07	intronic	GRID2	
AFR: aTRH (n = 768) versus normotensive controls (n = $5,032$)								
rs62434735	chr6:147600061	G/A	0.0569	2.06 (1.58, 2.68)	7.45E-08	intergenic	SAMD5;SASH1	
rs183634807	chr2:32050050	G/A	0.0138	3.88 (2.33, 6.47)	1.92E-07	intergenic	DPY30;SPAST	
rs11963682	chr6:147594975	C/T	0.0555	2.00 (1.54, 2.61)	2.85E-07	intergenic	SAMD5;SASH1	
rs117043225	chr8:56731207	C/T	0.0054	8.78 (3.75, 20.57)	5.71E-07	intergenic	LINC00968;IMPAD1	

TABLE 2 Top ten variants observed for association analysis of aTRH in each self-reported race/ethnicity-control strata.

(Continued on following page)

rsID	CHR:BP	REF/EA	EAF	Or (95% Cl)	<i>p</i> -value ^a	Consequence	Gene(s)	
rs73987603	chr2:212980098	G/A	0.1961	0.68 (0.59, 0.79)	6.68E-07	intergenic	MIR4776-2;IKZF2	
rs113287654	chr12:127301710	G/C	0.0761	1.75 (1.40, 2.20)	9.14E-07	intergenic	LINC02376;LINC02375	
rs11017247	chr10:130427166	A/T	0.3075	0.74 (0.65, 0.83)	1.09E-06	intergenic	GLRX3;MIR378C	
rs190813229	chr2:32006833	C/T	0.0115	3.97 (2.28, 6.91)	1.10E-06	intronic	MEMO1	
rs115965650	chr2:226205376	C/T	0.0070	7.53 (3.34, 16.97)	1.12E-06	intergenic	LOC646736;MIR5702	
rs74552450	chr2:226251404	T/A	0.0070	6.93 (3.17, 15.16)	1.23E-06	intergenic	LOC646736;MIR5702	
EUR: aTRH (n = 395) versus treated controls (n = 2,730)								
rs11711860	chr3:38776013	C/A	0.2566	1.64 (1.37, 1.97)	6.16E-08	intronic	SCN10A	
rs73064548	chr3:38782346	C/G	0.2567	1.64 (1.37, 1.97)	6.18E-08	intronic	SCN10A	
rs62244107	chr3:38778263	C/T	0.2566	1.64 (1.37, 1.96)	6.30E-08	intronic	SCN10A	
rs62244108	chr3:38781988	G/A	0.2567	1.64 (1.37, 1.96)	6.34E-08	intronic	SCN10A	
rs62244106	chr3:38777782	T/C	0.2570	1.64 (1.37, 1.96)	6.54E-08	intronic	SCN10A	
rs7430726	chr3:38776830	T/C	0.2570	1.64 (1.37, 1.96)	6.54E-08	intronic	SCN10A	
rs11129808	chr3:38785608	T/C	0.2571	1.64 (1.37, 1.96)	7.83E-08	intronic	SCN10A	
rs12172966	chr3:38785717	T/G	0.2570	1.63 (1.36, 1.95)	1.08E-07	intronic	SCN10A	
rs11927856	chr3:38779283	T/C	0.2586	1.62 (1.36, 1.94)	1.11E-07	intronic	SCN10A	
rs4072090	chr3:38773939	G/T	0.2574	1.62 (1.35, 1.94)	1.35E-07	intronic	SCN10A	
EUR: aTRH (n	= 395) versus no	rmotensive controls (n = 12	,483)					
rs62244107	chr3:38778263	C/T	0.2532	1.54 (1.30, 1.82)	5.64E-07	intronic	SCN10A	
rs11711860	chr3:38776013	C/A	0.2532	1.54 (1.30, 1.82)	5.73E-07	intronic	SCN10A	
rs62244108	chr3:38781988	G/A	0.2533	1.54 (1.30, 1.82)	6.02E-07	intronic	SCN10A	
rs7430726	chr3:38776830	T/C	0.2534	1.54 (1.30, 1.82)	6.24E-07	intronic	SCN10A	
rs73064548	chr3:38782346	C/G	0.2534	1.54 (1.30, 1.82)	6.31E-07	intronic	SCN10A	
rs62244106	chr3:38777782	T/C	0.2534	1.54 (1.30, 1.82)	6.35E-07	intronic	SCN10A	
rs11129808	chr3:38785608	T/C	0.2536	1.53 (1.30, 1.81)	6.76E-07	intronic	SCN10A	
rs34623024	chr10:9717155	T/G	0.2002	1.58 (1.32, 1.89)	8.42E-07	intergenic	LOC101928272;LOC1019282	
rs12779933	chr10:9715550	C/T	0.1994	1.58 (1.32, 1.89)	8.60E-07	intergenic	LOC101928272;LOC1019282	
rs12172966	chr3:38785717	T/G	0.2534	1.53 (1.29, 1.81)	8.67E-07	intronic	SCN10A	

TABLE 2 (Continued) Top ten variants observed for association analysis of aTRH in each self-reported race/ethnicity-control strata.

^ap-value using saddle point approximation. Statistical significance p < 5.00E-08 (p-value bolded).

Abbreviations: rsID-reference SNP, cluster ID; CHR-chromosome; BP- base position hg38 build; REF- reference allele; EA-effect allele; EAF- effect allele frequency; OR-odds, ratio; CIconfidence interval; aTRH- apparent treated-resistant hypertension; UTR5- 5' untranslated region; ncRNA-non-coding RNA; AFR-self-reported African American race/ethnicity; EUR-selfreported European American race/ethnicity.

4 and 5), or those with secondary causes of HTN. A second control group consisted of individuals without HTN as defined above (Shuey et al., 2018). In the ICAPS aTRH analysis, aTRH was defined as uncontrolled HTN using three or more AHTs from different classes (BP \geq 140/90 mmHg) or controlled HTN on four or more AHT medications. Treatment-responsive controls were individuals with controlled BP (<140/90) with the use of three or fewer AHTs. Quality control and analyses were performed at the cohort level for the five randomized controlled clinical trials, and then meta-analyzed (combined and race-stratified). Variant lookups were performed for both replication cohorts based on the top findings presented in Table 2.

Replication significance was determined as p < 1.39E-03 (=0.05/36 unique genetic loci).

3 Results

3.1 Study characteristics

The baseline characteristics of the study participants are shown in Table 1. Across the 12 TOPMed cohorts, 1,705 aTRH cases were compared to i) 5,424 treatment-responsive controls and ii) 22,079 normotensive controls. On average, aTRH cases (64.67 years) were older than treatment-responsive (60.80 years) and normotensive (54.79 years) controls and were more likely to be AA (45.04% cases). Cases also presented, on average, with a higher BMI (29.99 kg/m²) compared with treatment-responsive (28.48 kg/m²) and normotensive (26.55 kg/m²) controls, as well as lower glomerular filtration rate compared to treatment-responsive and normotensive controls (71.21 mL/min/1.73 m² versus 86.81 mL/min/1.73 m² and 92.00 mL/min/1.73 m², respectively), though on average still in the normal range (Table 1).

3.2 Single-variant analysis

The top 10 results for each case-control model are presented in Table 2. When comparing aTRH cases to normotensive controls, the top finding for the pooled, multi-race/ethnicity analysis was rs1175395756, an indel variant located in the sodium/potassium transporting ATPase interacting 2 (NKAIN2) gene. For rs1175395756, the odds of having aTRH were lower among those with the deletion (odds ratio [OR] = 0.32, 95% confidence interval [95% CI] = 0.23–0.43, *p* = 3.83E-13, effect allele frequency [EAF] = 0.27). Further, rs12476527, located in the 5' untranslated region of the potassium two pore domain channel subfamily K member 3 gene (KCNK3), resulted in lower odds of aTRH (OR [95% CI] = 0.80 [0.74–0.88], p = 8.23E-07) for those with an effect allele T. When comparing the aTRH cases to 5,424 treatment-responsive controls, the effect of rs1175395756 was consistent (OR [95% CI] = 0.35 [0.25-0.51], p = 1.38E-08, EAF = 0.35) with the analysis of normotensive controls. Further, the effect alleles of 11 intergenic variants located on chromosome 20 between the angiopoietin-4 (ANGPT4) and R-spondin 4 (RSPO4) genes (EAF ranging from 0.15-0.18) had increased odds of aTRH (OR [95% CI] = 1.35 [1.20–1.52], p = 4.86E-07 for the lead SNV rs6086545).

Among the EAs, the top findings for 395 aTRH cases compared to both control groups were intronic to the sodium voltage-gated channel alpha subunit 10 (SCN10A) gene. All 13 of the suggestive variants observed for the normotensive control group comparison were also associated with aTRH for the treatment-responsive control group comparison (Table 2 and Supplementary Tables S6, S7) with $p \le 2.36$ E-06. The majority of the suggestive *SCN10A* variants were in strong LD ($r^2 > 0.8$; Supplementary Table S8). The only statistically significant finding for the single variant analysis in the African-ethnicity stratum was an intergenic variant (rs1546149) between the zeta-sarcoglycan gene (SGCZ) and the tumor suppressor candidate 3 gene (TUSC3) on chromosome 8 (Table 2). All SNV results that met the suggestive statistical significance threshold p < 1.00E-05 in each ethnic strata and control group comparison can be found in Supplementary Tables S6, S7, S9-S12. While we observed some overlap by control group analysis in ethnic strata, there was limited overlap across race/ ethnicity-specific analyses. Manhattan and QQ plots for each analysis are presented in Supplementary Figures S1-S6, and there was no evidence for deviation of *p*-values from their expected values (Supplementary Table S13). In a secondary meta-analysis of the AA and EA strata results, we did not observe any statistically significant findings. rs1546149 had nominal significance for the treatmentresponsive control group (*p* = 7.31E-08) (Supplementary Tables S14, S15; Supplementary Figures S7, S8).

Upon variant look-up in BioVU and ICAPS, rs12476527 located in KCNK3 was replicated in the BioVU EA aTRH versus normotensive control analysis with the same direction of effect (OR [95% CI] = 0.88 [0.82-0.95], p = 6.10E-04) (Supplementary Table S16). While not statistically significantly associated with aTRH in our other analyses, the variant had a consistent direction of effect in all our analyses, with marginally significant p-values for the multi-ethnic aTRH versus treated controls (OR [95% CI] = 0.88 [0.80-0.96], p = 5.15E-03), AA aTRH versusnormotensive controls (OR [95% CI] = 0.83 [0.72-0.95], p = 8.47E-03), and EA aTRH versus normotensive controls (OR [95% CI] = 0.81 [0.69-0.94], p = 5.81E-03). Likewise, while this variant was not significant in the BioVU AA lookup, a consistent direction of effect (OR [95% CI] = 0.82 [0.68–0.98], *p* = 3.18E-02) was observed. Since rs12476527 was originally discovered in our analysis of aTRH cases versus normotensive controls, we did not look this variant up in the ICAPS replication cohort, which included treatment-responsive controls only.

3.3 Gene-based, rare-variant analysis

Gene-based results are presented in Table 3 (p < 2.5E-06). Expanded results for genes with suggestive significance are presented in Supplementary Tables S17-S19. Biologically plausible signals included the genes ATP/GTP binding carboxypeptidase 1 (AGTPBP1), myosin light chain 4 (MYL4), programmed cell death 4 (PDCD4), Bardet-Biedel Syndrome 9 (BBS9), erythroblast transformation-specific transcription factor ERG (ERG), and immediate early response 3 (IER3). In the multi-ethnic aggregate analysis of high-impact variants, AGTPBP1 was associated with aTRH in comparison to normotensive controls (5 variants, 4 singletons p = 3.04E-07). Among AA participants, the aggregate of 20 total variants (including 10 singletons) of high-moderate impact in the MYL4 gene region, was associated with aTRH in comparison to the normotensive control group (p = 8.83E-07). Also, in this stratum, the aggregate of five total high-impact variants (three singletons) in the PDCD4 gene region was associated (p = 1.59E-06) with case status when compared to the normotensive control group. Three gene regions of interest were observed in the EA analysis including BBS9 (high impact; normotensive controls; p = 1.29E-14), ERG (high-moderate impact; normotensive controls; p = 1.03E-10), and *IER3* (high-moderate impact; p = 2.48E-07 and p = 2.84E-07 for normotensive and treatment responsive controls, respectively).

4 Discussion

In the current study utilizing sequencing data from over 29,000 participants from TOPMed (26.44% AAs, 9.47% Asian, 53.44% EA, 9.48% Hispanic ethnicity, 1.17% other or unknown race/ethnicity), we investigated the genetic contributors to aTRH. Analyses were performed in a pooled, multi-ethnic analysis, as well as in self-reported AA and EA racial/ethnic strata. We identified

TABLE 3 Statistically significant (p < 2.50E-06) gene-based results.

CHR:BP (hg38)	Gene	Total variants	Singletons	<i>p</i> -value	Control group	Functional impact
Multi-ethnicity						
chr7:102573812-102616680	RASA4	5	4	2.44E-11	Normotensive	High-Moderate
chr11:617593-624900	CDHR5	6	4	2.93E-07	Normotensive	High
chr9:85547252-85741961	AGTPBP1	5	4	3.04E-07	Normotensive	High
chr20:44910144-44958409	PABPC1L	5	2	5.66E-07	Normotensive	High
chr3:157146572-157160085	CCNL1	51	34	7.75E-07	Normotensive	High-Moderate
chr20:35616035-35620978	SPAG4	6	2	1.23E-06	Normotensive	High
chr5:151016922-151017411	AC008641.1	2	0	2.16E-06	Normotensive	High-Moderate
AFR		I			1	1
chr17:47209444-47223029	MYL4	20	10	8.83E-07	Normotensive	High-Moderate
chr10:110876746-110896067	PDCD4	5	3	1.59E-06	Normotensive	High
EUR		·			I	l
chr7:33152724-33605225	BBS9	5	5	1.29E-14	Normotensive	High
chr19:2515043-2520681	GNG7	6	6	4.55E-13	Normotensive	High-Moderate
chr10:122454730-122456931	ARMS2	7	7	5.33E-12	Normotensive	High-Moderate
chr22:37971252-37983774	SOX10	11	9	3.60E-11	Treated	High-Moderate
chr21:38383429-38575700	ERG	28	23	1.03E-10	Normotensive	High-Moderate
chr6:31960118-31968877	SKIV2L	6	4	3.46E-10	Normotensive	High
chr3:150660005-150703898	ERICH6	5	5	5.27E-10	Normotensive	High
chr1:153070631-153070827	SPRR2B	5	2	1.32E-09	Normotensive	High-Moderate
chr16:87398822-87403097	MAP1LC3B	5	5	9.13E-09	Normotensive	High
chr22:37971252-37983774	SOX10	29	18	9.81E-09	Normotensive	High-Moderate
chr11:72216900-72221653	FOLR2	8	4	4.65E-08	Normotensive	High
chr19:42753372-42855618	PSG8	8	6	7.53E-08	Normotensive	High
chr12:48830188-48844031	DDX23	8	7	1.16E-07	Normotensive	High
chr9:128371381-128389729	URM1	23	18	1.28E-07	Normotensive	High-Moderate
chr3:50347615-50350643	NPRL2	27	17	2.09E-07	Normotensive	High-Moderate
chr6:159906956-159907931	MAS1	2	0	2.47E-07	Normotensive	High
chr6:30743938-30744514	IER3	14	10	2.48E-07	Normotensive	High-Moderate
chr6:30743938-30744514	IER3	7	5	2.84E-07	Treated	High-Moderate
chr16:56638740-56639943	MT1A	7	7	3.14E-07	Normotensive	High-Moderate
chr1:225887081-225888019	RP4-559A3.7	6	5	4.05E-07	Normotensive	High-Moderate
chr1:43307827-43321659	TIE1	5	4	4.74E-07	Normotensive	High
chr5:155013860-155017553	KIF4B	9	8	5.63E-07	Normotensive	High
chr3:10034704-10101203	FANCD2	5	3	7.28E-07	Normotensive	High
chr1:54609269-54619933	FAM151A	5	2	7.35E-07	Normotensive	High
chr19:2785665-2813226	THOP1	30	19	7.58E-07	Normotensive	High
chr5:151016922-151017411	AC008641.1	2	0	1.17E-06	Normotensive	High-Moderate

(Continued on following page)

CHR:BP (hg38)	Gene	Total variants	Singletons	<i>p</i> -value	Control group	Functional impact ^a
chr13:48342667-48476805	RB1	5	4	1.19E-06	Treated	High
chr22:21383992-21388771	RIMBP3B	5	5	1.35E-06	Normotensive	High-Moderate
chr11:94429946-94492670	MRE11A	27	20	1.40E-06	Normotensive	High
chr12:48342998-48350059	ZNF641	5	5	1.57E-06	Normotensive	High
chr20:5127096-5190229	CDS2	23	15	2.04E-06	Normotensive	High-Moderate
chr1:209617505-209650942	LAMB3	8	6	2.08E-06	Normotensive	High
chr2:74530009-74532941	HTRA2	6	4	2.33E-06	Normotensive	High
chr17:40074515-40093367	THRA	27	20	2.36E-06	Normotensive	High-Moderate
chr3:111999445-112013701	TAGLN3	17	12	2.37E-06	Normotensive	High-Moderate

TABLE 3 (Continued) Statistically significant (p < 2.50E-06) gene-based results.

^aDefinition of high versus high-moderate functional impact can be found in Supplementary Table S5.

Abbreviations: AFR-self-reported African American race/ethnicity; EUR-self-reported European American race/ethnicity.

three variants at the single-variant level and 44 gene regions that exceeded statistical significance for association with aTRH.

One of our top findings was located in the KCNK3 gene, which encodes a member of the superfamily of potassium channel proteins and was replicated in the BioVU data (Gene, 2004). KCNK3 has been previously linked to HTN in genetic studies. In a multi-racial/ethnic analysis of >7,800 individuals of European, African, Hispanic, and Chinese ethnicity within the Multi-Ethnic Study of Atherosclerosis (MESA) study, several KCNK3 variants were associated with measures of BP (including SBP, DBP, mean arterial pressure, and pulse pressure) across race/ethnic groups. Additionally, this study showed in a rodent model, that genetic disruption of KCNK3 produces hyperaldosteronism (Manichaikul et al., 2016). More recently, loss-of-function KCNK3 mutations have been identified as likely causative in heritable pulmonary arterial HTN (Ma et al., 2013; West et al., 2021). Specifically, rs12476527 has been associated with both pulse pressure (p = 2.55E-21) and SBP (p = 5.89E-50) across multi-ethnic meta-analyses (Giri et al., 2019).

A single-variant intronic to NKAIN2 was associated with aTRH in both control strata in the multi-ethnic analysis. NKAIN2 is a transmembrane protein that interacts with the beta subunit of a sodium/potassium transporting ATPase (Gene, 2004). Previous studies have shown that NKAIN2 may be related to neurological phenotypes, such as schizophrenia and bipolar disease (Blokland et al., 2022), as well as being a susceptibility locus for BMI (Yasukochi et al., 2018). This variant has not been directly associated with HTN to our knowledge, though a 2016 study reported a different SNV (not in LD with rs1175395756) in NKAIN2 (rs332607) was associated with pubertal SBP and adult SBP in the International Consortium of Blood Pressure (Parmar et al., 2016). Of note, the sodium/potassium transporting ATPase-s are ubiquitously expressed. In the heart they influence cell calcium and contractility by generating the Na + gradient driving Ca++ out of the cell via the Na+/Ca++ exchanger. They are also receptors for cardiac glycosides such as digitalis (a heart failure and antiarrhythmic drug) which increases cardiac contractility (McDonough et al., 2002). Since most AHTs act on the kidneys, heart and vasculature it is difficult to directly link NKAIN2 (primarily expressed in the brain) to AHT treatment response. However, some AHTs do influence the sympathetic nervous system (e.g., alpha blockers, centrally acting agents, and beta blockers) which could potentially link a mechanism to this gene, though that is beyond the scope of our study.

An additional intergenic region between ANGPT4 and RSPO4 was identified in our multi-ethnic analysis utilizing the treatmentresponsive controls. ANGPT4 encodes an angiopoietin shown to be involved in vascular development (Gene, 2004), angiogenesis, and vascular disease (Abu-Farha et al., 2018; Ali et al., 2022). RSPO4 is a member of the R-spondin family of proteins that have emerged as important regulators of WNT signaling (Jin and Yoon, 2012). There is evidence for Wnt's role in the regulation of BP and the pathogenesis of HTN (Abou Ziki and Mani, 2017). Lastly, and convincing in our European-specific analysis, 15 intronic variants of SCN10A met or exceeded our suggestive significance threshold (p <1.00E-05). SCN10A encodes the voltage-gated sodium channel, Nav1.8, and is flanked by SCN5A and SCN11A, both of which encode for voltage-gated sodium channels, Nav1.5 and Nav1.9, respectively (Macri et al., 2018). SCN10A is expressed in the myocardium and preferentially in the specialized Purkinje fibers of the cardiac conduction system (Sotoodehnia et al., 2010). Several genome-wide association studies have reported that SCN10A is involved in heart rate and dysrhythmia (Chambers et al., 2010; Holm et al., 2010; Pfeufer et al., 2010; Sotoodehnia et al., 2010; Liu et al., 2020). While the association between SCN10A and cardiac conduction is established making this gene especially interesting in our study of aTRH, SCN10A and other voltage gated sodium channels are not AHT drug targets. However, voltage gated sodium channels are important drug targets for other disorders (e.g., seizures, cardiac arrhythmias) (Wisedchaisri and Gamal El-Din, 2022) and further investigation of how different AHTs may indirectly affect their action is warranted (Liu et al., 2020). Most of our novel findings were derived from our aggregate, rare-variant (gene-based) analysis, which was to be expected due to the high coverage of rare genetic variants through TOPMed sequencing. In the multi-ethnic analysis, the gene region on chromosome 9 spanning the AGTPBP1 gene was of interest due to its previous association with cardiometabolic-related traits. AGTPBP1 is a metallocarboxypeptidase that mediates protein deglutamylation of tubulin and non-tubulin target proteins (Gene, 2004) and has been associated with BMI (Yasukochi et al., 2018), as well as increased blood pressure in spontaneously hypertensive rat lines (Bell et al., 2011).

An additional two gene regions were associated with aTRH in the AA stratum. MYL4, located on chromosome 17, and PDCD4, located on chromosome 10, both exceeded the significance threshold for the aTRH-normotensive case-control analysis. MYL4 encodes a myosin alkali light chain that is found in embryonic muscle and adult atria (Gene, 2004). Rs117154502, an intronic variant of MYL4, has been associated with both pulse pressure and SBP among Europeans (Plotnikov et al., 2022). PDCD4, the other gene region that was identified in the AA stratum, is a tumor suppressor gene that encodes a protein that binds to eukaryotic translation initiation factor 4A1 (Gene, 2004). PDCD4 is a direct target of miR-532-5p, a microRNA that has a potential function in cardiomyocyte response to hypoxia (Jin et al., 2018). BBS9, ERG, and IER3 were all gene regions implicated with aTRH in the European stratum. BBS9 is a gene associated with Bardet-Biedel Syndrome (BBS), an autosomal recessive condition with multiple clinical features including renal abnormalities, obesity, and HTN (Zhao and Rahmouni, 2022). ERG encodes a member of the erythroblast transformation-specific family of transcription factors (Gene, 2004). Recently, ERG has emerged as a major regulator of endothelial function (Shah et al., 2016; Kalna et al., 2019), and is important in the regulation of tissue-specific processes that include hematopoiesis, angiogenesis, and vascular inflammation (Seth and Watson, 2005; Shah et al., 2016). Of note, our variants comprising this gene region are not in LD with a previously reported ERG variant (rs117870289) associated with DBP (Giri et al., 2019). IER3 may play a role in the ERK signaling pathway by inhibiting the dephosphorylation of ERK (Gene, 2004). Animal studies have shown results that point to a cardiovascular role of IER related to blood pressure (De Keulenaer et al., 2002; Arlt and Schafer, 2011).

As one of the largest WGS analyses on aTRH, this study has several strengths. First, by leveraging the TOPMed consortium data, we were able to interrogate both common and rare variants in a racially/ethnically diverse sample. This was facilitated by the TOPMed Blood Pressure Working Group, which harmonized phenotypes and related variables in a pooled dataset across observational cohorts to pair with the pooled genomic data. Our study is not without limitations. Although it is one of the largest WGS interrogations of aTRH to date, the sample size, particularly in the AA and EA strata, was limited due to the extreme nature of this BP trait. Further, we had limited information on white coat HTN, adherence information, and medication doses, which could contribute to phenotype misclassification. While we did observe limited replication for rs12476527 in the BioVU EAs and AAs, we could not directly replicate the results from our multi-ethnic strata due to limitations of the summary statistics available (i.e., no racecombined analyses available in silico). Further, most of our novel findings were in the aggregate, gene-based analysis and we did not find another study population with appropriate phenotyping and sequencing data to facilitate replication. In general, studying rare variants linked to drug response introduces a series of challenges regarding the validation of findings both concerning the ascertainment of the replication sample and difficulties identifying the same variants comprising the gene region of interest (Hirschhorn and Altshuler, 2002). Difficulties identifying relevant variants may be further amplified by limited subsamples with specific drug exposures and/or strict inclusion criteria for clinical studies (Aslibekyan et al., 2013). Unfortunately, translation may be limited by the lack of validation in these types of studies. However, the increasing availability of large datasets from diverse biobanks linked to electronic medical records may alleviate current challenges. In conclusion, our main findings included variants in KCNK3, SCN10A (a known cardiac gene), NKAIN2, and the intergenic region between ANGPT4-RSPO4, and aggregate findings in the gene regions of AGTPBP1, MYL4, PDCD4, BBS9, ERG, and IER3. While many of these SNVs and gene regions have prior cardiovascular implications, most have not been associated with HTN or BP phenotypes (except for KCNK3) and remain worthy of further investigation of aTRH.

Data availability statement

Publicly available datasets were analyzed in this study. This data can be found here: The datasets analyzed for this study can be found in the National Center for Biotechnology Information database of Genotypes and Phenotypes (dbGaP) at the following accession numbers: Amish Complex Disease Research Program (AMISH) (phs000956); Atherosclerosis Risk in Communities (ARIC) study (phs001211); The BioMe Biobank at Mount Sinai (BioMe) (phs001644); Cleveland Family Study (CFS) (phs000954); Cardiovascular Health Study (CHS) (phs001368); Genetic Epidemiology Network of Arteriopathy (GENOA) (phs001345); Genetic Epidemiology Network of Saltandhyphen;Sensitivity (GenSALT) (phs001217); Hypertension Genetic Epidemiology Network (HyperGEN) (phs001293); Jackson Heart Study (JHS) (phs000964); Multi-Ethnic Study of Atherosclerosis (MESA) (phs001416. v1. p1); Taiwan Study of Hypertension using Rare Variants (THRV) (phs001387); and Women's Health Initiative (WHI) (phs001237).

Ethics statement

Informed consent was obtained from all participants, and the protocols for each study were approved by the institutional review board (IRB) of the participating institutions. For the Amish Study, all study protocols were approved by the IRB at the University of Maryland Baltimore. Informed consent was obtained from each study participant. The BioMe cohort was approved by the IRB at the Icahn School of Medicine at Mount Sinai. All BioMe participants provided written, informed consent for genomic data sharing. Cleveland Family Study was approved by the IRB of Case Western Reserve University and Mass General Brigham (formerly Partners Healthcare). Written informed consent was obtained from all participants. All CHS participants provided informed consent and the study was approved by the IRB (or ethics review committee) of University of Washington. GENOA-Written informed consent was obtained from all subjects and approval was granted by

participating IRBs (University of Michigan, University of Mississippi Medical Center, and Mayo Clinic). All GOLDN participants provided informed consent, and the study was approved by the IRB of the University of Kentucky. All HyperGEN participants provided informed consent, and the study was approved by the IRB of the University of Kentucky. The JHS study was approved by Jackson State University, Tougaloo College, and the University of Mississippi Medical Center IRBs, and all participants provided written informed consent. All MESA participants provided written informed consent, and the study was approved by the IRBs at the Lundquist Institute (formerly Los Angeles BioMedical Research Institute) at Harbor-UCLA Medical Center, University of Washington, Wake Forest School of Medicine, Northwestern University, University of Minnesota, Columbia University, and Johns Hopkins University. All THRV participants provided informed consent, and the study was approved by the IRB at The Lundquist Institute (formerly Los Angeles BioMedical Research Institute) at Harbor-UCLA Medical Center. All THRV participants provided informed consent, and the study was approved by the IRB at Washington University in St. Louis. All WHI participants provided informed consent and the study was approved by the IRB of the Fred Hutchinson Cancer Research Center. ARIC-All subjects provided informed consent and the study was approved by the Institutional Review Board (IRB) of the Johns Hopkins University School of Medicine. The studies were conducted in accordance with the local legislation and institutional requirements. Written informed consent for participation was not required from the participants or the participants' legal guardians/next of kin in accordance with the national legislation and institutional requirements.

Author contributions

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Conflict of interest

BP serves on the Steering Committee of the Yale Open Data Access Project funded by Johnson and Johnson.

References

Abou Ziki, M. D., and Mani, A. (2017). Wnt signaling, a novel pathway regulating blood pressure? State of the art review. *Atherosclerosis* 262, 171–178. doi:10.1016/j. atherosclerosis.2017.05.001

Abu-Farha, M., Cherian, P., Qaddoumi, M. G., AlKhairi, I., Sriraman, D., Alanbaei, M., et al. (2018). Increased plasma and adipose tissue levels of ANGPTL8/Betatrophin and ANGPTL4 in people with hypertension. *Lipids Health Dis.* 17 (1), 35. doi:10.1186/s12944-018-0681-0

Akinyelure, O. P., Sakhuja, S., Colvin, C. L., Clark, D., 3rd, Jaeger, B. C., Hardy, S. T., et al. (2020). Cardiovascular health and transition from controlled blood pressure to apparent treatment resistant hypertension: the Jackson heart study and the REGARDS study. *Hypertension* 76 (6), 1953–1961. doi:10.1161/HYPERTENSIONAHA.120.15890

Ali, F., Khan, A., Muhammad, S. A., and Hassan, S. S. U. (2022). Quantitative realtime analysis of differentially expressed genes in peripheral blood samples of hypertension patients. *Genes (Basel)*. 13 (2), 187. doi:10.3390/genes13020187

Arlt, A., and Schafer, H. (2011). Role of the immediate early response 3 (IER3) gene in cellular stress response, inflammation and tumorigenesis. *Eur. J. Cell Biol.* 90 (6-7), 545–552. doi:10.1016/j.ejcb.2010.10.002

Aslibekyan, S., Claas, S. A., and Arnett, D. K. (2013). To replicate or not to replicate: the case of pharmacogenetic studies: establishing validity of pharmacogenomic findings: from replication to triangulation. *Circ. Cardiovasc Genet.* 6 (4), 409–412. discussion 12. doi:10.1161/CIRCGENETICS.112.000010

Bell, R., Herring, S. M., Gokul, N., Monita, M., Grove, M. L., Boerwinkle, E., et al. (2011). High-resolution identity by descent mapping uncovers the genetic basis for blood pressure differences between spontaneously hypertensive rat lines. *Circ. Cardiovasc Genet.* 4 (3), 223–231. doi:10.1161/CIRCGENETICS.110.958934

Blokland, G. A. M., Grove, J., Chen, C. Y., Cotsapas, C., Tobet, S., Handa, R., et al. (2022). Sex-dependent shared and nonshared genetic architecture across mood and psychotic disorders. *Biol. Psychiatry* 91 (1), 102–117. doi:10.1016/j.biopsych.2021. 02.972

Carey, R. M., Calhoun, D. A., Bakris, G. L., Brook, R. D., Daugherty, S. L., Dennison-Himmelfarb, C. R., et al. (2018). Resistant hypertension: detection, evaluation, and

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management: a scientific statement from the American heart association. *Hypertension* 72 (5), e53–e90. doi:10.1161/HYP.00000000000084

Chambers, J. C., Zhao, J., Terracciano, C. M., Bezzina, C. R., Zhang, W., Kaba, R., et al. (2010). Genetic variation in SCN10A influences cardiac conduction. *Nat. Genet.* 42 (2), 149–152. doi:10.1038/ng.516

Chobanian, A. V., Bakris, G. L., Black, H. R., Cushman, W. C., Green, L. A., Izzo, J. L., Jr., et al. (2003). The seventh report of the Joint national committee on prevention, detection, evaluation, and treatment of high blood pressure: the JNC 7 report. *JAMA* 289 (19), 2560–2572. doi:10.1001/jama.289.19.2560

Conomos, M. P., Miller, M. B., and Thornton, T. A. (2015). Robust inference of population structure for ancestry prediction and correction of stratification in the presence of relatedness. *Genet. Epidemiol.* 39 (4), 276–293. doi:10.1002/gepi. 21896

De Keulenaer, G. W., Wang, Y., Feng, Y., Muangman, S., Yamamoto, K., Thompson, J. F., et al. (2002). Identification of IEX-1 as a biomechanically controlled nuclear factorkappaB target gene that inhibits cardiomyocyte hypertrophy. *Circ. Res.* 90 (6), 690–696. doi:10.1161/01.res.0000012922.40318.05

Dumitrescu, L., Ritchie, M. D., Denny, J. C., El Rouby, N. M., McDonough, C. W., Bradford, Y., et al. (2017). Genome-wide study of resistant hypertension identified from electronic health records. *PLoS One* 12 (2), e0171745. doi:10.1371/journal.pone. 0171745

El Rouby, N., McDonough, C. W., Gong, Y., McClure, L. A., Mitchell, B. D., Horenstein, R. B., et al. (2019). Genome-wide association analysis of common genetic variants of resistant hypertension. *Pharmacogenomics J.* 19 (3), 295–304. doi:10.1038/s41397-018-0049-x

EPACTS (2019). Efficient and parallelizable association container Toolbox. Available from: http://genome.sph.umich.edu/wiki/EPACTS.

Evangelou, E., Warren, H. R., Mosen-Ansorena, D., Mifsud, B., Pazoki, R., Gao, H., et al. (2018). Genetic analysis of over 1 million people identifies 535 new loci associated with blood pressure traits. *Nat. Genet.* 50 (10), 1412–1425. doi:10.1038/s41588-018-0205-x

Fontana, V., McDonough, C. W., Gong, Y., El Rouby, N. M., Sa, A. C., Taylor, K. D., et al. (2014). Large-scale gene-centric analysis identifies polymorphisms for resistant hypertension. *J. Am. Heart Assoc.* 3 (6), e001398. doi:10.1161/JAHA.114.001398

Franceschini, N., Reiner, A. P., and Heiss, G. (2011). Recent findings in the genetics of blood pressure and hypertension traits. *Am. J. Hypertens.* 24 (4), 392–400. doi:10.1038/ ajh.2010.218

Gene (2004). National library of medicine, national center for Biotechnology information. Available at: https://www.ncbi.nlm.nih.gov/gene/.2004.

Giri, A., Hellwege, J. N., Keaton, J. M., Park, J., Qiu, C., Warren, H. R., et al. (2019). Trans-ethnic association study of blood pressure determinants in over 750,000 individuals. *Nat. Genet.* 51 (1), 51–62. doi:10.1038/s41588-018-0303-9

Global Burden of Metabolic Risk Factors for Chronic Diseases Collaboration (2014). Cardiovascular disease, chronic kidney disease, and diabetes mortality burden of cardiometabolic risk factors from 1980 to 2010: a comparative risk assessment. *Lancet Diabetes Endocrinol.* 2 (8), 634–647. doi:10.1016/S2213-8587(14)70102-0

Gupta, A. K., Nasothimiou, E. G., Chang, C. L., Sever, P. S., Dahlof, B., Poulter, N. R., et al. (2011). Baseline predictors of resistant hypertension in the Anglo-Scandinavian Cardiac Outcome Trial (ASCOT): a risk score to identify those at high-risk. *J. Hypertens.* 29 (10), 2004–2013. doi:10.1097/HJH.0b013e32834a8a42

Hirschhorn, J. N., and Altshuler, D. (2002). Once and again-issues surrounding replication in genetic association studies. J. Clin. Endocrinol. Metab. 87 (10), 4438–4441. doi:10.1210/jc.2002-021329

Holm, H., Gudbjartsson, D. F., Arnar, D. O., Thorleifsson, G., Thorgeirsson, G., Stefansdottir, H., et al. (2010). Several common variants modulate heart rate, PR interval and QRS duration. *Nat. Genet.* 42 (2), 117–122. doi:10.1038/ng.511

Hyman, D. J., and Pavlik, V. N. (2001). Characteristics of patients with uncontrolled hypertension in the United States. *N. Engl. J. Med.* 345 (7), 479–486. doi:10.1056/ NEJMoa010273

Irvin, M. R., Sitlani, C. M., Floyd, J. S., Psaty, B. M., Bis, J. C., Wiggins, K. L., et al. (2019). Genome-wide association study of apparent treatment-resistant hypertension in the CHARGE consortium: the CHARGE Pharmacogenetics working group. *Am. J. Hypertens.* 32 (12), 1146–1153. doi:10.1093/ajh/hpz150

Jin, H., Li, D. Y., Chernogubova, E., Sun, C., Busch, A., Eken, S. M., et al. (2018). Local delivery of miR-21 stabilizes fibrous caps in vulnerable atherosclerotic lesions. *Mol. Ther.* 26 (4), 1040–1055. doi:10.1016/j.ymthe.2018.01.011

Jin, Y. R., and Yoon, J. K. (2012). The R-spondin family of proteins: emerging regulators of WNT signaling. *Int. J. Biochem. Cell Biol.* 44 (12), 2278–2287. doi:10.1016/ j.biocel.2012.09.006

Judd, E., and Calhoun, D. A. (2014). Apparent and true resistant hypertension: definition, prevalence and outcomes. *J. Hum. Hypertens.* 28 (8), 463–468. doi:10.1038/jhh.2013.140

Jun, G., Wing, M. K., Abecasis, G. R., and Kang, H. M. (2015). An efficient and scalable analysis framework for variant extraction and refinement from population-scale DNA sequence data. *Genome Res.* 25 (6), 918–925. doi:10.1101/gr.176552.114

Kalna, V., Yang, Y., Peghaire, C. R., Frudd, K., Hannah, R., Shah, A. V., et al. (2019). The transcription factor ERG regulates super-enhancers associated with an endothelial-specific gene expression program. *Circ. Res.* 124 (9), 1337–1349. doi:10.1161/CIRCRESAHA.118.313788

Kelly, T. N., Sun, X., He, K. Y., Brown, M. R., Taliun, S. A. G., Hellwege, J. N., et al. (2022). Insights from a large-scale whole-genome sequencing study of systolic blood pressure, diastolic blood pressure, and hypertension. *Hypertension* 79 (8), 1656–1667. doi:10.1161/HYPERTENSIONAHA.122.19324

Levy, D., Ehret, G. B., Rice, K., Verwoert, G. C., Launer, L. J., Dehghan, A., et al. (2009). Genome-wide association study of blood pressure and hypertension. *Nat. Genet.* 41 (6), 677–687. doi:10.1038/ng.384

Liu, B., Li, N., Zhang, J., Liu, Y., Zhang, M., Hong, Y., et al. (2020). The role of voltage-gated sodium channel 1.8 in the effect of atropine on heart rate: evidence from a retrospective clinical study and mouse model. *Front. Pharmacol.* 11, 1163. doi:10.3389/fphar.2020.01163

Liu, X., White, S., Peng, B., Johnson, A. D., Brody, J. A., Li, A. H., et al. (2016). WGSA: an annotation pipeline for human genome sequencing studies. *J. Med. Genet.* 53 (2), 111–112. doi:10.1136/jmedgenet-2015-103423

Lynch, A. I., Irvin, M. R., Davis, B. R., Ford, C. E., Eckfeldt, J. H., and Arnett, D. K. (2013). Genetic and adverse health outcome associations with treatment resistant hypertension in GenHAT. *Int. J. Hypertens.* 2013, 578578. doi:10.1155/2013/578578

Ma, L., Roman-Campos, D., Austin, E. D., Eyries, M., Sampson, K. S., Soubrier, F., et al. (2013). A novel channelopathy in pulmonary arterial hypertension. *N. Engl. J. Med.* 369 (4), 351–361. doi:10.1056/NEJMoa1211097

Macri, V., Brody, J. A., Arking, D. E., Hucker, W. J., Yin, X., Lin, H., et al. (2018). Common coding variants in SCN10A are associated with the Nav1.8 late current and cardiac conduction. *Circ. Genom Precis. Med.* 11 (5), e001663. doi:10.1161/CIRCGEN.116.001663

Manichaikul, A., Rich, S. S., Allison, M. A., Guagliardo, N. A., Bayliss, D. A., Carey, R. M., et al. (2016). KCNK3 variants are associated with hyperaldosteronism and hypertension. *Hypertension* 68 (2), 356–364. doi:10.1161/HYPERTENSIONAHA.116.07564

McDonough, A. A., Velotta, J. B., Schwinger, R. H., Philipson, K. D., and Farley, R. A. (2002). The cardiac sodium pump: structure and function. *Basic Res. Cardiol.* 97 (1), 119–124. doi:10.1007/s003950200024

Mikhaylova, A. V., McHugh, C. P., Polfus, L. M., Raffield, L. M., Boorgula, M. P., Blackwell, T. W., et al. (2021). Whole-genome sequencing in diverse subjects identifies genetic correlates of leukocyte traits: the NHLBI TOPMed program. *Am. J. Hum. Genet.* 108 (10), 1836–1851. doi:10.1016/j.ajhg.2021.08.007

NCD Risk Factor Collaboration (2019). Long-term and recent trends in hypertension awareness, treatment, and control in 12 high-income countries: an analysis of 123 nationally representative surveys. *Lancet* 394 (10199), 639–651. doi:10.1016/S0140-6736(19)31145-6

Newton-Cheh, C., Johnson, T., Gateva, V., Tobin, M. D., Bochud, M., Coin, L., et al. (2009). Genome-wide association study identifies eight loci associated with blood pressure. *Nat. Genet.* 41 (6), 666–676. doi:10.1038/ng.361

Parmar, P. G., Taal, H. R., Timpson, N. J., Thiering, E., Lehtimaki, T., Marinelli, M., et al. (2016). International genome-wide association study consortium identifies novel loci associated with blood pressure in children and adolescents. *Circ. Cardiovasc Genet.* 9 (3), 266–278. doi:10.1161/ CIRCGENETICS.115.001190

Pfeufer, A., van Noord, C., Marciante, K. D., Arking, D. E., Larson, M. G., Smith, A. V., et al. (2010). Genome-wide association study of PR interval. *Nat. Genet.* 42 (2), 153–159. doi:10.1038/ng.517

Plotnikov, D., Huang, Y., Khawaja, A. P., Foster, P. J., Zhu, Z., Guggenheim, J. A., et al. (2022). High blood pressure and intraocular pressure: a mendelian randomization study. *Invest. Ophthalmol. Vis. Sci.* 63 (6), 29. doi:10.1167/iovs.63.6.29

Sarafidis, P. A., and Bakris, G. L. (2008). Resistant hypertension: an overview of evaluation and treatment. J. Am. Coll. Cardiol. 52 (22), 1749–1757. doi:10.1016/j.jacc. 2008.08.036

Seth, A., and Watson, D. K. (2005). ETS transcription factors and their emerging roles in human cancer. *Eur. J. Cancer* 41 (16), 2462–2478. doi:10.1016/j.ejca.2005.08.013

Shah, A. V., Birdsey, G. M., and Randi, A. M. (2016). Regulation of endothelial homeostasis, vascular development and angiogenesis by the transcription factor ERG. *Vasc. Pharmacol.* 86, 3–13. doi:10.1016/j.vph.2016.05.003

Shuey, M. M., Gandelman, J. S., Chung, C. P., Nian, H., Yu, C., Denny, J. C., et al. (2018). Characteristics and treatment of African-American and European-American patients with resistant hypertension identified using the electronic health record in an academic health centre: a case-control study. *BMJ Open* 8 (6), e021640. doi:10.1136/ bmjopen-2018-021640

Sotoodehnia, N., Isaacs, A., de Bakker, P. I., Dorr, M., Newton-Cheh, C., Nolte, I. M., et al. (2010). Common variants in 22 loci are associated with QRS duration and cardiac ventricular conduction. *Nat. Genet.* 42 (12), 1068–1076. doi:10.1038/ng.716

Takahashi, Y., Yamazaki, K., Kamatani, Y., Kubo, M., Matsuda, K., and Asai, S. (2021). A genome-wide association study identifies a novel candidate locus at the DLGAP1 gene with susceptibility to resistant hypertension in the Japanese population. *Sci. Rep.* 11 (1), 19497. doi:10.1038/s41598-021-98144-z

Taliun, D., Harris, D. N., Kessler, M. D., Carlson, J., Szpiech, Z. A., Torres, R., et al. (2021). Sequencing of 53,831 diverse genomes from the NHLBI TOPMed Program. *Nature* 590 (7845), 290–299. doi:10.1038/s41586-021-03205-y

Vongpatanasin, W. (2014). Resistant hypertension: a review of diagnosis and management. JAMA 311 (21), 2216-2224. doi:10.1001/jama.2014.5180

West, J. D., Austin, E. D., Rizzi, E. M., Yan, L., Tanjore, H., Crabtree, A. L., et al. (2021). KCNK3 mutation causes altered immune function in pulmonary arterial hypertension patients and mouse models. *Int. J. Mol. Sci.* 22 (9), 5014. doi:10.3390/ijms22095014

Wijkman, M. O., Malachias, M. V. B., Claggett, B. L., Cheng, S., Matsushita, K., Shah, A. M., et al. (2021). Resistance to antihypertensive treatment and long-term risk: the Atherosclerosis Risk in Communities study. *J. Clin. Hypertens. (Greenwich)* 23 (10), 1887–1896. doi:10.1111/jch.14269

Willer, C. J., Li, Y., and Abecasis, G. R. (2010). METAL: fast and efficient metaanalysis of genomewide association scans. *Bioinformatics* 26 (17), 2190–2191. doi:10. 1093/bioinformatics/btq340

Wisedchaisri, G., and Gamal El-Din, T. M. (2022). Druggability of voltage-gated sodium channels-exploring old and new drug receptor sites. *Front. Pharmacol.* 13, 858348. doi:10.3389/fphar.2022.858348

Yasukochi, Y., Sakuma, J., Takeuchi, I., Kato, K., Oguri, M., Fujimaki, T., et al. (2018). Identification of three genetic variants as novel susceptibility loci for body mass index in a Japanese population. *Physiol. Genomics* 50 (3), 179–189. doi:10.1152/ physiolgenomics.00117.2017

Zhao, Y., and Rahmouni, K. (2022). BBSome: a new player in hypertension and other cardiovascular risks. *Hypertension* 79 (2), 303–313. doi:10.1161/ HYPERTENSIONAHA.121.17946

Zhou, W., Nielsen, J. B., Fritsche, L. G., Dey, R., Gabrielsen, M. E., Wolford, B. N., et al. (2018). Efficiently controlling for case-control imbalance and sample relatedness in large-scale genetic association studies. *Nat. Genet.* 50 (9), 1335–1341. doi:10.1038/s41588-018-0184-y

APPENDIX A

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