UC Davis UC Davis Previously Published Works

Title

Metagenomic Sequencing of Two Salton Sea Microbiomes

Permalink https://escholarship.org/uc/item/90w1p134

Journal

Microbiology Resource Announcements, 2(1)

ISSN 2576-098X

Authors

Hawley, Erik R Schackwitz, Wendy Hess, Matthias

Publication Date

2014-02-27

DOI

10.1128/genomea.01208-13

Peer reviewed



Metagenomic Sequencing of Two Salton Sea Microbiomes

Erik R. Hawley,^a Wendy Schackwitz,^b Matthias Hess^{a,c,d,e}

Washington State University Tri-Cities, Richland, Washington, USA^a; DOE Joint Genome Institute, Genome Analysis Group, Walnut Creek, California, USA^b; DOE Joint Genome Institute, Microbial Genomics Group, Walnut Creek, California, USA^c; Pacific Northwest National Laboratory, Chemical & Biological Process Development Group, Richland, Washington, USA^d; Washington State University, Pullman, Washington, USA^e

The Salton Sea is the largest inland body of water in California, with salinities ranging from brackish freshwater to hypersaline. The lake experiences high nutrient input, and its surface water is exposed to temperatures up to 40°C. Here, we report the community profiles associated with surface water from the Salton Sea.

Received 11 December 2013 Accepted 19 December 2013 Published 23 January 2014 Citation Hawley ER, Schackwitz W, Hess M. 2014. Metagenomic sequencing of two Salton Sea microbiomes. Genome Announc. 2(1):e1208-13. doi:10.1128/genomeA.01208-13. Copyright © 2014 Hawley et al. This is an open-access article distributed under the terms of the Creative Commons Attribution 3.0 Unported license. Address correspondence to Matthias Hess, mhess@lbl.gov.

ypersaline environments are found around the globe (1) and are often populated by dense communities of halophilic prokaryotes (2). The Salton Sea, a eutrophic lake with a salinity of ~48 g liter⁻¹, is the largest lake in California (3). The high nutrient loading of its influx waters facilitates algal blooms throughout the year (4), ultimately expediting the formation of anaerobic conditions associated with the microbially mediated formation of chemical species that have a negative impact on the food web (3, 5). To enhance our understanding of how environmental parameters affect the flora and fauna of the Salton Sea and its surrounding region, a thorough knowledge base of the Salton Sea microbiology is essential. Here, we report the community profile of two microbiomes associated with surface water from the Salton Sea.

Water samples were collected on 29 September and 1 October 2009 at site 1 (33°30.253'N 115°54.982'W) and site 2 (33°10.522'N 115°38.274'W), respectively. The water temperatures and pH levels of the samples were 29.9°C and 23.6°C and pH 8.09 and pH 8.20 from sites 1 and site 2, respectively. DNA was extracted using the FastDNA SPIN kit for soil from MP Biomedicals, according to the manufacturer's protocol. The V6 to V8 region of the bacterial 16S rRNA gene was amplified using the primer set 926f/1392r and sequenced using the Roche 454 FLX+ platform (Research and Testing Laboratory, Lubbock, TX). The raw pyrosequence reads were quality filtered and analyzed using QIIME version 1.7.0 (6). Prior to filtering, the sequencing primers and barcodes were removed, allowing 1.5 mismatches to the barcode and 2 mismatches to the primer. The sequences were removed from analysis if they contained homopolymers >6 bp, were <200 bp in length, contained a quality score <25, or if they were found to be chimeric. The sequences were clustered into operational taxonomic units (OTUs) at the 97% sequence identity level using UCLUST (7), and the most abundant sequence of each OTU was chosen as a representative. The OTU representative sequences were aligned using PyNAST (8) and then filtered to remove common gaps. The reference sequences of each OTU were taxonomically classified using the RDP Classifier (9) with an 80% confidence rating against the Greengenes database (10).

A total of 1,484 and 3,341 high-quality sequences, representing 492 and 1,043 distinct OTUs, were obtained from sites 1 and 2,

respectively. A total of 46 distinct phyla were identified in this study, with 34 of the phyla shared between the two samples. The two most abundant phyla detected were *Proteobacteria* (50.0% at site 1 and 52.1% at site 2) and *Bacteroidetes* (11.19% at site 1 and 8.50% at site 2). *Spirochaetes* (5.9%), *Planctomycetes* (5.1%), and unclassified bacteria (4.3%) were the next-most-abundant phyla for site 1, while unclassified bacteria (7.5%), *Planctomycetes* (4.2%), and *Cyanobacteria* (4.2%) were the next-most-abundant phyla for site 2.

Collectively, our data reveal a phylogenetic diversity and variance within the microbial communities of geospatially distinct sites from California's largest lake, the Salton Sea.

Nucleotide sequence accession number. The DNA sequences from this metagenomic project have been deposited in the NCBI Short Read Archive under the accession no. SRP033722.

ACKNOWLEDGMENT

M.H., E.R.H., and the work performed in the laboratory of M.H. were funded by Washington State University.

REFERENCES

- Andrei AŞ, Banciu HL, Oren A. 2012. Living with salt: metabolic and phylogenetic diversity of archaea inhabiting saline ecosystems. FEMS Microbiol. Lett. 330:1–9. http://dx.doi.org/10.1111/j.1574-6968.2012. 02526.x.
- Oren A. 2002. Molecular ecology of extremely halophilic archaea and bacteria. FEMS Microbiol. Ecol. 39:1–7. http://dx.doi.org/10.1111/j.1574 -6941.2002.tb00900.x.
- VillaRomero JF, Kausch M, Pallud C. 2013. Selenate reduction and adsorption in littoral sediments from a hypersaline California lake, the Salton Sea. Hydrobiologia 709:129–142. http://dx.doi.org/10.1007/s1075 0-013-1443-7.
- Carmichael WW, Li R. 2006. Cyanobacteria toxins in the Salton Sea. Saline Syst. 2:5. http://dx.doi.org/10.1186/1746-1448-2-5.
- Reese BK, Anderson MA, Amrhein C. 2008. Hydrogen sulfide production and volatilization in a polymictic eutrophic saline lake, Salton Sea, California. Sci. Total Environ. 406:205–218. http://dx.doi.org/10.1016/j .scitotenv.2008.07.021.
- 6. Caporaso JG, Kuczynski J, Stombaugh J, Bittinger K, Bushman FD, Costello EK, Fierer N, Peña AG, Goodrich JK, Gordon JI, Huttley GA, Kelley ST, Knights D, Koenig JE, Ley RE, Lozupone CA, McDonald D, Muegge BD, Pirrung M, Reeder J, Sevinsky JR, Turnbaugh PJ, Walters WA, Widmann J, Yatsunenko T, Zaneveld J, Knight R. 2010. QIIME

allows analysis of high-throughput community sequencing data. Nat. Methods 7:335–336. http://dx.doi.org/10.1038/nmeth.f.303.

- 7. Edgar RC. 2010. Search and clustering orders of magnitude faster than BLAST. Bioinformatics 26:2460-2461. http://dx.doi.org/10.1093 /bioinformatics/btq461.
- Caporaso JG, Bittinger K, Bushman FD, DeSantis TZ, Andersen GL, Knight R. 2010. PyNAST: a flexible tool for aligning sequences to a template alignment. Bioinformatics 26:266–267. http://dx.doi.org/10.1093 /bioinformatics/btp636.
- Wang Q, Garrity GM, Tiedje JM, Cole JR. 2007. Naive Bayesian classifier for rapid assignment of rRNA sequences into the new bacterial taxonomy. Appl. Environ. Microbiol. 73:5261–5267. http://dx.doi.org/10.1128/AEM .00062-07.
- McDonald D, Price MN, Goodrich J, Nawrocki EP, DeSantis TZ, Probst A, Andersen GL, Knight R, Hugenholtz P. 2012. An improved Greengenes taxonomy with explicit ranks for ecological and evolutionary analyses of bacteria and archaea. ISME J. 6:610–618. http://dx.doi.org/10.103 8/ismej.2011.139.