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The Microanatomy of the Leukemic Stem Cell Niche in Murine Chronic Myelogenous Leukemia

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Abstract		
Objectives and background: Constituents of the bone marrow microenvironment (BMM) Influence the proliferation, differentiation and location of hematopoletic stem and progenitor cells (HSPC). Dependent on their maturation stage, different subsets of HSPC are localized at distinct sites in the BMM. This location depends on HSPC-intrinsic, as well as HSPC- extrinsic factors. The BMM protects leukemic stem cells (LSC) from treatment with tyrosine kinase inhibitors or chemotherapy. We, therefore, investigated the microanantomy of the LSC niche hypothesizing that it may differ from the normal HSPC niche.	Volume: 124 Issue: 21 Pages: 351 - 351 DOI: http://dx.doi.org/	 Save to My Folders Request Permission Share
models of <i>BCR-ABL1</i> *chronic myelogenous leukemia (CML) and B-cell acute lymphoblastic leukemia (B-ALL).	Article	
Results: We show here that <i>BCR-ABL1</i> *Lin ⁻ c-Klt*Sca-1* (LKS) CD150*CD48 ⁻ (LKS SLAM) cells, which harbor the LSC fraction in the CML model, homed to locations further away from the endosteum than their normal counterparts.	Info & Metrics E-Letters	
standard of care in CML, reversed this phenotype and the cells were found closer to the endosteum. Native BCR-ABL1, as well as the imatinib-resistant BCR-ABL1 point mutants BCR-ABL1 ^{Y253F} , BCR-ABL1 ^{E255K} , BCR-ABL1 ^{T3151} and BCR-ABL1 ^{M351T} had similar intrinsic catalytic activity, but the BCR-ABL1 ^{Y253F} , BCR-ABL1 ^{E255K} , and BCR-ABL1 ^{F3151} mutants increased the IL-3-independent proliferative capacity of	Related Articles No related articles four	- Id.
32D cells relative to native BCR-ABL1. BCR-ABL1 ^{Y253F} and BCR-ABL1 ^{M35IT}		

caused increased transformation of primary BM B-lymphoid progenitors in vitro and led to accelerated induction of B-ALL in mice. In the CML model, *BCR-ABL1^{Y253F}* and *BCR-ABL1^{T315I}*induced myeloproliferative neoplasia with shortened survival and features of accelerated phase disease compared to native *BCR-ABL1*, whereas *BCR-ABL1^{T315I}*LKS cells homed closer to osteoblastic cells than LKS cells expressing native *BCR-ABL1*.

Sequential in vivo tracking of leukemic progenitor growth by IVM showed a similar nadir in the number of cells per leukemic cell 'nest' 11 days after irradiation and IV transplantation in recipients of *DsRed*BCR-ABL1*+ or empty vector control-transduced bone marrow. However, between days 18-25 after transplantation there was a significant increase in the number of cells per leukemic cell 'nest' compared to the empty vector control group. Sequential immunohistochemistry and TUNEL assays of leukemic bone sections in imatinib- or vehicle-treated recipient mice with CML showed that initial *BCR-ABL1*+ growth tends to occur at locations further away from the endosteum, whereas erythroid islands were found closer to the endosteum and trabeculae. Apoptosis in response to imatinib appeared most prominent in the metaphysis. Lastly, we could demonstrate by IVM in the CML model that treatment of mice with a combination of imatinib plus granulocyte colony-stimulating factor led to 'emptying' of the LSC niche and superior eradication of *BCR-ABL1*+ leukemic cells compared to treatment with imatinib alone.

Conclusions: In summary, these data suggest that the microanatomy of the LSC niche in CML differs from the normal hematopoietic niche. *BCR-ABL1* mutation status may affect the positioning of CML LSC in the microenvironment, and location in the niche may be altered pharmacologically, suggesting that niche location may influence clinical outcome.

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- □* Asterisk with author names denotes non-ASH members.
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