

UCLA

UCLA Previously Published Works

Title

Population-based targeted sequencing of 54 candidate genes identifies PALB2 as a susceptibility gene for high-grade serous ovarian cancer

Permalink

<https://escholarship.org/uc/item/5g21t5q9>

Journal

Journal of Medical Genetics, 58(5)

ISSN

0022-2593

Authors

Song, Honglin

Dicks, Ed M

Tyrer, Jonathan

et al.

Publication Date

2021-05-01

DOI

10.1136/jmedgenet-2019-106739

Peer reviewed



OPEN ACCESS

Original research

# Population-based targeted sequencing of 54 candidate genes identifies *PALB2* as a susceptibility gene for high-grade serous ovarian cancer

Honglin Song,<sup>1</sup> Ed M Dicks,<sup>1</sup> Jonathan Tyrer,<sup>1</sup> Maria Intermaggio,<sup>2</sup> Georgia Chenevix-Trench,<sup>3</sup> David D Bowtell,<sup>4</sup> Nadia Traficante,<sup>5</sup> AOCS Group,<sup>6,7</sup> James Brenton,<sup>8</sup> Teodora Goranova,<sup>8</sup> Karen Hosking,<sup>1</sup> Anna Piskorz,<sup>8</sup> Elke van Oudenhove,<sup>9</sup> Jen Doherty,<sup>10</sup> Holly R Harris,<sup>11,12</sup> Mary Anne Rossing,<sup>11,12</sup> Matthias Duerst,<sup>13</sup> Thilo Dork,<sup>14</sup> Natalia V Bogdanova,<sup>15,16</sup> Francesmary Modugno,<sup>17,18</sup> Kirsten Moysich,<sup>19</sup> Kunle Odunsi,<sup>20</sup> Roberta Ness,<sup>21</sup> Beth Y Karlan,<sup>22,23</sup> Jenny Lester,<sup>22,23</sup> Allan Jensen,<sup>24</sup> Susanne Krüger Kjaer,<sup>25</sup> Estrid Høgdall,<sup>24,26</sup> Ian G Campbell,<sup>5,27</sup> Conxi Lázaro,<sup>28</sup> Miguel Angel Pujara,<sup>29</sup> Julie Cunningham,<sup>30</sup> Robert Vierkant,<sup>31</sup> Stacey J Winham,<sup>31</sup> Michelle Hildebrandt,<sup>32</sup> Chad Huff ,<sup>32</sup> Donghui Li,<sup>32</sup> Xifeng Wu,<sup>32</sup> Yao Yu,<sup>32</sup> Jennifer B Permuth,<sup>33</sup> Douglas A Levine,<sup>34,35</sup> Joellen M Schildkraut,<sup>36</sup> Marjorie J Riggan,<sup>37</sup> Andrew Berchuck,<sup>37</sup> Penelope M Webb,<sup>38</sup> OPAL Study Group,<sup>38</sup> Cezary Cybulski ,<sup>39</sup> Jacek Gronwald,<sup>39</sup> Anna Jakubowska,<sup>39,40</sup> Jan Lubinski,<sup>39</sup> Jennifer Alsop,<sup>1</sup> Patricia Harrington,<sup>1</sup> Isaac Chan,<sup>2</sup> Usha Menon,<sup>41</sup> Celeste L Pearce,<sup>42</sup> Anna H Wu,<sup>43</sup> Anna de Fazio,<sup>44,45</sup> Catherine J Kennedy,<sup>44,45</sup> Ellen Goode,<sup>46</sup> Susan Ramus,<sup>2,47</sup> Simon Gayther,<sup>48</sup> Paul Pharoah <sup>49</sup>

► Additional material is published online only. To view please visit the journal online (<http://dx.doi.org/10.1136/jmedgenet-2019-106739>).

For numbered affiliations see end of article.

## Correspondence to

Dr Paul Pharoah, Department of Public Health and Primary Care, University of Cambridge, Cambridge CB2 1TN, UK; pp10001@medschl.cam.ac.uk

HS and EMD contributed equally.  
SR, SG and PP contributed equally.

Received 26 November 2019  
Revised 13 March 2020  
Accepted 12 May 2020  
Published Online First 16 June 2020



© Author(s) (or their employer(s)) 2021. Re-use permitted under CC BY. Published by BMJ.

**To cite:** Song H, Dicks EM, Tyrer J, et al. *J Med Genet* 2021;**58**:305–313.

## ABSTRACT

**Purpose** The known epithelial ovarian cancer (EOC) susceptibility genes account for less than 50% of the heritable risk of ovarian cancer suggesting that other susceptibility genes exist. The aim of this study was to evaluate the contribution to ovarian cancer susceptibility of rare deleterious germline variants in a set of candidate genes.

**Methods** We sequenced the coding region of 54 candidate genes in 6385 invasive EOC cases and 6115 controls of broad European ancestry. Genes with an increased frequency of putative deleterious variants in cases versus controls were further examined in an independent set of 14 135 EOC cases and 28 655 controls from the Ovarian Cancer Association Consortium and the UK Biobank. For each gene, we estimated the EOC risks and evaluated associations between germline variant status and clinical characteristics.

**Results** The ORs associated for high-grade serous ovarian cancer were 3.01 for *PALB2* (95% CI 1.59 to 5.68;  $p=0.00068$ ), 1.99 for *POLK* (95% CI 1.15 to 3.43;  $p=0.014$ ) and 4.07 for *SLX4* (95% CI 1.34 to 12.4;  $p=0.013$ ). Deleterious mutations in *FBXO10* were associated with a reduced risk of disease (OR 0.27, 95% CI 0.07 to 1.00,  $p=0.049$ ). However, based on the Bayes false discovery probability, only the association for *PALB2* in high-grade serous ovarian cancer is likely to represent a true positive.

**Conclusions** We have found strong evidence that carriers of *PALB2* deleterious mutations are at increased risk of high-grade serous ovarian cancer. Whether the magnitude of risk is sufficiently high to warrant the

inclusion of *PALB2* in cancer gene panels for ovarian cancer risk testing is unclear; much larger sample sizes will be needed to provide sufficiently precise estimates for clinical counselling.

## INTRODUCTION

Rare, predicted deleterious variants in multiple genes have been shown to be associated with a moderate to high risk of epithelial ovarian cancer (EOC). These include the DNA double strand break repair genes *BRCA1*,<sup>1</sup> *BRCA2*,<sup>2</sup> *BRIP1*,<sup>3</sup> *RAD51C*, and *RAD51*,<sup>4</sup> and the mismatch repair genes *MSH2* and *MSH6*.<sup>5,6</sup> *ANKRD11*, *FANCM*, *PALB2* and *POLE* have recently been reported as possible susceptibility genes.<sup>7–9</sup> Multiple common variants conferring weaker risk effects have also been identified,<sup>10–17</sup> some of which modify EOC risk in carriers of more highly penetrant gene mutations.<sup>18,19</sup>

EOC is heterogeneous with five main histotypes: high-grade serous (HGSOC), low-grade serous, endometrioid, clear cell and mucinous ovarian cancer. These have different clinical characteristics and outcomes and are characterised by different germline and somatic genetic changes that result in the perturbation of different molecular pathways. For example, germline mutations in DNA double break repair genes predispose to HGSOC while germline mutations in mismatch repair genes increase risk of the endometrioid and clear cell histotypes.<sup>6</sup>

The known susceptibility alleles account for less than 50% of the excess familial risk of ovarian cancer, suggesting that other susceptibility genes and alleles exist.<sup>15</sup> The unexplained genetic component of risk is likely to be made up of a combination of common genetic variants conferring weak effects and uncommon alleles conferring weak to moderate relative risks (less than 10-fold).

The aim of this study was to identify additional ovarian cancer susceptibility genes using case-control sequencing of candidate genes identified through various approaches including their known function in pathways that are associated with ovarian cancer development and from whole exome sequencing studies (WES) of ovarian cancer cases that have identified putative deleterious mutations in genes not previously evaluated for EOC risk.

## MATERIAL AND METHODS

### Selection of candidate genes

#### Genes based on known biological function

As several EOC susceptibility genes are involved in DNA double-strand break repair and Fanconi anaemia (FA),<sup>8</sup> we selected genes involved in these pathways. FA is a rare genetic disease characterised by chromosomal instability, hypersensitivity to DNA crosslinking agents, defective DNA repair, severe bone marrow failure, cancer susceptibility and many congenital defects. To date, 22 FA genes have been identified, of which eight have previously been evaluated in ovarian cancer case-control studies:<sup>3 4 6 8</sup> *FANCD1*, *FNACJ* (*BRIP1*), *FANCL*, *FANCN* (*PALB2*), *FANCM*, *FANCO* (*RAD51C*), *FANCS* (*BRCA1*) and *FANCV* (*MAD2L2*). We selected nine FA genes not previously studied in ovarian cancer: *FANCA*, *FANCB*, *FANCC*, *FANCD2*, *FANCE*, *FANCG*, *FANCI*, *FANCP* (*SLX4*), *FANCW*. We also included *FANCN* (*PALB2*), which has been studied previously in ovarian cancer<sup>3 9 20–22</sup> but its association with EOC risk is equivocal. Eight candidate genes involved in other aspects of DNA repair were also included: *ALKBH3*, *CHEK2*, *GTF2H4*, *POLE*, *POLK*, *RDM1* and *XRCC1*.

#### Genes from whole exome sequencing studies (WES)

Twelve genes (*BUB1B*, *C5orf28*, *C6*, *DNAJB4*, *EXO1*, *LIG4*, *MKNK2*, *MMRN1*, *PARP1*, *RAD52*, *SMC1A* and *SNRNP200*) were selected from WES analysis of EOC cases where putative deleterious (truncating) mutations were identified at a greater frequency in cases compared with publicly available WES data from controls reported by the NHLBI GO Exome Sequencing Project and The Exome Aggregation Consortium databases (<http://exac.broadinstitute.org>). Germline WES data for EOC cases were available for 412 HGSOC cases from the Cancer Genome Atlas ovarian cancer study; 513 ovarian cancer cases from an Australian case series 6; 97 familial non-*BRCA1/BRCA2* ovarian cancer cases from Gilda Radner Familial Ovarian Cancer Registry and 54 ovarian cancer cases from the UK Familial Ovarian Cancer Registry.

Four genes from these WES studies (*GANC*, *KNTC1*, *PSG6* and *UPK2*) were selected because more than one family member diagnosed with ovarian cancer from 10 familial cases carried the same truncating mutation in one of these genes.

Finally, 21 genes were selected from analyses of several other unpublished EOC WES studies (personal communications) where the frequency of truncating mutations was greater in cases compared with controls. These genes were *ANAPC2*, *CNKSRI1*, *DUOX1*, *FBXO10*, *NAT10*, *OSGIN1*, *PAK4*, *PHF20L1*,

*PIK3C2G*, *PTGER3*, *PTX3*, *RAD54B*, *RECQL*, *RIPK3*, *RNASEL*, *SMG5*, *SPHK1*, *SULT1C2*, *UHRF2*, *WNT5A* and *ZFH3*.

### Study subjects

We used case-control data from targeted sequencing, exome and array-based genotyping.

#### Targeted sequencing

We included 5914 EOC cases and 5479 controls of European ancestries from 19 studies—13 case-control studies, 1 familial ovarian cancer study from Poland, 2 clinical trials and 3 case-only studies (online supplementary table 1).<sup>14</sup> HGSOC cases were preferentially plated out for sequencing where possible.

#### Exome sequencing

We extracted data on the 54 candidate genes from 829 case and 913 controls from two ovarian cancer case-control studies (MDA<sup>23–25</sup> and NCO<sup>14</sup>) for which whole exome sequence data were available (online supplementary table 1).

#### Variants from genotyping array data

For genes that reached nominal significance in the combined analysis of the targeted sequencing and exome sequencing data, we extracted genotypes of any deleterious variants included on the OncoArray and UK Biobank Axiom Array. These two arrays were used to genotype up to 18936 controls and 13288 cases from the Ovarian Cancer Association Consortium (OCAC),<sup>15</sup> 9725 controls and 858 cases from UK Biobank GWAS (<https://www.ukbiobank.ac.uk/>), respectively. Samples overlapped with the sequencing studies were excluded from the analysis.

All studies had ethics committee approval, and all participants provided informed consent.

### Sequencing methods

Target sequence enrichment followed by sequencing was performed on the coding sequence and splice-sites of *ALKBH3*, *ANAPC2*, *BUB1B*, *C5orf28*, *C6*, *CHEK2*, *CNKSRI1*, *DNAJB4*, *DUOX1*, *EXO1*, *FANCA*, *FANCB*, *FANCC*, *FANCD2*, *FANCE*, *FANCG*, *FANCI*, *FBXO10*, *GANC*, *GTF2H4*, *KNTC1*, *LIG4*, *MKNK2*, *MMRN1*, *NAT10*, *OSGIN1*, *PAK4*, *PALB2*, *PARP1*, *PHF20L1*, *PIK3C2G*, *POLE*, *POLK*, *PSG6*, *PTGER3*, *PTX3*, *RAD52*, *RAD54B*, *RDM1*, *RECQL*, *REV3L*, *RIPK3*, *RNASEL*, *SLX4*, *SMC1A*, *SMG5*, *SNRNP200*, *SPHK1*, *SULT1C2*, *UHRF2*, *UPK2*, *WNT5A*, *XRCC1* and *ZFH3*. The target sequence was identified from the NCBI Reference Sequence Database<sup>48.48</sup> Fluidigm access arrays as previously described.<sup>6</sup> A total of 1663 amplicons were designed to cover the 159kb target region. Libraries were sequenced using 150bp paired-end sequencing on the Illumina HiSeq4000 or HiSeq2500.

Sequencing reads were demultiplexed and then aligned against the human genome reference sequence (hg19) using the Burrows-Wheeler Aligner.<sup>26</sup> The Genome Analysis Toolkit<sup>27</sup> was used for base quality-score recalibration, local indel realignment and variant calling. Finally, ANNOVAR<sup>28</sup> was used for variant annotation. Variants were called if (1) genotype information was available from a chip genotype for that sample or (2) the variants were presented in more than one amplicon or (3) read depth  $\geq 15$  and alternate allele frequency  $\geq 40\%$  or (4) read depth  $\geq 100$  and alternate allele frequency  $\geq 25\%$ . These thresholds were defined using the results from sequencing of positive controls with known variants and genotype information from chip array genotyping of overlapping samples.

**Table 1** Frequency of mutations and estimated risk of EOC in candidate genes ( $p < 0.05$ ) from targeted sequencing and exome sequencing

Set*	Histotype	Gene	Controls		Cases		Or (95% CI)	P value
			No.	%	No.	%		
TS	Overall	<i>POLK</i>	9	0.17	29	0.52	3.04 (1.43 to 6.43)	0.0037
		<i>PALB2</i>	6	0.12	19	0.34	3.10 (1.23 to 7.78)	0.016
		<i>SLX4</i>	4	0.08	13	0.23	3.08 (1.00 to 9.48)	0.0049
		<i>FBXO10</i>	9	0.17	3	0.053	0.30 (0.08 to 1.11)	0.071
		Non-carrier	5174	99.5	5492	98.8		
HGSOC	HGSOC	<i>POLK</i>	9	0.17	27	0.53	3.17 (1.48 to 6.79)	0.003
		<i>PALB2</i>	6	0.12	18	0.35	3.30 (1.30 to 8.38)	0.012
		<i>SLX4</i>	4	0.08	13	0.25	3.51 (1.13 to 10.9)	0.029
		<i>FBXO10</i>	9	0.17	3	0.059	0.32 (0.09 to 1.18)	0.086
		Non-carrier	5174	99.5	5062	98.8		
ES	Overall	<i>POLK</i>	7	0.77	6	0.72	0.94 (0.32 to 2.82)	0.92
		<i>PALB2</i>	2	0.22	3	0.36	1.65 (0.28 to 9.93)	0.58
		<i>SLX4</i>	0	0	2	0.24	NA	
		<i>FBXO10</i>	1	0.11	0	0		
		Non-carrier	903	98.9	818	98.7		
HGSOC	HGSOC	<i>POLK</i>	7	0.77	6	0.72	0.94 (0.32 to 2.82)	0.92
		<i>PALB2</i>	2	0.22	3	0.36	1.66 (0.28 to 9.94)	0.58
		<i>SLX4</i>	0	0	2	0.24	NA	
		<i>FBXO10</i>	1	0.11	0	0	NA	
		Non-carrier	903	98.9	817	98.7		

\*TS: targeted sequencing; ES: exome sequencing.

EOC, epithelial ovarian cancer; HGSOC, high-grade serous ovarian cancer; OCAC, Ovarian Cancer Association Consortium.

We excluded 356 cases and 269 controls because <80% of the target sequence bases had a read depth of at least 15. The average percentage coverage of the genes at 15X read depth ranged from 64% to 99% (online supplementary table 2). The mean sequencing depth for these genes ranged from 130 (IQR 104–152) to 432 (IQR 364–492). Concordance for 111 duplicate pairs was 98% (7384 concordant variants out of total 7572 variants called).

For the exome sequencing, sonication fragmentation was used to fragment DNA samples. Fragments with an average size of 200 bp were selected to generate libraries for sequencing. Agilent SureSelect Clinical Research Exome (CRE) v1 was used for exome enrichment and sequencing was performed on an Illumina HiSeq 4000 using 2×150 bp paired-end reads. Cutadapt (<https://doi.org/10.14806/ej.17.1.200>) was used to locate and remove residual adapters in reads. FLASH (Fast Length Adjustment of SHort reads)<sup>29</sup> was used to merge the overlapped paired-end reads into one read, using default parameters. Reference genome alignment and joint genotype calling according to a pipeline described in Yu *et al.*<sup>30</sup> The coding sequences and splice sites of all 54 genes were extracted. Fifty-three genes with 100% average coverage at 10X were included in the analysis. *GTF2H4* was excluded from the analysis, as the average coverage was only 43%.

Deleterious variants were defined as those predicted to result in protein truncation (frameshift indel, splice site, nonsense mutations and start loss) or predicted to be deleterious and/or likely deleterious by Clinvar.<sup>31</sup> Any exonic single nucleotide variants within 3 bp of the exon-intron boundary and any intronic variants within 20 bp of the exon-intron boundary at the 5-prime end, and 6 bp at the 3-prime end, were evaluated using the software MaxEntScan to identify those most likely to disrupt splicing.<sup>32</sup> Variants with a MaxEntScan score that decreased by more than 40% compared with the reference sequence and having a reference sequence score  $\geq 3$  were considered

deleterious. Sequencing alignments were confirmed by visual inspection using the Integrative Genomic Viewer.<sup>33</sup>

## Statistical methods

### Risk estimation and genotype-phenotype analyses

We used a simple burden test for association between deleterious variants and ovarian cancer risk on a gene-by-gene basis. The burden test was based on unconditional logistic regression adjusted for country (Australia, Denmark, German, Poland, the UK and the USA) and sequencing method (targeted sequencing or exome sequencing). ORs and associated 95% CI were calculated.

### Missense variant analyses

We also identified multiple rare (minor allele frequency <1%) missense variants that have an unknown functional effect on the protein. We used the rare admixture likelihood burden test<sup>34</sup> to test these variants for association. We excluded any missense variants classified as deleterious and classified the remaining variants by whether or not they are predicted to have a damaging effect on protein function by two out of three prediction tools—SIFT (score <0.05),<sup>35</sup> polyphen-2<sup>36</sup> (classified as probably damaging or damaging) and Provean<sup>37</sup> (score  $\leq -2.5$ ). Subjects with a missense variant call rate less than 80% and variants with a call rate less than 80% or with genotype frequencies inconsistent with Hardy-Weinberg equilibrium ( $p < 10^{-5}$ ) were excluded.

## RESULTS

### Germline deleterious mutations in ovarian cancer cases and controls

Sequencing results were available for 6385 EOC cases and 6115 controls after quality control analysis. The characteristics of these individuals by study are summarised in online supplementary table 1. Most EOC cases were serous histotype ( $n=6304$ , 98.7%), of which 5951 were the HGSOC histotype (93.2%).

**Table 2** Frequency of mutations and estimated risk of EOC in candidate genes for validation chip genotyping data

Set*	Histotype	Gene	Controls		Cases		OR (95% CI)	P value
			No.	%	No.	%		
OCAC	Overall	<i>PALB2</i>	6	0.03	11	0.08	2.10 (0.74 to 5.94)	0.16
		Non-carrier	18930	99.97	13277	99.9		
	HGSOC	<i>PALB2</i>	6	0.03	6	0.097	3.48 (1.10 to 11.1)	0.035
Biobank	Overall	<i>PALB2</i>	11	0.11	3	0.35	3.12 (0.87 to 11.2)	0.081
		<i>POLK</i>	29	0.30	2	0.23	0.78 (0.19 to 3.29)	0.74
		Non-carrier	9685	99.6	853	99.4		
	HGSOC†	<i>PALB2</i>	11	0.11	1	0.28	2.49 (0.32 to 19.4)	0.38
		<i>POLK</i>	29	0.30	1	0.28	0.92 (0.12 to 6.74)	0.93
		Non-carrier	9685	99.6	361	99.4		

\*OCAC: OCAC sample genotype on the OncoArray; Biobank: genotype from UK Biobank Axiom Array.

†Information on tumour grade was not available for UK Biobank cases, all the serous cases in UK Biobank were assumed to be HGSOC. EOC, epithelial ovarian cancer.

We identified 629 unique, putative-deleterious variants (online supplementary table 3) in 1051 ovarian cancer cases (967 high-grade serous histotype) and 964 controls. There was a nominally significant higher frequency of mutations in cases compared with controls for *POLK*, *PALB2* and *SLX4* and a lower frequency of mutations in cases compared with controls for *FBXO10* (table 1). The associated ORs are shown in table 1—for *POLK*, *PALB2* and *SLX4* the effect size was slightly larger for HGSOC. The frequency of deleterious variants in the other genes was similar in cases compared with controls (online supplementary table 4). Given the evidence for association of multiple FA genes with EOC risk, we also carried out a burden test to compare the frequency of deleterious variants in any of the eight genes which were not significantly associated with ovarian cancer risks individually (*FANCA*, *FANCB*, *FANCC*, *FANCD2*, *FANCE*, *FANCG*, *FANCI* and *FANCL*). A combined analysis will have greater power if multiple genes were associated but the effect sizes too small to detect individually. There was no significant difference in the frequency of deleterious variants in cases (96/6184, 1.6%) and controls (85/6089, 1.4%) (p=0.50).

**Validation analyses in ovarian cancer case-control studies**

We also evaluated risk associations between deleterious variants in *POLK*, *PALB2*, and *SLX4* with EOC risk based on germline genotyping data for 13 277 EOC cases and 18 930 controls from OCAC and for 858 EOC cases and 9725 controls and from UK Biobank. For OCAC samples, data were available for six deleterious non-monomorphic variants in *PALB2*; for UK Biobank samples, data were available for seven *PALB2* and one *POLK* deleterious variants (table 2, list of variants in online supplementary table 5).

In OCAC case-control analyses, *PALB2* variants showed a non-significant increased risk of EOC (OR 2.10, 95% CI 0.74 to

5.94, p=0.16). The strength of this association increased when the analysis was restricted to 6181 HGSOC cases (OR 3.48, 95% CI 1.10 to 11.1, p=0.035). In UK Biobank, we observed a weak association for *PALB2* mutations with EOC risk (OR 3.12, 95% CI 0.87 to 11.2, p=0.081). There was no evidence of risk association for mutations in *POLK* (table 2).

We then performed a meta-analysis by combining the targeted sequencing, WES and chip genotyping data. Taken together, putative deleterious mutations were associated with increased risk for *PALB2* (OR 2.60, 95% CI 1.45 to 4.64; p=0.0013), *POLK* (OR 1.77, 95% CI 1.07 to 2.93; p=0.026) and *SLX4* (OR 3.37, 95% CI 1.17 to 9.70, p=0.024) and decreased risk for *FBXO10* (95% CI 0.07 to 1.00; p=0.049). After stratifying cases by histological subtype, the estimated risks were higher for HGSOC for *PALB2* (OR 3.01, 95% CI 1.59 to 5.68; p=0.00068), *POLK* (OR 1.99, 95% CI 1.15 to 3.43; p=0.014) and *SLX4* (OR 3.92, 95% CI 1.33 to 11.5; p=0.013).

We used an approximate Bayes factor to calculate the Bayes false discovery probability (BFDP) described by Wakefield<sup>38</sup> for *PALB2*, *SLX4*, *POLK* and *FBXO10* based on several different priors and assuming that the associated risk is unlikely to be greater than an OR of 4 (table 3). The evidence for association of *PALB2* was strong with a BFDP of less than 15% when the prior on the alternative hypothesis is 0.1. The nominally significant associations for the other three genes are likely to be false positives.

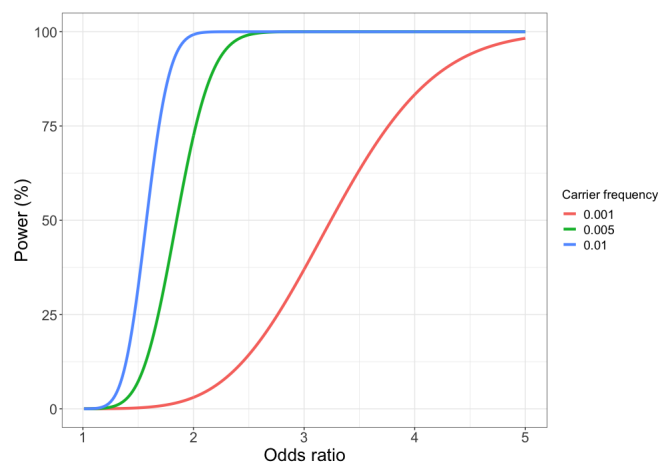
**Predicting the functional impact of missense coding variants**

Combining the whole exome and targeted sequencing data, we identified 5265 unique missense variants with minor allele frequency less than 1% in the 54 genes (online supplementary table 6). We used the in silico software programs SIFT, Polyphen-2 and Provean to evaluate the predicted impact of these variants on protein function for each gene. Of the 5265 variants, 2111 were classified as ‘deleterious’ based on at least 2 out of 3 of these classifiers. We found weak evidence for association with increased EOC risk for rare missense variants in *DUOX1* and *PAK4* using burden testing (p=0.015 and 0.025, respectively) (online supplementary table 7); for *DUOX1*, the strength of this association improved when the analyses were restricted to the HGSOC histotype (p=0.0061). When we performed the same analyses for 1493 very rare variants (MAF<0.001), we observed significant association for missense variants in *DUOX1* and *FANCE* (p=0.015 and 0.034, respectively).

**Table 3** Bayes false discovery probability for the associations reported for *PALB2*, *SLX4*, *POLK* and *FBXO10* from the meta-analysis of all the available data

Gene	Histotype	OR (95% CI)	P value	Prior probability		
				0.1	0.05	0.01
<i>PALB2</i>	HGSC	3.01 (1.59 to 5.68)	0.00068	0.14	0.26	0.65
<i>SLX4</i>	HGSC	3.92 (1.33 to 11.5)	0.013	0.65	0.80	0.95
<i>POLK</i>	HGSC	1.99 (1.15 to 3.43)	0.014	0.65	0.80	0.95
<i>FBXO10</i>	Overall	0.27 (0.07 to 1.00)	0.026	0.75	0.86	0.97



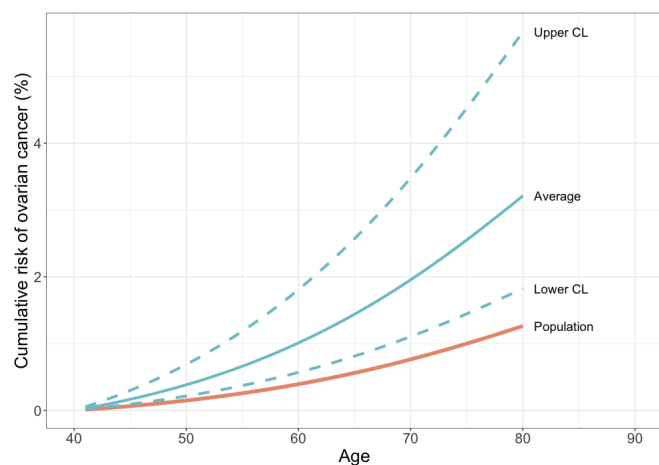


**Figure 1** Power to detect association for 5951 cases and 6385 controls at a Type I error rate of 0.0001 by deleterious variant carrier frequency and effect size (OR).

## DISCUSSION

We have evaluated the association between putative deleterious variants in 54 genes with the risk of HGSOV through a combination of whole exome and targeted sequencing analysis in 5951 cases and 6115 controls of broad European ancestries. We found evidence for four genes—*PALB2*, *POLK*, *SLX4* and *FBXO10*—associated with HGSOV risk. Association analysis in an additional 14135 ovarian cancer cases and 28655 controls genotyped through OCAC and the UK Biobank provided further support for *PALB2* as a HGSOV susceptibility gene.

The probability that a genetic association deemed statistically significant is a false positive depends on the prior of the null hypothesis and the power of the study to detect an effect size plausible under the alternative hypothesis. We calculated Wakefield's BFDP<sup>38</sup> based on several different priors to further evaluate the likelihood that *PALB2*, *POLK*, *SLX4* and *FBXO10* are EOC susceptibility genes. If we assume the prior on the alternative to be 1 in 10 or 1 in 20, the BFDPs for the association of deleterious variants in *PALB2* with HGSOV are 0.14 and 0.26, respectively. These moderately strong priors are reasonable given the evidence for the association from previously published studies.<sup>20</sup> Two studies have reported nominally significant associations for *PALB2* with



**Figure 2** Estimated cumulative risk (%) of ovarian cancer in a *PALB2* deleterious variant carrier compared with population risks for England and Wales, 2016.

OR 4.4 (95% CI 2.1 to 9.1)<sup>20</sup> and (2.87, 95% CI 1.61 to 4.74).<sup>21</sup> Kotsopoulos and colleagues reported an increased risk that was not significant (OR, 4.55, 95% CI 0.76 to 27) and, in a subset of the samples included in this study, we also found a non-significant increase in risk (OR 3.2, 95% CI 0.86 to 12).<sup>3</sup>

It is possible that cryptic population structure could cause spurious association in these data. Principal component analysis is one approach to reducing the risk of such bias, but there are too few common variants in the regions covered by the targeted sequencing panel to do a principal component analysis and chip genotyping data that would be required for such an analysis is not available for all the samples. Adjusting for country of origin and restricting the analysis to samples from individuals of broad European ancestries should reduce any problem with population stratification.

We lacked the statistical power to identify susceptibility genes conferring relative risks of less than 2 (figure 1). Our use of targeted sequencing and a definition of deleterious variants as those that likely truncate the protein product will have probably underestimated the true prevalence of deleterious variants in these genes. Incomplete coverage of each gene will have missed some small indels and single nucleotide variants. Amplicon based sequencing will also miss large deletions and rearrangements, which are relatively common in some genes.<sup>39,40</sup> Finally, any functional mutations in the non-coding region of these genes will have been missed.<sup>41</sup>

Some commercial gene-panel tests for hereditary breast-ovarian cancer already include *PALB2*. However, whether there is clinical utility in testing unaffected women for deleterious mutations in *PALB2* is not clear given the uncertainties in the risk estimates for this gene. There is no consensus over the risk threshold at which preventative surgery should be offered; many cancer genetics clinics in the UK will refer women if their predicted lifetime risk of EOC is greater than 10%. Others have suggested that the risk threshold should be lower given the low risk nature of the intervention; prophylactic surgery has been shown to be cost-effective for women at a lifetime risk of 5%. Recent updates to the US National Comprehensive Cancer Network Guidelines recommend considering risk reducing salpingo-oophorectomy in carriers of moderate risk genes if the lifetime risk of such mutation carriers exceeds 2.6%. Based on our data and population data for ovarian cancer incidence in England and Wales in 2016, the cumulative risk of ovarian cancer by age 80 for a carrier of a deleterious *PALB2* mutation is 3.2% (figure 2). Thus, a woman carrying a *PALB2* deleterious mutation would be eligible for prophylactic surgery. However, the CIs for this estimate range from 1.8% to 5.7%. Very large, well-designed case-control studies will be required to provide more precise, unbiased estimates of risk suitable for clinical counselling.

In summary, we have found relatively strong evidence that deleterious germline mutations in *PALB2* are associated with a moderate increase in the risk of HGSOV with weak evidence for *POLK*, *SLX4* and *FBXO10*. Mutations in the other 50 genes we tested are unlikely to contribute meaningfully to genetic predisposition to HGSOV. This study highlights the importance of large sample sizes needed to obtain risk estimates with the precision necessary for clinical use.

## Author affiliations

<sup>1</sup>Department of Oncology, University of Cambridge, Cambridge, UK

<sup>2</sup>School of Women's and Children's Health, Faculty of Medicine, University of New South Wales, Sydney, New South Wales, Australia

<sup>3</sup>Cancer Genetics, Queensland Institute of Medical Research—QIMR, Herston, Queensland, Australia

<sup>4</sup>Cancer Genomics and Genetics and Women's Cancer Programs, Peter MacCallum Cancer Centre, Melbourne, Victoria, Australia

<sup>5</sup>Sir Peter MacCallum Department of Oncology, The University of Melbourne, Melbourne, Victoria, Australia  
<sup>6</sup>QIMR Berghofer Department of Genetics and Computational Biology, Herston, Queensland, Australia  
<sup>7</sup>Department of Research, Peter MacCallum Cancer Centre, Melbourne, Victoria, Australia  
<sup>8</sup>Cancer Research UK Cambridge Institute, University of Cambridge, Cambridge, UK  
<sup>9</sup>Laura and Isaac Perlmutter Cancer Center, New York University, New York, New York, USA  
<sup>10</sup>Huntsman Institute, University of Utah, Salt Lake City, Utah, USA  
<sup>11</sup>Division of Public Health Sciences, Fred Hutchinson Cancer Research Center, Seattle, Washington, USA  
<sup>12</sup>Department of Epidemiology, University of Washington, Seattle, Washington, USA  
<sup>13</sup>Department of Gynaecology, Jena University Hospital, Friedrich Schiller University Jena, Jena, Thüringen, Germany  
<sup>14</sup>Gynaecology Research Unit, Hannover Medical School, Hannover, Niedersachsen, Germany  
<sup>15</sup>Department of Radiation Oncology, Hannover Medical School, Hannover, Niedersachsen, Germany  
<sup>16</sup>Department of Gynaecology, NN Alexandrov National Cancer Centre, Minsk, Minsk, Belarus  
<sup>17</sup>Womens Cancer Research Center, Magee-Womens Research Institute, Pittsburgh, Pennsylvania, USA  
<sup>18</sup>Department of Obstetrics, Gynecology and Reproductive Sciences, University of Pittsburgh School of Medicine, Pittsburgh, Pennsylvania, USA  
<sup>19</sup>Division of Cancer Prevention and Control, Roswell Park Cancer Institute, Buffalo, New York, USA  
<sup>20</sup>Department of Gynecologic Oncology, Roswell Park Cancer Institute, Buffalo, New York, USA  
<sup>21</sup>School of Public Health, University of Texas Health Science Center at Houston, Houston, Texas, USA  
<sup>22</sup>Department of Obstetrics and Gynecology, David Geffen School of Medicine, University of California Los Angeles, Los Angeles, California, USA  
<sup>23</sup>Women's Cancer Program at the Samuel Oschin Comprehensive Cancer Institute, Cedars-Sinai Medical Center, Los Angeles, California, USA  
<sup>24</sup>Department of Virus, Lifestyle and Genes, Danish Cancer Society Research Center, Copenhagen, Denmark  
<sup>25</sup>Department of Gynaecology, Rigshospitalet, University of Copenhagen, Copenhagen, Denmark  
<sup>26</sup>Department of Pathology, Herlev Hospital, University of Copenhagen, Copenhagen, Denmark  
<sup>27</sup>Department of Research, Cancer Genomics and Genetics, Peter MacCallum Cancer Centre, Melbourne, Victoria, Australia  
<sup>28</sup>Hereditary Cancer Program, Catalan Institute of Oncology, Barcelona, Catalunya, Spain  
<sup>29</sup>Translational Research Laboratory, Catalan Institute of Oncology, Barcelona, Catalunya, Spain  
<sup>30</sup>Department of Laboratory Medicine and Pathology, Mayo Clinic, Rochester, Minnesota, USA  
<sup>31</sup>Department of Health Science Research, Division of Biomedical Statistics and Informatics, Mayo Clinic, Rochester, Minnesota, USA  
<sup>32</sup>Department of Epidemiology, University of Texas MD Anderson Cancer Center, Houston, Texas, USA  
<sup>33</sup>Department of Cancer Epidemiology, Moffitt Cancer Center, Tampa, Florida, USA  
<sup>34</sup>Department of Surgery, Memorial Sloan Kettering Cancer Center, New York, New York, USA  
<sup>35</sup>Gynecologic Oncology, Laura and Isaac Perlmutter Cancer Center, New York University, New York, New York, USA  
<sup>36</sup>Department of Public Health Sciences, University of Virginia, Charlottesville, Virginia, USA  
<sup>37</sup>Department of Gynecologic Oncology, Duke University Hospital, Durham, North Carolina, USA  
<sup>38</sup>Population Health Department, QIMR Berghofer Medical Research Institute, Herston, Queensland, Australia  
<sup>39</sup>Department of Genetics and Pathology, Pomeranian Medical University in Szczecin, Szczecin, Zachodniopomorskie, Poland  
<sup>40</sup>Independent Laboratory of Molecular Biology and Genetic Diagnostics, Pomeranian Medical University in Szczecin, Szczecin, Zachodniopomorskie, Poland  
<sup>41</sup>MRC Clinical Trials Unit, Institute of Clinical Trials & Methodology, University College London, London, UK  
<sup>42</sup>Department of Epidemiology, University of Michigan School of Public Health, Ann Arbor, Michigan, USA  
<sup>43</sup>Department of Preventive Medicine, Keck School of Medicine, University of Southern California, Los Angeles, California, USA  
<sup>44</sup>Centre for Cancer Research, The Westmead Institute for Medical Research, The University of Sydney, Sydney, New South Wales, Australia  
<sup>45</sup>Department of Gynaecological Oncology, Westmead Hospital, Westmead, New South Wales, Australia

<sup>46</sup>Department of Health Science Research, Division of Epidemiology, Mayo Clinic, Rochester, Minnesota, USA  
<sup>47</sup>Kinghorn Cancer Centre, Garvan Institute of Medical Research, Darlinghurst, New South Wales, Australia  
<sup>48</sup>Center for Bioinformatics and Functional Genomics and the Cedars Sinai Genomics Core, Cedars-Sinai Medical Center, Los Angeles, California, USA  
<sup>49</sup>Department of Public Health and Primary Care, University of Cambridge, Cambridge, UK

**Acknowledgements** We thank all the study participants who contributed to this study and all the researchers, clinicians and technical and administrative staff who have made possible this work. In particular, we thank: the clinical and scientific collaborators listed at [www.aocstudy.org/](http://www.aocstudy.org/) (AOCS) and [opalstudy.qimrberghofer.edu.au](http://opalstudy.qimrberghofer.edu.au) (OPL).

**Contributors** HS planned the study, co-ordinated the sequencing, collated the results and drafted the manuscript. EDM carried out the analysis of the sequencing data. JT carried out the biostatistical analyses. GC-T, DDB, NT, JB, TG, KH, AP, EVO, JD, HRH, MAR, MD, TD, NVB, FM, KM, KO, RN, BYK, JLe, AJe, SKK, EH, CL, Miquel Angel Pujana, JC, RV, SJW, MH, CH, DL, XW, YY, JBP, DAL, JMS, AB, PMW, CC, JG, AJa, JLu, JA, CJK, IC, UM, CLP, AHW and AdF were responsible for data and sample collection for the contributing studies. MJR co-ordinated the collection of the OCAC phenotype data. Maria Intermaggio and PH carried out the targeted sequencing. IGC, EG, SR, SG and PP planned the study.

**Funding** American Cancer Society: SIOP-06-258-01-COUN. Cancer Councils of New South Wales, Victoria, Queensland, South Australia and Tasmania and Cancer Foundation of Western Australia: Multi-State Applications 191, 211 and 182. Cancer Institute NSW: 12/RIG/1-17, 15/RIG/1-16. Cancer Research UK: C490/A10119, C490/A10124, C490/A16561, Cambridge Cancer Centre. Kræftens Bekæmpelse: 94 222 52. Medical Research Council: MR\_UU\_12023. U.S. Department of Health and Human Services, National Institutes of Health, National Cancer Institute: K07CA080668, K22 CA193860, MO1RR000056, P50CA159981, R01CA112523, R01CA122443, P30CA15083, P50CA136393, R01CA76016, R01CA126841, R01CA178535, R01CA188943, R01CA61107, R01CA87538, R01CA95023. U.S. Department of Health and Human Services, National Institutes of Health, National Center for Advancing Translational Sciences: UL1TR000124. National Health and Medical Research Council of Australia 199600, 310670, 400281, 400413, 628903, APP1025142. National Institutes of Health Research: Cambridge Biomedical Research Centre, University College London Hospitals Biomedical Research Centre. The Eve Appeal: UKOPS Study. U.S. Army Medical Research and Materiel Command: DAMD17-01-1-0729, DAMD17-02-1-0669, DAMD17-02-1-06. U.S. Department of Defense Ovarian Cancer Research Program: W81XWH-07-0449. The University of Cambridge has received salary support in respect of PDPP from the NHS in the East of England through the Clinical Academic Reserve. SG is a recipient of the Barth Family Chair in Cancer Genetics.

**Competing interests** None declared.

**Patient consent for publication** Not required.

**Provenance and peer review** Not commissioned; externally peer reviewed.

**Data availability statement** Data are available from the authors on request.

**Open access** This is an open access article distributed in accordance with the Creative Commons Attribution 4.0 Unported (CC BY 4.0) license, which permits others to copy, redistribute, remix, transform and build upon this work for any purpose, provided the original work is properly cited, a link to the licence is given, and indication of whether changes were made. See: <https://creativecommons.org/licenses/by/4.0/>.

#### ORCID iDs

Chad Huff <http://orcid.org/0000-0002-2386-1503>  
 Cezary Cybulski <http://orcid.org/0000-0002-2819-3057>  
 Paul Pharos <http://orcid.org/0000-0001-8494-732X>

#### REFERENCES

- Miki Y, Swensen J, Shattuck-Eidens D, Futreal PA, Harshman K, Tavtigian S, Liu Q, Cochran C, Bennett LM, Ding W, Bell R, Rosenthal J, Hussey C, Tran T, McClure M, Frye C, Hattier T, Phelps R, Haugenstrano A, Katcher H, Yakumo K, Gholami Z, Shaffer D, Stone S, Bayer S, Wray C, Bogden R, Dayananth P, Ward J, Tonin P, Narod S, Bristow PK, Norris FH, Helvering L, Morrison P, Rostock P, Lai M, Barrett JC, Lewis C, Neuhausen S, Cannon-Albright L, Goldgar D, Wiseman R, Kamb A, Skolnick MH. A strong candidate for the breast and ovarian cancer susceptibility gene BRCA1. *Science* 1994;266:66–71.
- Wooster R, Bignell G, Lancaster J, Swift S, Seal S, Mangion J, Collins N, Gregory S, Gumbs C, Micklem G. Identification of the breast cancer susceptibility gene BRCA2. *Nature* 1995;378:789–92.
- Ramus SJ, Song H, Dicks E, Tyrer JP, Rosenthal AN, Intermaggio MP, Fraser L, Gentry-Maharaj A, Hayward J, Philpott S, Anderson C, Edlund CK, Conti D, Harrington P,

- Barrowdale D, Bowtell DD, Alsop K, Mitchell G, Cicek MS, Cunningham JM, Fridley BL, Alsop J, Jimenez-Linan M, Poblete S, Lele S, Sucheston-Campbell L, Moysich KB, Sieh W, McGuire V, Lester J, Bogdanova N, Dürst M, Hillemanns P, Odunsi K, Whittemore AS, Karlan BY, Dörk T, Goode EL, Menon U, Jacobs IJ, Antoniou AC, Pharoah PDP, Gayther SA, AOCs Study Group, Ovarian Cancer Association Consortium. Germline mutations in the BRIP1, BARD1, PALB2, and NBN genes in women with ovarian cancer. *J Natl Cancer Inst* 2015;107. doi:10.1093/jnci/djv214. [Epub ahead of print: 27 Aug 2015].
- 4 Song H, Dicks E, Ramus SJ, Tyrer JP, Intermaggio MP, Hayward J, Edlund CK, Conti D, Harrington P, Fraser L, Philpott S, Anderson C, Rosenthal A, Gentry-Maharaj A, Bowtell DD, Alsop K, Cicek MS, Cunningham JM, Fridley BL, Alsop J, Jimenez-Linan M, Høgdall E, Høgdall CK, Jensen A, Kjaer SK, Lubiński J, Huzarski T, Jakubowska A, Gronwald J, Poblete S, Lele S, Sucheston-Campbell L, Moysich KB, Odunsi K, Goode EL, Menon U, Jacobs IJ, Gayther SA, Pharoah PDP. Contribution of germline mutations in the RAD51B, RAD51C, and RAD51D genes to ovarian cancer in the population. *J Clin Oncol* 2015;33:2901–7.
- 5 Pal T, Permuth-Wey J, Sellers TA. A review of the clinical relevance of mismatch-repair deficiency in ovarian cancer. *Cancer* 2008;113:733–42.
- 6 Song H, Cicek MS, Dicks E, Harrington P, Ramus SJ, Cunningham JM, Fridley BL, Tyrer JP, Alsop J, Jimenez-Linan M, Gayther SA, Goode EL, Pharoah PDP. The contribution of deleterious germline mutations in BRCA1, BRCA2 and the mismatch repair genes to ovarian cancer in the population. *Hum Mol Genet* 2014;23:4703–9.
- 7 Zhu Q, Zhang J, Chen Y, Hu Q, Shen H, Huang R-Y, Liu Q, Kaur J, Long M, Battaglia S, Eng KH, Lele SB, Zsiros E, Villella J, Lugade A, Yao S, Liu S, Moysich K, Odunsi KO. Whole-Exome sequencing of ovarian cancer families uncovers putative predisposition genes. *Int J Cancer* 2020;146:2147–55.
- 8 Dicks E, Song H, Ramus SJ, Oudenhove EV, Tyrer JP, Intermaggio MP, Kar S, Harrington P, Bowtell DD, Group AS, Cicek MS, Cunningham JM, Fridley BL, Alsop J, Jimenez-Linan M, Piskorz A, Goranova T, Kent E, Siddiqui N, Paul J, Crawford R, Poblete S, Lele S, Sucheston-Campbell L, Moysich KB, Sieh W, McGuire V, Lester J, Odunsi K, Whittemore AS, Bogdanova N, Dürst M, Hillemanns P, Karlan BY, Gentry-Maharaj A, Menon U, Tischkowitz M, Levine D, Brenton JD, Dörk T, Goode EL, Gayther SA, Pharoah DPP. Germline whole exome sequencing and large-scale replication identifies *FANCM* as a likely high grade serous ovarian cancer susceptibility gene. *Oncotarget* 2017;8:50930–40.
- 9 Yang X, Leslie G, Doroszuk A, Schneider S, Allen J, Decker B, Dunning AM, Redman J, Scarth J, Plaskocinska I, Luccarini C, Shah M, Pooley K, Dorling L, Lee A, Adank MA, Adlard J, Aittomäki K, Andrulis IL, Ang P, Barwell J, Bernstein JL, Bobolis K, Borg A, Blomqvist C, Claes KBM, Concannon P, Cuggia A, Culver JO, Damiola F, de Pauw A, Diez O, Dolinsky JS, Domchek SM, Engel C, Evans DG, Fostira F, Garber J, Golmard L, Goode EL, Gruber SB, Hahnen E, Hake C, Heikinen T, Hurley JE, Janavicius R, Kleibl Z, Kleiblova P, Konstantopoulou I, Kvist A, Laduca H, ASG L, Lesueur F, Maher ER, Mannerman A, Manoukian S, McFarland R, McKinnon W, Meindl A, Metcalfe K, Mohd Taib NA, Moilanen J, Nathanson KL, Neuhausen S, PS N, Nguyen-Dumont T, Nielsen SM, Obermair F, Offit K, Olopade OI, Ottini L, Penkert J, Pylkas K, Radice P, Ramus SJ, Rudaitis V, Side L, Silva-Smith R, Silvestri V, Skytte AB, Slavin T, Soukupova J, Tondini C, Trainer AH, Unzeitig G, Usha L, van Overeem Hansen T, Whitworth J, Wood M, Yip CH, Yoon SY, Yussuf A, Zogopoulos G, Goldgar D, Hopper JL, Chenevix-Trench G, Pharoah P, George SHL, Balmana J, Houdayer C, James P, El-Haffaf Z, Ehrencrona H, Janatova M, Peterlongo P, Nevanlinna H, Schmutzler R, Teo SH, Robson M, Pal T, Couch F, Weitzel JN, Elliott A, Southey M, Winqvist R, Easton DF, Foulkes WD, Antoniou AC, Tischkowitz M. Cancer risks associated with germline PALB2 pathogenic variants: an international study of 524 families. *J Clin Oncol* 2019;JCO1901907.
- 10 Bojesen SE, Pooley KA, Johnatty SE, Beesley J, Michailidou K, Tyrer JP, Edwards SL, Pickett HA, Shen HC, Smart CE, Hillman KM, Mai PL, Lawrenson K, Stutz MD, Lu Y, Karevan R, Woods N, Johnston RL, French JD, Chen X, Weischer M, Nielsen SF, Maranian MJ, Ghousaini M, Ahmed S, Baynes C, Bolla MK, Wang Q, Dennis J, McGuffog L, Barrowdale D, Lee A, Healey S, Lush M, Tessier DC, Vincent D, Bacot F, Vergote I, Lambrechts S, Despiere E, Risch HA, González-Neira A, Rossing MA, Pita G, Doherty JA, Alvarez N, Larson MC, Fridley BL, Schoof N, Chang-Claude J, Cicek MS, Peto J, Kalli KR, Broeks A, Armasu SM, Schmidt MK, Braaf LM, Winterhoff B, Nevanlinna H, Konecny GE, Lambrechts D, Rogmann L, Guénel P, Teoman A, Milne RL, Garcia JJ, Cox A, Shridhar V, Burwinkel B, Marme F, Hein R, Sawyer EJ, Haiman CA, Wang-Gohrke S, Andrulis IL, Moysich KB, Hopper JL, Odunsi K, Lindblom A, Giles GG, Brenner H, Simard J, Lurie G, Fasching PA, Carney ME, Radice P, Wilkens LR, Swerdlow A, Goodman MT, Brauch H, Garcia-Closas M, Hillemanns P, Winqvist R, Dürst M, Devilee P, Runnebaum I, Jakubowska A, Lubinski J, Mannerman A, Butzow R, Bogdanova NV, Dörk T, Pelttari LM, Zheng W, Leminen A, Anton-Culver H, Bunker CH, Kristensen V, Ness RB, Muir K, Edwards R, Meindl A, Heitz F, Matsuo K, du Bois A, Wu AH, Harter P, Teo S-H, Schwaab I, Shu X-O, Blot W, Hosono S, Kang D, Nakanishi T, Hartman M, Yatabe Y, Hamann U, Karlan BY, Sangrajrang S, Kjaer SK, Gaborieau V, Jensen A, Eccles D, Høgdall E, Shen C-Y, Brown J, Woo YL, Shah M, Azmi MAN, Luben R, Omar SZ, Czene K, Vierkant RA, Nordestgaard BG, Flyger H, Vachon C, Olson JE, Wang X, Levine DA, Rudolph A, Weber RP, Flesch-Janys D, Iversen E, Nickels S, Schildkraut JM, Silva IDS, Cramer DW, Gibson L, Terry KL, Fletcher O, Vitis AF, van der Schoot CE, Poole EM, Hogervorst FBL, Tworoger SS, Liu J, Bandera EV, Li J, Olson SH, Humphreys K, Orlow I, Blomqvist C, Rodriguez-Rodriguez L, Aittomäki K, Salvesen HB, Muranen TA, Wik E, Brouwers B, Krakstad C, Wauters E, Halle MK, Wildiers H, Kiemeny LA, Mulot C, Aben KK, Laurent-Puig P, Altena AM, Truong T, Massuger L FAG, Benitez J, Pejovic T, Perez JIA, Hoatlin M, Zamora MP, Cook LS, Balasubramanian SP, Kelemen LE, Schneeweiss A, Le ND, Sohn C, Brooks-Wilson A, Tomlinson I, Kerin MJ, Miller N, Cybulski C, Henderson BE, Menkiszak J, Schumacher F, Wentzensen N, Le Marchand L, Yang HP, Mulligan AM, Glendon G, Engelholm SA, Knight JA, Høgdall CK, Apicella C, Gore M, Tsimiklis H, Song H, Southey MC, Jager A, den Ouweland AMW, Brown R, Martens JWM, Flanagan JM, Kriege M, Paul J, Margolin S, Siddiqui N, Severi G, Whittemore AS, Baglietto L, McGuire V, Stegmaier C, Sieh W, Müller H, Arndt V, Labréche F, Gao Y-T, Goldberg MS, Yang G, Dumont M, McLaughlin JR, Hartmann A, Ekici AB, Beckmann MW, Phelan CM, Lux MP, Permuth-Wey J, Peissel B, Sellers TA, Ficarazzi F, Barile M, Ziogas A, Ashworth A, Gentry-Maharaj A, Jones M, Ramus SJ, Orr N, Menon U, Pearce CL, Brüning T, Pike MC, Ko Y-D, Lissowska J, Figueroa J, Kupryjanczyk J, Chanock SJ, Dansonka-Mieszowska A, Jukkola-Vuorinen A, Rzepecka IK, Pylkäs K, Bidzinski M, Kauppila S, Hollestelle A, Seynaeve C, Tollenaar RAEM, Durda K, Jaworska K, Hartikainen JM, Kosma V-M, Kataja V, Anttonenkovana NN, Long J, Shrubsole M, Deming-Halverson S, Lophatananon A, Siriwanarangsang P, Stewart-Brown S, Ditsch N, Lichtner P, Schmutzler RK, Ito H, Iwata H, Tajima K, Tseng C-C, Stram DO, van den Berg D, Yip CH, Ikram MK, Teh Y-C, Cai H, Lu W, Signorello LB, Cai Q, Noh D-Y, Yoo K-Y, Miao H, Iau PT-C, Teo YY, McKay J, Shapiro C, Ademuyiwa F, Fountzilas G, Hsiung C-N, Yu J-C, Hou M-F, Healey CS, Luccarini C, Peock S, Stoppa-Lyonnet D, Peterlongo P, Rebbeck TR, Piedmonte M, Singer CF, Friedman E, Thomassen M, Offit K, Hansen TVO, Neuhausen SL, Szabo CI, Blanco I, Garber J, Narod SA, Weitzel JN, Montagna M, Olah E, Godwin AK, Yannoukakos D, Goldgar DE, Caldes T, Imyanitov EN, Tihomirova L, Arun BK, Campbell I, Mensenkamp AR, van Asperen CJ, van Roozendaal KEP, Meijers-Heijboer H, Collé JM, Oosterwijk JC, Hooning MJ, Rookus MA, van der Luijt RB, Os TAM, Evans DG, Frost D, Finberg E, Barwell J, Walker L, Kennedy MJ, Platte R, Davidson R, Ellis SD, Cole T, Bressac-de Paillerets B, Buecher B, Damiola F, Faivre L, Frenay M, Sinilnikova OM, Caron O, Giraud S, Mazoyer S, Bonadona V, Caux-Moncoulier V, Toloczko-Grabarek A, Gronwald J, Byrski T, Spurdle AB, Bonanni B, Zaffaroni D, Giannini G, Bernard L, Dolcetti R, Manoukian S, Arnold N, Engel C, Deissler H, Rhiem K, Niederacher D, Plendl H, Sutter C, Wappenschmidt B, Borg A, Melin B, Rantalä J, Soller M, Nathanson KL, Domchek SM, Rodriguez GC, Salani R, Kaulich DG, Tea M-K, Paluch SS, Laitman Y, Skytte A-B, Kruse TA, Jensen UB, Robson M, Gerdes A-M, Ejlersen B, Foretova L, Savage SA, Lester J, Soucy P, Kuchenbaecker KB, Olswold C, Cunningham JM, Slager S, Pankratz VS, Dicks E, Lakhani SR, Couch FJ, Hall P, Monteiro ANA, Gayther SA, Pharoah PDP, Reddel RR, Goode EL, Greene MH, Easton DF, Berchuck A, Antoniou AC, Chenevix-Trench G, Dunning AM, Australian Cancer Study, Australian Ovarian Cancer Study, Kathleen Cunningham Foundation Consortium for Research into Familial Breast Cancer (kConFab), Gene Environment Interaction and Breast Cancer (GENICA), Swedish Breast Cancer Study (SWE-BRCA), Hereditary Breast and Ovarian Cancer Research Group Netherlands (HEBON), Epidemiological study of BRCA1 & BRCA2 Mutation Carriers (EMBRACE), Genetic Modifiers of Cancer Risk in BRCA1/2 Mutation Carriers (GEMO). Multiple independent variants at the TERT locus are associated with telomere length and risks of breast and ovarian cancer. *Nat Genet* 2013;45:371–84.
- 11 Bolton KL, Tyrer J, Song H, Ramus SJ, Notaridou M, Jones C, Sher T, Gentry-Maharaj A, Wozniak E, Tsai Y-Y, Weidhaas J, Paik D, Van Den Berg DJ, Stram DO, Pearce CL, Wu AH, Brewster W, Anton-Culver H, Ziogas A, Narod SA, Levine DA, Kaye SB, Brown R, Paul J, Flanagan J, Sieh W, McGuire V, Whittemore AS, Campbell I, Gore ME, Lissowska J, Yang HP, Medrek K, Gronwald J, Lubinski J, Jakubowska A, Le ND, Cook LS, Kelemen LE, Brooks-Wilson A, Brook-Wilson A, Massuger L FAG, Kiemeny LA, Aben KKH, van Altena AM, Houlston R, Tomlinson I, Palmieri RT, Moorman PG, Schildkraut J, Iversen ES, Phelan C, Vierkant RA, Cunningham JM, Goode EL, Fridley BL, Kruger-Kjaer S, Blaeker J, Hogdall E, Hogdall C, Gross J, Jarlan V, Ness RB, Edwards RP, Odunsi K, Moysich KB, Baker JA, Modugno F, Heikkinen T, Butzow R, Nevanlinna H, Leminen A, Bogdanova N, Anttonenkovana N, Doerk T, Hillemanns P, Dürst M, Runnebaum I, Thompson PJ, Carney ME, Goodman MT, Lurie G, Wang-Gohrke S, Hein R, Chang-Claude J, Rossing MA, Cushing-Haugen KL, Doherty J, Chen C, Rafnar T, Besenbacher S, Sulem P, Stefansson K, Birrer MJ, Terry KL, Hernandez D, Cramer DW, Vergote I, Amant F, Lambrechts D, Despiere E, Fasching PA, Beckmann MW, Thiel FC, Ekici AB, Chen X, Johnatty SE, Webb PM, Beesley J, Chanock S, Garcia-Closas M, Sellers T, Easton DF, Berchuck A, Chenevix-Trench G, Pharoah PDP, Gayther SA, Australian Ovarian Cancer Study Group, Australian Cancer Study (Ovarian Cancer), Ovarian Cancer Association Consortium. Common variants at 19p13 are associated with susceptibility to ovarian cancer. *Nat Genet* 2010;42:880–4.
- 12 Goode EL, Chenevix-Trench G, Song H, Ramus SJ, Notaridou M, Lawrenson K, Widschwendter M, Vierkant RA, Larson MC, Kjaer SK, Birrer MJ, Berchuck A, Schildkraut J, Tomlinson I, Kiemeny LA, Cook LS, Gronwald J, Garcia-Closas M, Gore ME, Campbell I, Whittemore AS, Sutphen R, Phelan C, Anton-Culver H, Pearce CL, Lambrechts D, Rossing MA, Chang-Claude J, Moysich KB, Goodman MT, Dörk T, Nevanlinna H, Ness RB, Rafnar T, Hogdall C, Hogdall E, Fridley BL, Cunningham JM, Sieh W, McGuire V, Godwin AK, Cramer DW, Hernandez D, Levine D, Lu K, Iversen ES, Palmieri RT, Houlston R, van Altena AM, Aben KKH, Massuger L FAG, Brooks-Wilson A, Kelemen LE, Le ND, Jakubowska A, Lubinski J, Medrek K, Stafford A, Easton DF, Tyrer J, Bolton KL, Harrington P, Eccles D, Chen A, Molina AN, Davila BN, Arango H, Tsai Y-Y, Chen Z, Risch HA, McLaughlin J, Narod SA, Ziogas A, Brewster W, Gentry-Maharaj A, Menon U, Wu AH, Stram DO, Pike MC, Beesley J, Webb PM, Chen X, Ekici AB, Thiel FC, Beckmann MW, Yang H, Wentzensen N, Lissowska J, Fasching PA, Despiere E,



- Amant F, Vergote I, Doherty J, Hein R, Wang-Gohrke S, Lurie G, Carney ME, Thompson PJ, Runnebaum I, Hillemanns P, Dürst M, Antonenkova N, Bogdanova N, Leminen A, Butzow R, Heikkinen T, Stefansson K, Sulem P, Besenbacher S, Sellers TA, Gayther SA, Pharoah PDP, Wellcome Trust Case-Control Consortium, Australian Cancer Study (Ovarian Cancer), Australian Ovarian Cancer Study Group, Ovarian Cancer Association Consortium (OCAC). A genome-wide association study identifies susceptibility loci for ovarian cancer at 2q31 and 8q24. *Nat Genet* 2010;42:874–9.
- 13 Permut-Wey J, Lawrenson K, Shen HC, Velkova A, Tyrer JP, Chen Z, Lin H-Y, Chen YA, Tsai Y-Y, Qu X, Ramus SJ, Karevan R, Lee J, Lee N, Larson MC, Aben KK, Anton-Culver H, Antonenkova N, Antoniou AC, Armasu SM, Bacot F, Baglietto L, Bandera EV, Barnholtz-Sloan J, Beckmann MW, Birrer MJ, Bloom G, Bogdanova N, Brinton LA, Brooks-Wilson A, Brown R, Butzow R, Cai Q, Campbell I, Chang-Claude J, Chanock S, Chenevix-Trench G, Cheng JQ, Cicek MS, Coetzee GA, Cook LS, Couch FJ, Cramer DW, Cunningham JM, Dansonka-Mieszkowska A, Despierre E, Doherty JA, Dörk T, du Bois A, Dürst M, Easton DF, Eccles D, Edwards R, Ekici AB, Fasching PA, Fenstermacher DA, Flanagan JM, Garcia-Closas M, Gentry-Maharaj A, Giles GG, Glasspool RM, Gonzalez-Bosquet J, Goodman MT, Gore M, Górski B, Gronwald J, Hall P, Halle MK, Harter P, Heitz F, Hillemanns P, Hoatlin M, Høgdall CK, Høgdall E, Hosono S, Jakubowska A, Jensen A, Jim H, Kalli KR, Karlan BY, Kaye SB, Kelemen LE, Kiemeny LA, Kikkawa F, Konecny GE, Krakstad C, Kjaer SK, Kupryjanczyk J, Lambrechts D, Lambrechts S, Lancaster JM, Le ND, Leminen A, Levine DA, Liang D, Lim BK, Lin J, Lissowska J, Lu KH, Lubiński J, Lurie G, Massuger LFAG, Matsuo K, McGuire V, McLaughlin JR, Menon U, Modugno F, Moysich KB, Nakanishi T, Narod SA, Nedergaard L, Ness RB, Nevanlinna H, Nickels S, Noushmehr H, Odunsi K, Olson SH, Orlov I, Paul J, Pearce CL, Pejovic T, Pelttari LM, Pike MC, Poole EM, Raska P, Renner SP, Risch HA, Rodriguez-Rodriguez L, Rossing MA, Rudolph A, Runnebaum IB, Rzepecka IK, Salvesen HB, Schwaab I, Severi G, Shridhar V, Shu X-O, Shvetsov YB, Sieh W, Song H, Southey MC, Spiwakiewicz B, Stram D, Sutphen R, Teo S-H, Terry KL, Tessier DC, Thompson PJ, Tworoger SS, van Altena AM, Vergote I, Vierkant RA, Vincent D, Vitonis AF, Wang-Gohrke S, Palmieri Weber R, Wentzensen N, Whittemore AS, Wik E, Wilkens LR, Winterhoff B, Woo YL, Wu AH, Xiang Y-B, Yang HP, Zheng W, Ziogas A, Zulkifli F, Phelan CM, Iversen E, Schildkraut JM, Berchuck A, Fridley BL, Goode EL, Pharoah PDP, Monteiro ANA, Sellers TA, Gayther SA, Australian Cancer S, Australian Cancer Study, Australian Ovarian Cancer Study, Consortium of Investigators of Modifiers of BRCA1/2. Identification and molecular characterization of a new ovarian cancer susceptibility locus at 17q21.31. *Nat Commun* 2013;4:1627.
- 14 Pharoah PDP, Tsai Y-Y, Ramus SJ, Phelan CM, Goode EL, Lawrenson K, Buckley M, Fridley BL, Tyrer JP, Shen H, Weber R, Karevan R, Larson MC, Song H, Tessier DC, Bacot F, Vincent D, Cunningham JM, Dennis J, Dicks E, Aben KK, Anton-Culver H, Antonenkova N, Armasu SM, Baglietto L, Bandera EV, Beckmann MW, Birrer MJ, Bloom G, Bogdanova N, Brenton JD, Brinton LA, Brooks-Wilson A, Brown R, Butzow R, Campbell I, Carney ME, Carvalho RS, Chang-Claude J, Chen YA, Chen Z, Chow W-H, Cicek MS, Coetzee G, Cook LS, Cramer DW, Cybulski C, Dansonka-Mieszkowska A, Despierre E, Doherty JA, Dörk T, du Bois A, Dürst M, Eccles D, Edwards R, Ekici AB, Fasching PA, Fenstermacher D, Flanagan J, Guo Y-T, Garcia-Closas M, Gentry-Maharaj A, Giles G, Gjyshi A, Gore M, Gronwald J, Guo Q, Halle MK, Harter P, Hein A, Heitz F, Hillemanns P, Hoatlin M, Høgdall E, Høgdall CK, Hosono S, Jakubowska A, Jensen A, Kalli KR, Karlan BY, Kelemen LE, Kiemeny LA, Kjaer SK, Konecny GE, Krakstad C, Kupryjanczyk J, Lambrechts D, Lambrechts S, Le ND, Lee N, Lee J, Leminen A, Lim BK, Lissowska J, Lubiński J, Lundvall L, Lurie G, Massuger LFAG, Matsuo K, McGuire V, McLaughlin JR, Menon U, Modugno F, Moysich KB, Nakanishi T, Narod SA, Ness RB, Nevanlinna H, Nickels S, Noushmehr H, Odunsi K, Olson S, Orlov I, Paul J, Pejovic T, Pelttari LM, Permut-Wey J, Pike MC, Poole EM, Qu X, Risch HA, Rodriguez-Rodriguez L, Rossing MA, Rudolph A, Runnebaum I, Rzepecka IK, Salvesen HB, Schwaab I, Severi G, Shen H, Shridhar V, Shu X-O, Sieh W, Southey MC, Spellman P, Tajima K, Teo S-H, Terry KL, Thompson PJ, Timorek A, Tworoger SS, van Altena AM, van den Berg D, Vergote I, Vierkant RA, Vitonis AF, Wang-Gohrke S, Wentzensen N, Whittemore AS, Wik E, Winterhoff B, Woo YL, Wu AH, Yang HP, Zheng W, Ziogas A, Zulkifli F, Goodman MT, Hall P, Easton DF, Pearce CL, Berchuck A, Chenevix-Trench G, Iversen E, Monteiro ANA, Gayther SA, Schildkraut JM, Sellers TA, Australian Cancer S, Australian Cancer Study, Australian Ovarian Cancer Study Group. Gwas meta-analysis and replication identifies three new susceptibility loci for ovarian cancer. *Nat Genet* 2013;45:362–70.
- 15 Phelan CM, Kuchenbaecker KB, Tyrer JP, Kar SP, Lawrenson K, Winham SJ, Dennis J, Pirie A, Riggan MJ, Chornokur G, Earp MA, Lyra PC, Lee JM, Coetzee S, Beesley J, McGuffog L, Soucy P, Dicks E, Lee A, Barrowdale D, Lecarpentier J, Leslie G, Aalfs CM, Aben KKH, Adams M, Adlard J, Andrlus I, Anton-Culver H, Antonenkova N, Aravantinos G, Arnold N, Arun BK, Arver B, Azzollini J, Balmaña J, Banerjee SN, Barjhoux L, Barkardottir RB, Bean Y, Beckmann MW, Beeghly-Fadiel A, Benitez J, Bermisheva M, Bernardini MQ, Birrer MJ, Bjorge L, Black A, Blankstein K, Blok MJ, Bodelon C, Bogdanova N, Bojesen A, Bonanni B, Borg Åke, Bradbury AR, Brenton JD, Brewer C, Brinton L, Broberg P, Brooks-Wilson A, Bruinsma F, Brunet J, Buecher B, Butzow R, Buys SS, Caldes T, Caligo MA, Campbell I, Cannioto R, Carney ME, Cescon T, Chan SB, Chang-Claude J, Chanock S, Chen XQ, Chiew Y-E, Chiquette J, Chung WK, Claes KBM, Conner T, Cook LS, Cook J, Cramer DW, Cunningham JM, D'Aloisio AA, Daly MB, Damiola F, Damirovna SD, Dansonka-Mieszkowska A, Dao F, Davidson R, DeFazio A, Delnatte C, Doheny KF, Diez O, Ding YC, Doherty JA, Domchek SM, Dorfling CM, Dörk T, Dossus L, Duran M, Dürst M, Dworniczak B, Eccles D, Edwards T, Eeles R, Eilber U, Ejlertsen B, Ekici AB, Ellis S, Elvira M, Eng KH, Engel C, Evans DG, Fasching PA, Ferguson S, Ferrer SF, Flanagan JM, Fogarty ZC, Fortner RT, Fostira F, Foulkes WD, Fountzilas G, Fridley BL, Friebel TM, Friedman E, Frost D, Ganz PA, Garber J, Garcia MJ, Garcia-Barberan V, Gehrig A, Gentry-Maharaj A, Gerdes A-M, Giles GG, Glasspool R, Glendon G, Godwin AK, Goldgar DE, Goranova T, Gore M, Greene MH, Gronwald J, Gruber S, Hahnen E, Haiman CA, Håkansson N, Hamann U, Hansen TVO, Harrington PA, Harris HR, Hauke J, Hein A, Henderson A, Hildebrandt MAT, Hillemanns P, Hodgson S, Høgdall CK, Høgdall E, Hogervorst FBL, Holland H, Hoening MJ, Hosking K, Huang R-Y, Hulick PJ, Hung J, Hunter DJ, Huntsman DG, Huzarski T, Imyanitov EN, Isaacs C, Iversen ES, Izatt L, Izquierdo A, Jakubowska A, James P, Janavicius R, Jernetz M, Jensen A, Jensen UB, John EM, Johnatty S, Jones ME, Kannisto P, Karlan BY, Karnezis A, Kast K, Kennedy CJ, Khusnutdinova E, Kiemeny LA, Kiiski JI, Kim S-W, Kjaer SK, Köbel M, Kopperud RK, Kruse TA, Kupryjanczyk J, Kwong A, Laitman Y, Lambrechts D, Larrañaga N, Larson MC, Lazaro C, Le ND, Le Marchand L, Lee JW, Lele SB, Leminen A, Leroux D, Lester J, Lesueur F, Levine DA, Liang D, Liebrich C, Lilyquist J, Lipworth L, Lissowska J, Lu KH, Lubinski J, Luccarini C, Lundvall L, Mai PL, Mendoza-Fandiño G, Manoukian S, Massuger LFAG, May T, Mazoyer S, McAlpine JN, McGuire V, McLaughlin JR, McNeish I, Meijers-Heijboer H, Meindl A, Menon U, Mensenkamp AR, Merritt MA, Milne RL, Mitchell G, Modugno F, Moes-Sosnowska J, Moffitt M, Montagna M, Moysich KB, Mulligan AM, Musini G, Nattar L, Nedergaard L, Ness RB, Neuhausen SL, Nevanlinna H, Niederacher D, Nussbaum RL, Odunsi K, Olah E, Olopade OI, Olson H, Olswold C, O'Malley DM, Ong K-R, Onland-Moret NC, Orr N, Orsulic S, Osorio A, Palli D, Papi L, Park-Simon T-W, Paul J, Pearce CL, Pedersen IS, Peeters PHM, Peissel B, Peixoto A, Pejovic T, Pelttari LM, Permut-Wey J, Peterlongo P, Pezzani L, Pfeiler G, Phillips K-A, Piedmonte M, Pike MC, Piskorz AM, Poblete SR, Poczta T, Poole EM, Poppe B, Porteous ME, Prieur F, Prokofyeva D, Pugh E, Pujana MA, Pujol P, Radice P, Rantala J, Rappaport-Fuerhauser C, Rennert G, Rhiem K, Rice P, Richardson A, Robson M, Rodriguez GC, Rodriguez-Antona C, Romm J, Rookus MA, Rossing MA, Rothstein JH, Rudolph A, Runnebaum IB, Salvesen HB, Sandler DP, Schoemaker MJ, Senter L, Setiawan VW, Severi G, Sharma P, Shelford T, Siddiqui N, Side LE, Sieh W, Singer CF, Sobol H, Song H, Southey MC, Spurdle AB, Stadler Z, Steinemann D, Stoppa-Lyonnet D, Sucheston-Campbell LE, Sukiennicki G, Sutphen R, Sutter C, Swerdlow AJ, Szabo CI, Szafron L, Tan YY, Taylor JA, Tea M-K, Teixeira MR, Teo S-H, Terry KL, Thompson PJ, Thomsen LCV, Thull DL, Tihomirova L, Tinker AV, Tischkowitz M, Tognazzo S, Toland AE, Tone A, Trabert B, Travis RC, Trichopoulos A, Tung N, Tworoger SS, van Altena AM, Van Den Berg D, van der Hout AH, van der Luijt RB, Van Heetvelde M, Van Nieuwenhuysen E, van Rensburg EJ, Vanderstichele A, Varon-Mateeva R, Vega A, Edwards DV, Vergote I, Vierkant RA, Vijai J, Vratimos A, Walker L, Walsh C, Wand D, Wang-Gohrke S, Wappenschmidt B, Webb PM, Weinberg CR, Weitzel JN, Wentzensen N, Whittemore AS, Wijnen JT, Wilkens LR, Wolk A, Woo M, Wu X, Wu AH, Yang H, Yannoukakis D, Ziogas A, Zorn KK, Narod SA, Easton DF, Amos CI, Schildkraut JM, Ramus SJ, Ottini L, Goodman MT, Park SK, Kelemen LE, Risch HA, Thomassen M, Offit K, Simard J, Schmutzler RK, Hazelett D, Monteiro AN, Couch FJ, Berchuck A, Chenevix-Trench G, Goode EL, Sellers TA, Gayther SA, Antoniou AC, Pharoah PDP, AOCs study group, EMBRACE Study, GEMO Study Collaborators, HEBON Study, KConFab Investigators, OPAL study group. Identification of 12 new susceptibility loci for different histotypes of epithelial ovarian cancer. *Nat Genet* 2017;49:680–91.
- 16 Shen H, Fridley BL, Song H, Lawrenson K, Cunningham JM, Ramus SJ, Cicek MS, Tyrer J, Stram D, Larson MC, Kobel M, Consortium P, Ziogas A, Zheng W, Yang HP, AH W, Wozniak EL, Woo YL, Winterhoff B, Wik E, Whittemore AS, Wentzensen N, Weber RP, Vitonis AF, Vincent D, Vierkant RA, Vergote I, Van Den Berg D, Van Altena AM, Tworoger SS, Thompson PJ, Tessier DC, Terry KL, Teo SH, Templeman C, Stram DO, Southey MC, Sieh W, Siddiqui N, Shvetsov YB, Shu XO, Shridhar V, Wang-Gohrke S, Severi G, Schwaab I, Salvesen HB, Rzepecka IK, Runnebaum IB, Rossing MA, Rodriguez-Rodriguez L, Risch HA, Renner SP, Poole EM, Pike MC, Phelan CM, Pelttari LM, Pejovic T, Paul J, Orlov I, Omar SZ, Olson SH, Odunsi K, Nickels S, Nevanlinna H, Ness RB, Narod SA, Nakanishi T, Moysich KB, Monteiro AN, Moes-Sosnowska J, Modugno F, Menon U, McLaughlin JR, McGuire V, Matsuo K, Adenan NA, Massuger LF, Lurie G, Lundvall L, Lubinski J, Lissowska J, Levine DA, Leminen A, Lee AW, ND L, Lambrechts S, Lambrechts D, Kupryjanczyk J, Krakstad C, Konecny GE, Kjaer SK, Kiemeny LA, Kelemen LE, Keeney GL, Karlan BY, Karevan R, Kalli KR, Kajiyama H, BT J, Jensen A, Jakubowska A, Iversen E, Hosono S, Høgdall CK, Høgdall E, Hoatlin M, Hillemanns P, Heitz F, Hein R, Harter P, Halle MK, Hall P, Gronwald J, Gore M, Goodman MT, Giles GG, Gentry-Maharaj A, Garcia-Closas M, Flanagan JM, Fasching PA, Ekici AB, Edwards R, Eccles D, Easton DF, Dürst M, du Bois A, Dörk T, Doherty JA, Despierre E, Dansonka-Mieszkowska A, Cybulski C, Cramer DW, Cook LS, Chen X, Charbonneau B, Chang-Claude J, Campbell I, Butzow R, Bunker CH, Bruggemann D, Brown R, Brooks-Wilson A, Brinton LA, Bogdanova N, Block MS, Benjamin E, Beesley J, Beckmann MW, Bandera EV, Baglietto L, Bacot F, Armasu SM, Antonenkova N, Anton-Culver H, Aben KK, Liang D, Wu X, Lu K, Hildebrandt MA. Australian ovarian cancer study G, Australian cancer S, Schildkraut JM, sellers TA, Huntsman D, Berchuck a, Chenevix-Trench G, Gayther SA, Pharoah PD, Laird PW, Goode EL, Pearce CL. epigenetic analysis leads to identification of HNF1B as a subtype-specific susceptibility gene for ovarian cancer. *Nat Commun* 2013;4:1628.
- 17 Song H, Ramus SJ, Tyrer J, Bolton KL, Gentry-Maharaj A, Wozniak E, Anton-Culver H, Chang-Claude J, Cramer DW, DiCioccio R, Dörk T, Goode EL, Goodman MT, Schildkraut JM, Sellers T, Baglietto L, Beckmann MW, Beesley J, Blaakaer J, Carney ME, Chanock S, Chen Z, Cunningham JM, Dicks E, Doherty JA, Dürst M, Ekici AB, Fenstermacher D,

- Fridley BL, Giles G, Gore ME, De Vivo I, Hillemanns P, Hogdall C, Hogdall E, Iversen ES, Jacobs IJ, Jakubowska A, Li D, Lissowska J, Lubiński J, Lurie G, McGuire V, McLaughlin J, Medrek K, Moorman PG, Moysich K, Narod S, Phelan C, Pye C, Risch H, Runnebaum IB, Severi G, Southey M, Stram DO, Thiel FC, Terry KL, Tsai Y-Y, Tworoger SS, Van Den Berg DJ, Vierkant RA, Wang-Gohrke S, Webb PM, Wilkens LR, Wu AH, Yang H, Brewster W, Ziogas A, Houlston R, Tomlinson I, Whittemore AS, Rossing MA, Ponder BAJ, Pearce CL, Ness RB, Menon U, Kiaer SK, Gronwald J, Garcia-Closas M, Fasching PA, Easton DF, Chenevix-Trench G, Berchuck A, Pharoah PDP, Gayther SA, Vivo D I, Hogdall E, AH W, Australian Cancer S, Australian Cancer (Ovarian) Study, Australian Ovarian Cancer Study Group, Ovarian Cancer Association Consortium. A genome-wide association study identifies a new ovarian cancer susceptibility locus on 9p22.2. *Nat Genet* 2009;41:996–1000.
- 18 Kuchenbaecker KB, Ramus SJ, Tyrer J, Lee A, Shen HC, Beesley J, Lawrenson K, McGuffog L, Healey S, Lee JM, Spindler TJ, Lin YG, Pejovic T, Bean Y, Li Q, Coetzee S, Hazelett D, Miron A, Southey M, Terry MB, Goldgar DE, Buys SS, Janavicius R, Dorfling CM, van Rensburg EJ, Neuhausen SL, Ding YC, Hansen TV, Jonson L, Gerdes AM, Ejlersen B, Barrowdale D, Dennis J, Benitez J, Osorio A, Garcia MJ, Komenaka I, Weitzel JN, Ganschow P, Peterlongo P, Bernard L, Viel A, Bonanni B, Peissel B, Manoukian S, Radice P, Papi L, Ottini L, Fostira F, Konstantopoulou I, Garber J, Frost D, Perkins J, Platte R, Ellis S, Embrace GAK, Schmutzler RK, Meindl A, Engel C, Sutter C, Sinilnikova OM, Collaborators GS, Damiola F, Mazoyer S, Stoppa-Lyonnet D, Claes K, De Leener K, Kirk J, Rodriguez GC, Piedmonte M, O'Malley DM, de la Hoya M, Caldes T, Aittomaki K, Nevanlinna H, Collee JM, Rookus MA, Oosterwijk JC. Consortium of Investigators of modifiers of B, BRCA. Identification of six new susceptibility loci for invasive epithelial ovarian cancer. *Nat Genet* 2015;47:164–71.
- 19 Ramus SJ, Kartsonaki C, Gayther SA, Pharoah PD, Sinilnikova OM, Beesley J, Chen X, McGuffog L, Healey S, Couch FJ, Wang X, Fredericksen Z, Peterlongo P, Manoukian S, Peissel B, Zaffaroni D, Roversi G, Barile M, Viel A, Allavena A, Ottini L, Papi L, Gismondi V, Capra F, Radice P, Greene MH, Mai PL, Andrulis IL, Glendon G, Oczelik H, Ocg TM, Gerdes AM, Kruse TA, Cruger D, Jensen UB, Caligo MA, Olsson H, Kristoffersson U, Lindblom A, Arver B, Karlsson P, Stenmark Askmal M, Borg A, Neuhausen SL, Ding YC, Nathanson KL, Domchek SM, Jakubowska A, Lubinski J, Huzarski T, Byrski T, Gronwald J, Gorski B, Cybulski C, Debniak T, Osorio A, Duran M, Tejada MI, Benitez J, Hamann U, Rookus MA, Verhoef S, Tilanus-Linthorst MA, Vreeswijk MP, Bodmer D, Ausems MG, van Os TA, Asperen CJ, Blok MJ, Meijers-Heijboer HE, Hebon E, Peock S, Cook M, Oliver C, Frost D, Dunning AM, Evans DG, Eeles R, Pichert G, Cole T, Hodgson S, Brewer C, Morrison PJ, Porteous M, Kennedy MJ, Rogers MT, Side LE, Donaldson A, Gregory H, Godwin A, Stoppa-Lyonnet D, Moncoutier V, Castera L, Mazoyer S, Barjhoux L, Bonadona V, Leroux D, Fauriv L, Lidereau R, Nogues C, Bignon YJ, Prieur F, Collonge-Rame MA, Venat-Bouvet L, Fert-Ferrer S, Collaborators GS, Miron A, Buys SS, Hopper JL, Daly MB, John EM, Terry MB, Goldgar D, Bcfr HT, Jonson L, Ejlersen B, Agnarsson BA, Offit K, Kirchhoff T, Vijai J, Dutra-Clarke AV, Przybylo JA, Montagna M, Casella C, Imyanitov EN, Janavicius R, Blanco I, Lazaro C, Moysich KB, Karlan BY, Gross J, Beattie MS, Schmutzler R, Wappenschmidt B, Meindl A, Ruehl I, Fiebig B, Sutter C, Arnold N, Deissler H, Varon-Mateeva R, Kast K, Niederacher D, Gadzicki D, Caldes T, de la Hoya M, Nevanlinna H, Aittomaki K, Simard J, Soucy P, kConFab I, Spurdle AB, Holland H, Chenevix-Trench G, Easton DF, Antoniou AC. Consortium of Investigators of modifiers of B. genetic variation at 9p22.2 and ovarian cancer risk for BRCA1 and BRCA2 mutation carriers. *J Natl Cancer Inst* 2011;103:105–16.
- 20 Norquist BM, Harrell MI, Brady MF, Walsh T, Lee MK, Gulsuner S, Bernards SS, Casadei S, Yi Q, Burger RA, Chan JK, Davidson SA, Mannel RS, DiSilvestro PA, Lankes HA, Ramirez NC, King MC, Swisher EM, Birrer MJ. Inherited mutations in women with ovarian carcinoma. *JAMA Oncol* 2016;2:482–90.
- 21 Lilyquist J, LaDuca H, Polley E, Davis BT, Shimelis H, Hu C, Hart SN, Dolinsky JS, Couch FJ, Goldgar DE. Frequency of mutations in a large series of clinically ascertained ovarian cancer cases tested on multi-gene panels compared to reference controls. *Gynecol Oncol* 2017;147:375–80.
- 22 Kotsopoulos J, Sopik V, Rosen B, Fan I, McLaughlin JR, Risch H, Sun P, Narod SA, Akbari MR. Frequency of germline PALB2 mutations among women with epithelial ovarian cancer. *Fam Cancer* 2017;16:29–34.
- 23 Wu X, Ye Y, Kiemeny LA, Sulem P, Rafnar T, Matullo G, Semina D, Yoshida T, Saeki N, Andrew AS, Dinney CP, Czerniak B, Zhang Z-feng, Kiltie AE, Bishop DT, Vineis P, Porru S, Buntinx F, Kellen E, Zeegers MP, Kumar R, Rudnai P, Gurdau E, Koppova K, Mayordomo JL, Sanchez M, Saez B, Lindblom A, de Verdier P, Steineck G, Mills GB, Schned A, Guarrera S, Polidoro S, Chang S-C, Lin J, Chang DW, Hale KS, Majewski T, Grossman HB, Thorlacius S, Thorsteinsdottir U, Aben KKH, Witjes JA, Stefansson K, Amos CI, Karagas MR, Gu J. Genetic variation in the prostate stem cell antigen gene PSCA confers susceptibility to urinary bladder cancer. *Nat Genet* 2009;41:991–5.
- 24 Wu X, Hildebrandt MA, Ye Y, Chow W-H, Gu J, Cunningham S, Zhao H, Hawk ET, Wagar E, Rodriguez A, Hamilton SR. Cohort profile: the MD anderson cancer patients and survivors cohort (MDA-CPSC). *Int J Epidemiol* 2016;45:713–713f.
- 25 Hassan MM, Bondy ML, Wolff RA, Abbruzzese JL, Vauthey J-N, Pisters PW, Evans DB, Khan R, Chou T-H, Lenzi R, Jiao L, Li D. Risk factors for pancreatic cancer: case-control study. *Am J Gastroenterol* 2007;102:2696–707.
- 26 Li H, Durbin R. Fast and accurate long-read alignment with Burrows-Wheeler transform. *Bioinformatics* 2010;26:589–95.
- 27 McKenna A, Hanna M, Banks E, Sivachenko A, Cibulskis K, Kernytsky A, Garimella K, Altshuler D, Gabriel S, Daly M, DePristo MA. The genome analysis toolkit: a MapReduce framework for analyzing next-generation DNA sequencing data. *Genome Res* 2010;20:297–303.
- 28 Wang K, Li M, Hakonarson H. ANNOVAR: functional annotation of genetic variants from high-throughput sequencing data. *Nucleic Acids Res* 2010;38:e164.
- 29 Magoč T, Salzberg SL. Flash: fast length adjustment of short reads to improve genome assemblies. *Bioinformatics* 2011;27:2957–63.
- 30 Yu Y, Hu H, Bohlender RJ, Hu F, Chen J-S, Holt C, Fowler J, Guthery SL, Scheet P, Hildebrandt MAT, Yandell M, Huff CD. XPAT: a toolkit to conduct cross-platform association studies with heterogeneous sequencing datasets. *Nucleic Acids Res* 2018;46:e32.
- 31 Landrum MJ, Lee JM, Benson M, Brown G, Chao C, Chitipiralla S, Gu B, Hart J, Hoffman D, Hoover J, Jang W, Katz K, Ovetsky M, Riley G, Sethi A, Tully R, Villamarin-Salomon R, Rubinstein W, Maglott DR. ClinVar: public archive of interpretations of clinically relevant variants. *Nucleic Acids Res* 2016;44:D862–8.
- 32 Yeo G, Burge CB. Maximum entropy modeling of short sequence motifs with applications to RNA splicing signals. *J Comput Biol* 2004;11:377–94.
- 33 Robinson JT, Thorvaldsdóttir H, Winckler W, Guttman M, Lander ES, Getz G, Mesirov JP. Integrative genomics viewer. *Nat Biotechnol* 2011;29:24–6.
- 34 Tyrer JP, Guo Q, Easton DF, Pharoah PDP. The admixture maximum likelihood test to test for association between rare variants and disease phenotypes. *BMC Bioinformatics* 2013;14:177.
- 35 Ng PC, Henikoff S. SIFT: predicting amino acid changes that affect protein function. *Nucleic Acids Res* 2003;31:3812–4.
- 36 Adzhubei I, Jordan DM, Sunyaev SR. Predicting functional effect of human missense mutations using PolyPhen-2. *Curr Protoc Hum Genet* 2013;Chapter 7:7.20.1–7.20.41.
- 37 Choi Y, Chan AP. PROVEAN web server: a tool to predict the functional effect of amino acid substitutions and indels. *Bioinformatics* 2015;31:2745–7.
- 38 Wakefield J. A Bayesian measure of the probability of false discovery in genetic epidemiology studies. *Am J Hum Genet* 2007;81:208–27.
- 39 Duraturo F, Cavallo A, Liccardo R, Cudia B, De Rosa M, Diana G, Izzo P. Contribution of large genomic rearrangements in Italian Lynch syndrome patients: characterization of a novel alu-mediated deletion. *Biomed Res Int* 2013;2013:1–7.
- 40 Ramus SJ, Harrington PA, Pye C, DiCioccio RA, Cox MJ, Garlinghouse-Jones K, Oakley-Girvan I, Jacobs IJ, Hardy RM, Whittemore AS, Ponder BAJ, Piver MS, Pharoah PDP, Gayther SA. Contribution of BRCA1 and BRCA2 mutations to inherited ovarian cancer. *Hum Mutat* 2007;28:1207–15.
- 41 Vaz-Drago R, Custódio N, Carmo-Fonseca M. Deep intronic mutations and human disease. *Hum Genet* 2017;136:1093–111.