# UCLA UCLA Previously Published Works

# Title

Large-Scale Genome-Wide Association Studies and Meta-Analyses of Longitudinal Change in Adult Lung Function

**Permalink** https://escholarship.org/uc/item/541887v9

**Journal** PLOS ONE, 9(7)

**ISSN** 1932-6203

# **Authors**

Tang, Wenbo Kowgier, Matthew Loth, Daan W <u>et al.</u>

**Publication Date** 2014

# DOI

10.1371/journal.pone.0100776

Peer reviewed

# Large-Scale Genome-Wide Association Studies and Meta-Analyses of Longitudinal Change in Adult Lung Function



Bruce M. Psaty<sup>42,53,54,55</sup>, Fernando Rivadeneira<sup>35,56</sup>, Jerome I. Rotter<sup>57</sup>, Holger Schulz<sup>34</sup>, Lewis J. Smith<sup>58</sup>, Akshay Sood<sup>59</sup>, John M. Starr<sup>17,60</sup>, David P. Strachan<sup>61</sup>, Alexander Teumer<sup>62</sup>, André G. Uitterlinden<sup>35,56</sup>, Henry Völzke<sup>63</sup>, Arend Voorman<sup>64</sup>, Louise V. Wain<sup>5,6</sup>, Martin T. Wells<sup>65</sup>, Jemma B. Wilk<sup>28,66</sup>, O. Dale Williams<sup>67</sup>, Susan R. Heckbert<sup>42,53,54</sup>, Bruno H. Stricker<sup>3,4</sup>, Stephanie J. London<sup>7</sup>, Myriam Fornage<sup>30,50‡</sup>, Martin D. Tobin<sup>5,6‡</sup>, George T. O'Connor<sup>28,68‡</sup>, Ian P. Hall<sup>8‡</sup>,

Patricia A. Cassano<sup>1,69</sup>\*<sup>‡</sup>

1 Division of Nutritional Sciences, Cornell University, Ithaca, New York, United States of America, 2 Ontario Institute for Cancer Research and Biostatistics Division, Dalla Lana School of Public Health, University of Toronto, Toronto, Ontario, Canada, 3 Department of Epidemiology, Erasmus Medical Center, Rotterdam, the Netherlands, 4 Netherlands Healthcare Inspectorate. The Haque, the Netherlands, 5 University of Leicester, Genetic Epidemiology Group, Department of Health Sciences, Leicester, United Kingdom, 6 National Institute for Health Research (NIHR) Leicester Respiratory Biomedical Research Unit, Glenfield Hospital, Leicester, United Kingdom, 7 Epidemiology Branch, National Institute of Environmental Health Sciences, National Institutes of Health, U.S. Department of Health and Human Services, Research Triangle Park, North Carolina, United States of America, 8 Division of Respiratory Medicine, University Hospital of Nottingham, Nottingham, United Kingdom, 9 Computational Medicine Core, Center for Lung Biology, Division of Pulmonary & Critical Care Medicine, Department of Medicine, University of Washington, Seattle, Washington, United States of America, 10 Icelandic Heart Association, Kopavogur, Iceland, 11 University of Iceland, Reykjavik, Iceland, 12 Department of Biostatistics, Bloomberg School of Public Health, Johns Hopkins University, Baltimore, Maryland, United States of America, 13 Department of Medicine, School of Medicine, Johns Hopkins University, Baltimore, Maryland, United States of America, 14 Laboratory of Epidemiology, Demography, and Biometry, National Institute on Aging, National Institutes of Health, Bethesda, Maryland, United States of America, 15 Institute of Genetic Epidemiology, Helmholtz Zentrum München - German Research Center for Environmental Health, Neuherberg, Germany, 16 Department of Thoracic Surgery and Division of Epidemiology, Vanderbilt University Medical Center, Nashville, Tennessee, United States of America, 17 Centre for Cognitive Ageing and Cognitive Epidemiology, University of Edinburgh, Edinburgh, United Kingdom, 18 Division of General Medicine, Pulmonary, Allergy and Critical Care, Department of Medicine, College of Physicians and Surgeons, Columbia University, New York, New York, United States of America, 19 Department of Epidemiology, Mailman School of Public Health, Columbia University, New York, New York, United States of America, 20 Department of Respiratory Medicine, Ghent University Hospital, Ghent, Belgium, 21 Department of Respiratory Medicine, Erasmus Medical Center, Rotterdam, the Netherlands, 22 22 Department of Biostatistics, University of North Carolina at Chapel Hill, Chapel Hill, North Carolina, United States of America, 23 Swiss Tropical and Public Health Institute, Basel, Switzerland, 24 University of Basel, Basel, Switzerland, 25 Medical Genetics Section, University of Edinburgh Molecular Medicine Centre and MRC Institute of Genetics and Molecular Medicine, Western General Hospital, Edinburgh, United Kingdom, 26 Department of Psychology, University of Edinburgh, Edinburgh, United Kingdom, 27 Biostatistics Department, Boston University School of Public Health, Boston, Massachusetts, United States of America, 28 The National Heart, Lung, and Blood Institute's Framingham Heart Study, Framingham, Massachusetts, United States of America, 29 Department of Medical Sciences, Molecular Epidemiology and Science for Life Laboratory, Uppsala University, Uppsala, Sweden, 30 Institute of Molecular Medicine, University of Texas Health Science Center at Houston, Houston, Texas, United States of America, 31 Gillings School of Global Public Health, Department of Epidemiology, University of North Carolina at Chapel Hill, Chapel Hill, North Carolina, United States of America, 32 Department of Internal Medicine B; Pneumology, Cardiology, Intensive Care Medicine; Field of Research: Pneumology and Pneumological Epidemiology, University Medicine Greifswald, Greifswald, Germany, 33 Behavioral Health Epidemiology Program, Research Triangle Institute, Research Triangle Park, North Carolina, United States of America, 34 Institute of Epidemiology I, Helmholtz Zentrum München - German Research Center for Environmental Health, Neuherberg, Germany and Comprehensive Pneumology Center Munich (CPC-M), Member of the German Center for Lung Research, Munich, Germany, 35 Netherlands Consortium for Healthy Aging, Rotterdam, the Netherlands, 36 Wellcome Trust Centre for Human Genetics, University of Oxford, Oxford, UK and Department of Biostatistics, University of Liverpool, Liverpool, United Kingdom, 37 School of Medicine and Pharmacology, University of Western Australia, Perth, Western Australia, Australia, 38 Institute and Outpatient Clinic for Occupational, Social and Environmental Medicine, Ludwig-Maximilians-Universität, Munich, Germany, 39 Institute of General Practice, University Hospital Klinikum rechts der Isar, Technische Universität München, Munich, Germany, 40 Institute of Epidemiology I, Helmholtz Zentrum München - German Research Center for Environmental Health, Neuherberg, Germany, 41 Sticht Center on Aging, Wake Forest School of Medicine, Winston-Salem, North Carolina, United States of America, 42 Cardiovascular Health Research Unit, University of Washington, Seattle, Washington, United States of America, 43 Department of Medical Sciences, Uppsala University, Uppsala, Sweden, 44 Broad Institute of MIT and Harvard, Cambridge, Massachusetts, United States of America, 45 Department of Epidemiology and Prevention, Division of Public Health Sciences, Wake Forest School of Medicine, Winston-Salem, North Carolina, United States of America, 46 Department of Biostatistical Sciences, Division of Public Health Sciences, Wake Forest School of Medicine, Winston-Salem, North Carolina, United States of America, 47 Department of Statistics, University of Auckland, Auckland, New Zealand, 48 School of Social and Community Medicine, University of Bristol, Bristol, United Kingdom, 49 College of Pharmacy, University of Tennessee Health Science Center, Memphis, Tennessee, United States of America, 50 Human Genetics Center, School of Public



PLOS ONE

Health, University of Texas Health Science Center at Houston, Houston, Texas, United States of America, **51** Epidemiology and Obstetrics & Gynaecology, University of Toronto, Toronto, Ontario, Canada, **52** Samuel Lunenfeld Research Institute, Toronto, Ontario, Canada, **53** Department of Epidemiology, University of Washington, Seattle, Washington, United States of America, **54** Group Health Research Institute, Group Health Cooperative, Seattle, Washington, United States of America, **55** Department of Medicine, University of Washington, Seattle, Washington, United States of America, **55** Department of Medicine, University of Washington, Seattle, Washington, United States of America, **56** Department of Internal Medicine, Erasmus Medical Center, Rotterdam, the Netherlands, **57** Institute for Translational Genomics and Population Sciences, Los Angeles Biomedical Research Institute and Department of Pediatrics at Harbor-UCLA Medical Center, Torrance, California, United States of America, **58** Northwestern University Feinberg School of Medicine, Chicago, Illinois, United States of America, **59** University of New Mexico, Albuquerque, New Mexico, United States of America, **60** Alzheimer Scotland Dementia Research Centre, University of Edinburgh, United Kingdom, **61** Division of Population Health Sciences and Education, St George's, University of London, London, United Kingdom, **62** Department for Genetics and Functional Genomics, Interfaculty Institute for Genetics and Functional Genomics, University Medicine Greifswald, Greifswald, Germany, **64** Department of Biostatistics, University of Washington, Seattle, Washington, United States of America, **65** Department of Statistical Science, Cornell University, Ithaca, New York, United States of America, **66** Division of Aging, Department of Medicine, Brigham and Women's Hospital and Harvard Medical School, Boston, Massachusetts, United States of America, **69** Department of Health Care Policy and Research, Division of Biostatistics and University School of Medicine, Boston, M

# Abstract

**Background:** Genome-wide association studies (GWAS) have identified numerous loci influencing cross-sectional lung function, but less is known about genes influencing longitudinal change in lung function.

*Methods:* We performed GWAS of the rate of change in forced expiratory volume in the first second ( $FEV_1$ ) in 14 longitudinal, population-based cohort studies comprising 27,249 adults of European ancestry using linear mixed effects model and combined cohort-specific results using fixed effect meta-analysis to identify novel genetic loci associated with longitudinal change in lung function. Gene expression analyses were subsequently performed for identified genetic loci. As a secondary aim, we estimated the mean rate of decline in  $FEV_1$  by smoking pattern, irrespective of genotypes, across these 14 studies using meta-analysis.

**Results:** The overall meta-analysis produced suggestive evidence for association at the novel *IL16/STARD5/TMC3* locus on chromosome 15 ( $P = 5.71 \times 10^{-7}$ ). In addition, meta-analysis using the five cohorts with  $\geq 3$  FEV<sub>1</sub> measurements per participant identified the novel *ME3* locus on chromosome 11 ( $P = 2.18 \times 10^{-8}$ ) at genome-wide significance. Neither locus was associated with FEV<sub>1</sub> decline in two additional cohort studies. We confirmed gene expression of *IL16, STARD5,* and *ME3* in multiple lung tissues. Publicly available microarray data confirmed differential expression of all three genes in lung samples from COPD patients compared with controls. Irrespective of genotypes, the combined estimate for FEV<sub>1</sub> decline was 26.9, 29.2 and 35.7 mL/year in never, former, and persistent smokers, respectively.

**Conclusions:** In this large-scale GWAS, we identified two novel genetic loci in association with the rate of change in  $FEV_1$  that harbor candidate genes with biologically plausible functional links to lung function.

Citation: Tang W, Kowgier M, Loth DW, Soler Artigas M, Joubert BR, et al. (2014) Large-Scale Genome-Wide Association Studies and Meta-Analyses of Longitudinal Change in Adult Lung Function. PLoS ONE 9(7): e100776. doi:10.1371/journal.pone.0100776

Editor: Lin Chen, The University of Chicago, United States of America

Received January 2, 2014; Accepted April 17, 2014; Published July 1, 2014

This is an open-access article, free of all copyright, and may be freely reproduced, distributed, transmitted, modified, built upon, or otherwise used by anyone for any lawful purpose. The work is made available under the Creative Commons CC0 public domain dedication.

Funding: The AGES-Reykjavik Study is funded by NIH contract N01-AG-12100, the NIA Intramural Research Program, Hjartavernd (the Icelandic Heart Association), and the Althingi (the Icelandic Parliament). The Atherosclerosis Risk in Communities Study is carried out as a collaborative study supported by National Heart, Lung, and Blood Institute contracts (HHSN268201100005C, HHSN268201100006C, HHSN268201100007C, HHSN268201100008C, HHSN268201100009C HHSN268201100010C, HHSN268201100011C, and HHSN268201100012C), R01HL087641, R01HL59367 and R01HL086694; National Human Genome Research Institute contract U01HG004402; and National Institutes of Health contract HHSN268200625226C. The authors thank the staff and participants of the ARIC study for their important contributions. Infrastructure was partly supported by Grant Number UL1RR025005, a component of the National Institutes of Health and NIH Roadmap for Medical Research. Supported in part by the Intramural Research Program of the NIH, National Institute of Environmental Health Sciences ZO1 ES43012. The authors acknowledge use of phenotype and genotype data from the British 1958 Birth Cohort DNA collection, funded by the Medical Research Council grant G0000934 and the Wellcome Trust grant 068545/Z/02 (http://www.b58cgene.sgul.ac.uk/). Genotyping for the B58C-WTCCC subset was funded by the Wellcome Trust grant 076113/ B/04/Z. The B58C-T1DGC genotyping utilized resources provided by the Type 1 Diabetes Genetics Consortium, a collaborative clinical study sponsored by the National Institute of Diabetes and Digestive and Kidney Diseases (NIDDK), National Institute of Allergy and Infectious Diseases (NIAID), National Human Genome Research Institute (NHGRI), National Institute of Child Health and Human Development (NICHD), and Juvenile Diabetes Research Foundation International (JDRF) and supported by U01 DK062418. B58C-T1DGC GWAS data were deposited by the Diabetes and Inflammation Laboratory, Cambridge Institute for Medical Research (CIMR), University of Cambridge, which is funded by Juvenile Diabetes Research Foundation International, the Wellcome Trust and the National Institute for Health Research Cambridge Biomedical Research Centre; the CIMR is in receipt of a Wellcome Trust Strategic Award (079895). The B58C-GABRIEL genotyping was supported by a contract from the European Commission Framework Programme 6 (018996) and grants from the French Ministry of Research. The 1994 Busselton follow-up Health Study was supported by Healthways, Western Australia. The Busselton Health Study is supported by The Great Wine Estates of the Margaret River region of Western Australia. The study gratefully acknowledges the assistance of the Western Australian DNA Bank (NHMRC Enabling Facility) with DNA samples and the support provided by The Ark at University of Western Australia for this study. The Coronary Artery Risk Development in Young Adults (CARDIA) study was funded by contracts N01-HC-95095, N01-HC-48047, N01-HC-48048, N01-HC-48049, N01-HC-48050, N01-HC-45134, N01-HC-05187, N01-HC-45205, and N01-HC-45204 from NHLBI to the CARDIA investigators. Genotyping of the CARDIA participants was supported by grants U01-HG-004729, U01-HG-004446, and U01-HG-004424 from the NHGRI. Statistical analyses were supported by grants U01-HG-004729 and R01-HL-084099 to MF. This Cardiovascular Health Study (CHS) research was supported by NHLBI contracts HISN268201200036C, HISN26820080007C, N01HC55222, N01HC8S079, N01HC8S080, N01HC8S081, N01HC8S082, N01HC8S083, N01HC8S086, and HISN26820096009C; and NHLBI grants HL080295, HL087652, HL105756, HL103612, and HL085251 with additional contribution from the National Institute of Neurological Disorders and Stroke (NINDS). Additional support was provided through AG023629 from the National Institute on Aging (NIA). A full list of CHS investigators and institutions can be found at http://chs-nhlbi.org. The provision of genotyping data was supported in part by the National Center for Advancing Translational Sciences, CTSI grant UL1TR000124, and the National Institute of Diabetes and Digestive and Kidney Disease Diabetes Research Center (DRC) grant DK063491 to the Southern California Diabetes Endocrinology Research Center. Framingham Heart Study (FHS) research was conducted in part using data and resources of the NHLBI and Boston University School of Medicine. The analyses reflect intellectual input and resource development from the FHS investigators participating in the SNP Health Association Resource (SHARe) project. This work was partially supported by NHLBI (contract no. N01-HC-25195) and its contract with Affymetrix for genotyping services (contract no. N02-HL-6-4278). A portion of this research utilized the Linux Cluster for Genetic Analysis (LinGA-II) funded by the Robert Dawson Evans Endowment of the Department of Medicine at Boston University School of Medicine and Boston Medical Center. JBW was supported by a Young Clinical Scientist Award from the Flight Attendant Medical Research Institute (FAMRI). The Health, Aging, and Body Composition Study was supported by NIA contracts N01AG62101, N01AG2103, and N01AG62106, NIA grant R01-AG028050, NINR grant R01-NR012459, and in part by the Intramural Research Program of the NIA, NIH. The genome-wide association study was funded by NIA grant 1R01AG032098-01A1 to Wake Forest Health Sciences, and genotyping services were provided by the Center for Inherited Disease Research, which is fully funded through a federal contract from the National Institutes of Health to The Johns Hopkins University, contract number HHSN268200782096C. This research was further supported by RC1AG035835. The KORA study was funded by the Helmholtz Zentrum München German Research Center for Environmental Health, German Federal Ministry of Education and Research, State of Bavaria, Munich Center of Health Sciences (MC Health), Ludwig-Maximilians-Universität, as part of LMUinnovativ, and Competence Network ASCONET, subnetwork COSYCONET (FKZ 01GI0882). The Lothian Birth Cohorts 1921 and 1936 were funded by the Lifelong Health and Wellbeing Initiative (BBSRC, EPSRC, ESRC and MRC). The authors thank the cohort participants and team members who contributed to these studies. Phenotype collection in the Lothian Birth Cohort 1921 was supported by the BBSRC, The Royal Society and The Chief Scientist Office of the Scottish Government. Phenotype collection in the Lothian Birth Cohort 1936 was supported by Research Into Ageing (continues as part of Age UK The Disconnected Mind project). Genotyping of the cohorts was funded by the UK Biotechnology and Biological Sciences Research Council (BBSRC). The work was undertaken by the University of Edinburgh Centre for Cognitive Ageing and Cognitive Epidemiology, part of the cross council Lifelong Health and Wellbeing Initiative (G0700704/84698). Funding from the BBSRC, Engineering and Physical Sciences Research Council (EPSRC), Economic and Social Research Council (ESRC), and MRC is gratefully acknowledged. The Lung Health Study (LHS) was supported by GENEVA (U01HG 004738), and by the Mary Beryl Patch Turnbull Scholar Program (KCB, in part). The PIVUS study was funded by the Swedish Foundation for Strategic Research (ICA08-0047), the Swedish Research Council (2012-1397), the Swedish Heart-Lung Foundation (20120197), the Swedish Society of Medicine, and Uppsala University. CML is a Wellcome Trust Research Career Development Fellow (086596/Z/08/Z). The computations were performed on resources provided by SNIC through Uppsala Multidisciplinary Center for Advanced Computational Science (UPPMAX) under Project p2013056. APM acknowledges funding from the Wellcome Trust under grants WT098017, WT064890 and WT090532, and APM is a Senior Research Fellow in Basic and Biomedical Science (grant number WT098017). The Rotterdam Studies were funded by the Netherlands Organization of Scientific Research NWO Investments, nr. 175.010.2005.011, 911-03-012, Research Institute for Diseases in the Elderly, 014-93-015; RIDE2, the Netherlands Genomics Initiative (NGI)/Netherlands Organization for Scientific Research (NWO) project nr. 050-060-810, the Ministry of Education, Culture and Science, the Ministry for Health, Welfare and Sports, the European Commission (DG XII) Municipality of Rotterdam. SAPALDIA was supported by the Swiss National Science Foundation (grants no 33CS30-134276/1, 33CSCO-108796, 3247BO-104283, 3247BO-104288, 3247BO-104284, 3247-065896, 3100-059302, 3200-052720,3200-042532, 4026-028099, 3233-054996, PDFMP3-123171), the Federal Office for Forest, Environment, and Landscape, the Federal Office of Public Health, the Federal Office of Roads and Transport, the canton's government of Aargau, Basel-Stadt, Basel-Land, Geneva, Luzern, Ticino, Valais, Zurich, the Swiss Lung League, the canton's Lung League of Basel Stadt/Basel Landschaft, Geneva, Ticino, Valais and Zurich, Schweizerische Unfallversicherungsanstalt (SUVA), Freiwillige Akademische Gesellschaft, UBS Wealth Foundation, Talecris Biotherapeutics GmbH, Abbott Diagnostics. Genotyping in the GABRIEL framework was supported by grants European Commission 018996 and Wellcome Trust WT 084703MA. SHIP is part of the Community Medicine Research net of the University of Greifswald, Germany, which is funded by the Federal Ministry of Education and Research, the Ministry of Cultural Affairs as well as the Social Ministry of the Federal State of Mecklenburg-West Pomerania, and the network 'Greifswald Approach to Individualized Medicine (GANI\_MED)' funded by the Federal Ministry of Education and Research, and the German Asthma and COPD Network (COSYCONET), under 01ZZ9603, 01ZZ0103, 01ZZ0403, 03IS2061A, and BMBP 01GI0883. Genome-wide data have been supported by the Federal Ministry of Education and Research and a joint grant from Siemens Healthcare, Erlangen, Germany and the Federal State of Mecklenburg- West Pomerania (03ZIK012). The University of Greifswald is a member of the 'Center of Knowledge Interchange' program of the Siemens AG and the Caché Campus program of the InterSystems GmbH. The research undertaken by MDT, LVW and MSA was partly funded by the National Institute for Health Research (NIHR). The views expressed are those of the author(s) and not necessarily those of the NHS, the NIHR or the Department of Health. MDT holds a Medical Research Council Senior Clinical Fellowship (G0902313). The expression analysis undertaken by IPH and EH was funded by the Medical Research Council of UK (grant no. G1000861). The University of Greifswald is a member of the 'Center of Knowledge Interchange' program of the Siemens AG and the Caché Campus program of the InterSystems GmbH. The funders had no role in study design, data collection and analysis, decision to publish, or preparation of the manuscript.

Competing Interests: All authors have read the journal's policy, and 57 of the 81 authors have declared that no competing interests exist. The following 24 authors have possible conflicts, as follows: Dr. Aldrich reports grants from NIH, during the conduct of the study; Dr. Barnes reports grants from NIH, during the conduct of the study; Dr. Barr reports grants from NIH and US-EPA, during the conduct of the study; Dr. Couper reports grants from NIH, during the conduct of the study; Dr. Deary reports grants from Age UK, grants from BBSRC, during the conduct of the study; Dr. Dupuis reports grants from Boston University, during the conduct of the study; Dr. Fall reports personal fees from MSD (Merck), outside the submitted work; Dr. Gudnason reports other from NIH, during the conduct of the study; Dr. Gläser reports grants from BMBF (German Ministry for Research and Education), during the conduct of the study; personal fees from Actelion Pharma, personal fees from Novartis Pharma, personal fees from GSK, personal fees from Pfizer, personal fees from Boehringer Ingelheim, personal fees from Bayer Pharma, all those apply outside of the submitted work; Dr. Hall reports grants received from MRC and Pfizer, and Vertex sponsored lecture at ERS, outside the submitted work; Dr. Hodge reports grants from The Medical Research Council, UK, during the conduct of the study; Dr. Koch reports grants from BMBF, during the conduct of the study, travel fees from Actelion Pharma, Pfizer, Bayer Pharma, the German Academic Exchange Service, and the Research Network for Community Medicine of the University of Greifswald, one research prize of the Society of Internal Medicine Mecklenburg-Vorpommern, Germany, outside the submitted work; Dr. Lahousse reports grant from Belgian Society of Pneumology, during the conduct of the study; Dr. London is funded in full by the Division of Intramural Research, NIEHS, NIH, DHHS; Dr. Lumley reports grants from NIH, during the conduct of the study; Dr. Mathias reports grants from NIH, during the conduct of the study; Dr. Meibohm reports grants from NIH, during the conduct of the study: Dr. O'Connor reports personal fees from Sunovion. Inc., outside the submitted work; Dr. Psaty reports grants from NIH, during the conduct of the study, and Dr. Psaty serves on the DSMB for a clinical trial of a device, which is funded by the manufacturer (Zoll LifeCor), and he is on the Steering Committee of the Yale Open Data Access Project funded by Johnson & Johnson; Dr. L Smith reports personal fees as member of Merck Data Safety and Monitoring Board, outside the submitted work; Dr. Tobin reports grants from Medical Research Council grant G0902313, grants from National Institute for Health Research (NIHR), Leicester Respiratory Biomedical Research Unit, during the conduct of the study (the views expressed are those of the authors and not necessarily those of the NHS, the NIHR or the Department of Health), grants from Pfizer for collaborative research project (on rare sequence variants and the smoking resistant lung, Nov. 2010 to Nov 2012), outside the submitted work; Dr. Wain reports grants from Pfizer collaborative research project (onrare sequence variants and the smoking resistant lung, Nov. 2010 to Nov 2012), outside the submitted work; Dr. Wilk reports grants from FAMRI, grants from NIH, during the conduct of the study; personal fees from Pfizer, outside the submitted work. This does not alter the authors' adherence to all PLOS ONE policies on sharing data and materials.

\* Email: pac6@cornell.edu

These authors are joint first authors on this work.These authors are joint last authors on this work.

### Introduction

Forced expiratory volume in the first second (FEV<sub>1</sub>) is a reliable spirometric parameter that reflects the physiological state of the lungs and airways. Reduced FEV<sub>1</sub> relative to forced vital capacity (FVC), is a defining feature of chronic obstructive pulmonary disease (COPD), a leading cause of death globally.[1] FEV<sub>1</sub> is also a predictor of morbidity and mortality in the general population.[2,3] Lung function reaches its peak in early adulthood, followed by a plateau, and then subsequently declines. As first reported by Fletcher and Peto,[4] decline in lung function is accelerated in smokers, leading to increased risks of COPD and premature death. While cigarette smoking is a key risk factor for accelerated loss of lung function, genetic variation is hypothesized to also play an important role.[5,6] Family and twin studies of the longitudinal change in lung function report heritability estimates between 10 and 39%.[7,8]

Recent large-scale genome-wide association studies (GWAS) identified 26 novel loci for cross-sectional lung function,[9–11] demonstrating the power of GWAS with large sample size to identify common genetic variants with modest effect sizes. However, cross-sectional measurements in adults reflect the combination of maximal attained lung growth and subsequent decline. GWAS that specifically study the longitudinal change in lung function are needed to distinguish the genetic contributions to age-related decline. To date, only one population-based GWAS meta-analysis of longitudinal change in lung function has been reported.[12] Separate analyses were conducted in 1,441 asthmatic and 2,667 non-asthmatic participants; association was found at one novel locus in each analysis, though only the locus in non-asthmatics replicated.

In this study, we conducted primary GWAS of the rate of change in FEV<sub>1</sub> in each of 14 population-based cohort studies from the Cohorts for Heart and Aging Research in Genomic Epidemiology (CHARGE) and SpiroMeta consortia, comprising 27,249 adult participants of European ancestry and 62,130 FEV<sub>1</sub> measurements. We then performed meta-analysis of the cohort-specific results, followed up our most statistically significant associations in the AGES-Reykjavík cohort study and the Lung Health Study (LHS) for corroborative evidence, and explored the biological basis for identified associations using cell-specific gene

expression studies, and expression quantitative trait loci (eQTL) look-up.

# Methods

#### Study populations

All 14 cohort studies are members of the CHARGE or SpiroMeta Consortium (Table 1). The respective local Institutional Review Boards approved all study protocols, and written informed consent for genetic studies was obtained from all participants. Spirometry tests were performed at baseline and at least one follow-up time point by trained technicians and in accordance with the American Thoracic Society or European Respiratory Society recommendations (Methods S1 in File S1 for further details).[13] FEV<sub>1</sub> measurements meeting acceptability criteria were included in the current study.

Studies performed genotyping following standard quality control measures; imputation was conducted based on the HapMap CEU reference panel to generate genotype dosages for  $\sim 2.5$  million autosomal single nucleotide polymorphisms (SNPs) (Table S1 in File S1).

# Statistical analysis

For the analysis of repeated measurement data such as longitudinal change in lung function, mixed effects models offer more flexibility and statistical power than alternative approaches; the model allows for the use of unbalanced data and does not exclude individuals with incomplete records. Each cohort study performed the GWAS using a linear mixed effects model. The model included a random intercept and a random slope, and fixed effects for time (a continuous variable quantifying the time distance between each FEV1 measurement and baseline), SNP and its interaction with time (SNP-by-time), baseline age, gender, standing height, smoking pattern during follow-up and its interaction with time (smoking-by-time), baseline smoking packyears, study site, and principal components for genetic ancestry (as needed). Cohort-specific results for the SNP-by-time interaction term, which estimates the effect of genotype on the rate of change in FEV<sub>1</sub>, were shared, and two meta-analyses, one using all 14 studies and the other using the five studies with  $\geq 3$  FEV<sub>1</sub>

Table 1. Baseline chara	acteristics of cohort st	tudies included in the	e meta-analysis <sup>*</sup> .				
Cohort:	ARIC	B58C	BHS	CARDIA	CHS	FHS	Health ABC
No. of participants	8,242	827	1,009	1,492	3,159	3,230	1,586
No. of FEV <sub>1</sub> measurements	15,582	1,653	3,073	6,140	7,140	11,275	4,426
No. of FEV <sub>1</sub> per person	2	2	7	5	£	5	4
Follow-up duration, yr	5.6	10	29	20.1	7.9	14.7	9.5
Males, %	46.5	48.6	41.6	46.9	39	47	52.7
Baseline age, yr	54.6 (5.7)	35.0 (0.2)	37.5 (12.8)	27.5 (2.3)	72.3 (5.4)	50.9 (10.3)	73.8 (2.8)
Baseline height, cm	168.7 (9.4)	170.1 (9.5)	168.1 (8.9)	171.2 (9.3)	164.6 (9.4)	168.4 (9.3)	166.8 (9.3)
Current smokers, %	20.2	27.1	20.9	24.8	10.8	24.6	6.4
Former smokers, %	32.6	41.5	16.5	17.3	35.7	39.8	49.9
Baseline pack-years $^{\dagger}$	25.9 (21.7)	7.5 (11.4)	8.2 (17.8)	6.0 (6.5)	33.2 (27.0)	25.4 (21.3)	36.8 (32.2)
Baseline FEV <sub>1</sub> , mL	2972 (758)	3631 (744)	3230 (927)	3818 (781)	2123 (652)	2989 (806)	2308 (649)
Baseline FEV <sub>1</sub> /FVC, %	74.1 (7.1)	80.6 (5.8)	78.2 (9.2)	81.6 (6.5)	70.5 (10.5)	75.7 (8.0)	74.7 (7.8)
Cohort:	KORA	LBC1921	LBC1936	PIVUS	RS	SAPALDIA	SHIP
No. of participants	890	512	1,002	818	1,321	1,401	1,760
No. of FEV <sub>1</sub> measurements	1,597	706	1,790	1,469	2,016	2,692	2,571
No. of FEV1 per person	2	2	2	2	2	2	2
Follow-up duration, yr	3.2	8.9	4.8	5.8	8.3	10.9	7.9
Males, %	47.2	41.4	50.8	49.9	45.1	48	49.4
Baseline age, yr	53.8 (4.5)	79.1 (0.6)	69.6 (0.8)	70.2 (0.2)	74.4 (5.6)	41.1 (11.2)	52.4 (13.6)
Baseline height, cm	169.3 (9.3)	163.2 (9.4)	166.5 (8.9)	169.0 (9.3)	167.3 (9.1)	169.4 (9.1)	169.5 (9.7)
Current smokers, %	20.5	7.0	12.9	10.2	11.1	26.9	32.8
Former smokers, %	40.9	50.4	42.6	39.6	56.7	25.8	23.8
Baseline pack-years†	11.2 (17.1)	15.3 (22.3)	16.9 (25.8)	14.3 (15.8)	25.7 (21.3)	17.4 (18.0)	11.3 (11.9)
Baseline FEV <sub>1</sub> , mL	3280 (792)	1887 (625)	2371 (687)	2452 (682)	2215 (652)	3516 (861)	3238 (876)
Baseline FEV <sub>1</sub> /FVC, %	77.5 (6.2)	79.0 (11.8)	78.3 (10.2)	76.0 (10.0)	74.8 (7.9)	78.5 (8.2)	83.1 (6.6)
Definition of abbreviations: ARI Health Study = FHS, Framingh Birth Cohort 1936; PIVUS = Pro Health in Pomerania. *Data are presented as mean ( *Pack-years are calculated amo doi:10.1371/journal.pone.01000	C = Atherosclerosis Risk in nam Heart Study; Health AB <sup>1</sup> sspective Investigation of th (5D) unless otherwise indica ng current and former smo 776.t001	Communities, B58C = Briti C = Health, Aging, and Boc ie Vasculature in Uppsala Sei ated; total no. participants okers at study baseline.	ish 1958 Birth Cohort; BHS y Composition; KORA = C niors; RS = Rotterdam Stuc = 27,249, total no. FEV <sub>1</sub> m	= Busselton Health Study coperative Health Research dy SAPALDIA = Swiss Study dy casurements = 62,130.	; CARDIA = Coronary Arter n in the Region of Augsburg y on Air Pollution and Lung	y Risk Development in Young J; LBC1921 = Lothian Birth Cc Diseases in Adults; SD = stanc	Adults; CHS = Cardiovascular hort 1921; LBC1936 = Lothian lard deviation; SHIP = Study of

\_\_\_\_\_

5

**Table 2.** Model estimates for the rate of change in FEV<sub>1</sub> in never smokers and effects of other smoking patterns (compared with never smokers) on the rate of change in FEV<sub>1</sub> (mL/year)<sup>\*</sup> L.

1

1

Study	Annual FEV <sub>1</sub> cha	inge in never smokers	Additional Effect	<sup>™</sup> of smoking patterns on ann	al FEV <sub>1</sub> change			
	(referent group)		Persistent smoke	rs	Intermittent smokers		Former smokers	
	ß	SE	ß	SE	Ø	SE	đ	SE
ARIC	- 14.0	1.3	-12.4	1.7	-5.5	2.1	-5.3	1.4
B58C	- 29.6	1.5	-9.4	2.8	-2.2	3.4	-3.0	3.0
BHS	-23.0	1.0	-20.0	3.0	-8.0	2.0	- 9.0	2.0
CARDIA	-26.4	0.5	-6.7	1.3	-0.2	1.0	1.0	1.2
CHS	-35.0	1.1	-2.2	3.3	-4.6	2.2	-2.4	1.7
FHS	-26.0	0.6	-8.1	1.3	-2.9	1.0	-1.1	0.8
Health ABC	- 39.7	1.3	-12.9	6.1	-6.8	4.4	-2.6	1.7
KORA	- 22.1	3.7	2.2	7.2	-10.4	9.3	2.8	5.2
LBC1921	- 10.0	3.6	-11.6	15.7	2.8	14.4	- 18.8	4.9
LBC1936	- 32.3	3.6	-19.0	9.6	40.1	16.8	4.3	5.3
PIVUS	-21.1	2.5	-15.9	8.2	-21.7	13.4	-3.9	3.9
RS	- 27.5	3.7	-1.8	9.0	9.3	8.6	-4.6	4.5
SAPALDIA	-29.7	1.2	-7.4	2.3	-2.0	2.6	2.8	2.1
SHIP	-31.8	2.8	-0.4	10.9	- 0.1	3.9	- 15.0	7.3
14-cohort meta- analyzed estimate	-26.9	0.3	-8.8	0.7	-2.6	0.6	-2.3	0.5
Definition of abbrevi	ations: ARIC = Ather	osclerosis Risk in Communi	ities; B58C = British	1958 Birth Cohort; BHS = Bussel	ton Health Study; CARDIA = Coro	nary Artery Risk Develo	pment in Young Adul	s; CHS = Cardiovascul

F Lothian = Study of Data shown are the effect estimates ( $\beta$  and SE) of the time and smoking-by-time interaction terms in the preliminary mixed effects model fully adjusted for all specified variables except the SNP terms. Time represents the rate of П = Health, Aging, and Body Composition; KORA = Cooperative Health Research in the Region of Augsburg; LBC1921 = Lothian Birth Cohort 1921; LBC1936 = standard error; SHIP Birth Cohort 1936; PIVUS = Prospective Investigation of the Vasculature in Uppsala Seniors; RS = Rotterdam Study; SAPALDIA = Swiss Study on Air Pollution and Lung Diseases in Adults; SE Health Study; FHS = Framingham Heart Study; Health ABC Health in Pomerania.

change in FEV, in never smokers and the smoking-by-time interaction term represents the effects of the other three smoking patterns on the rate of change in FEV, compared with never smokers. Smoking categories are defined as persistent (smoke throughout follow-up), intermittent (stop and/or start smoking during follow-up) and former (smoke only prior to start of follow-up).

<sup>1</sup> Effect estimates in smoking categories are added to estimates in never smokers to compute the actual rate of change in each group (for example, in ARIC, the point estimate of the rate of change in FEV, in persistent smokers was -14.0 - 12.4 = -26.4 mL/year

doi:10.1371/journal.pone.0100776.t002

SNP	Chr	Position	Closest Gene(s)	Coded Allele	Frequency	β	SE	<i>P</i> Value
rs12137475	-	44059735	ST3GAL3	Т	0.11	-3.5	0.8	$3.90 \times 10^{-6}$
rs766488	-	61583103	NFIA	Α	0.31	1.4	0.3	$6.60 \times 10^{-6}$
rs17698444	-	215483178	ESRRG/GPATCH2	C	0.89	-2.2	0.5	$2.62 \times 10^{-6}$
rs12692550	2	159958017	BAZ2B	Т	0.17	-1.7	0.4	$5.16 \times 10^{-6}$
rs2260722	13	113236292	TMCO3	Α	0.72	-1.5	0.3	$1.83 \times 10^{-6}$
rs4077833	15	79419738	IL16/STARD5/TMC3	C	0.10	2.3	0.5	$5.71 \times 10^{-7}$
rs8027498	15	89595638	SV2B	A	0.25	1.4	0.3	$9.41 \times 10^{-6}$
rs8051319	16	15794449	1 1HAW	Т	0.60	1.7	0.3	$5.12 \times 10^{-6}$
rs740557	17	62451139	CACNG4	U	0.85	-2.3	0.5	$3.59 \times 10^{-6}$
Definition of abbreviations: C Data reported are the meta-	hr = chroi analvsis res	mosome; SE = standard err sults of the SNP-bv-time inte	ror; SNP = single-nucleotide polymorphism. raction term from the GWAS mixed effects mo	del. A positive B-coefficient indic	ates an attenuation of FEV,	decline and a	negative B-c	oefficient an acceleration o

GWAS of Longitudinal Change in Adult Lung Function

measurements per participant, were performed using METAL software with inverse variance weighting to combine effect estimates after applying genomic control correction.[14]

We sought corroborative evidence for SNPs with  $P < 1 \times 10^{-5}$  in the AGES-Reykjavík cohort study (n = 1,494), and in LHS (n = 4,048), a clinical cohort study of smokers with mild COPD, in which a longitudinal GWAS was recently reported.[15]

## Gene expression analyses

Expression profiles of genes at the novel loci were evaluated in human lung tissues and primary cell samples using RT-PCR (Table S7 in File S1). Using publicly available data from the Lung Genomics Research Consortium (LGRC), expression profiles of these genes were compared in lung specimens of 219 COPD patients and 137 controls, and sentinel (most associated) SNPs at the novel loci were also searched against an eQTL database of lymphoblastoid cell lines.[16]

This manuscript follows the PRISMA statement and a checklist is available online (Checklist S1).

# Results

#### Population characteristics

The majority of the 14 cohort studies had  $\text{FEV}_1$  at two times, but five studies (BHS, CARDIA, CHS, FHS, Health ABC) had  $\geq$ 3 FEV<sub>1</sub> measurements per participant. The maximum length of follow-up ranged from 4 to 29 years. Studies with older participants generally had fewer current smokers and more former smokers, and had lower mean baseline FEV<sub>1</sub>.

# Smoking patterns and rate of decline in FEV<sub>1</sub>

All 14 studies implemented a preliminary mixed model adjusted for all specified variables except the SNP terms and reported the estimated rate of change in FEV<sub>1</sub> by smoking pattern (Table 2). The rate of decline in FEV<sub>1</sub> in never smokers ranged from 10.0 to 39.7 mL/year, and was generally steeper in studies with older participants, as expected.[4] Across all 14 studies, the metaanalyzed rate of change in FEV<sub>1</sub> was a decline of  $26.9\pm0.3$  mL/ year in never smokers, and was  $8.8\pm0.7$ ,  $2.6\pm0.6$ , and  $2.3\pm0.5$  mL/year steeper in persistent, intermittent, and former smokers, respectively (Table 2). We repeated the meta-analyses in the five cohort studies with  $\geq 3$  FEV<sub>1</sub> measurements per participant, and found similar, although less statistically significant results.

#### Discovery meta-analyses

Study-specific genomic inflation factors ( $\lambda_{gc}$ ) were calculated for the SNP-by-time interaction term and used for study-level genomic control prior to the meta-analyses. Study-specific  $\lambda_{gc}$ values ranged from 0.96 to 1.11 (Table S1 in File S1) and the meta-analysis  $\lambda_{gc}$  was 1.01 for both the 14-study and five-study meta-analyses. Figures S1 and S2 in File S1 present the Manhattan and quantile-quantile (QQ) plots.

In the meta-analysis including all 14 cohort studies, 15 SNPs at nine independent loci were associated with the rate of change in FEV<sub>1</sub> at  $P < 1 \times 10^{-5}$ , and none reached the genome-wide significance threshold of  $P < 5 \times 10^{-8}$ . The association results for the sentinel SNPs at these nine loci are presented in Table 3, and more detailed results for all 15 SNPs are included in Table S2 in File S1. The most statistically significant association, and the only one that reached  $P < 1 \times 10^{-6}$ , was for rs4077833, an intronic SNP located in the novel *IL16/STARD5/TMC3* gene region on chromosome 15 ( $P = 5.71 \times 10^{-7}$ ; Figures 1A and 1B). The C allele of rs4077833, with a frequency of 10%, was associated with

FEV<sub>1</sub> decline. doi:10.1371/journal.pone.0100776.t003



B



**Figure 1.** Association of the chromosome 15 locus with the rate of change in FEV<sub>1</sub> in the meta-analysis of 14 cohort studies. A) Regional association plot, where the X-axis is Megabase (Mb) position and Y-axes are the negative log of the *P* value on the left and recombination rate on the right. The sentinel SNP is colored in purple and linkage disequilibrium to the sentinel SNP is depicted by degree of color according to the legend. **B**) Forest plot for rs4077833, where the size of the square for each study represents its contributing weight to the meta-analysis. doi:10.1371/journal.pone.0100776.g001

an attenuation of the rate of decline in  $FEV_1$  by 2.3 mL/year in comparison to the G allele.

For estimation of longitudinal trajectory in lung function, having more than two measurements over time provides greater precision.[4] We performed a further meta-analysis with the five cohort studies (BHS, CHS, CARDIA, FHS, Health ABC) having  $\geq$ 3 FEV<sub>1</sub> measurements per participant, with a combined sample size of 10,476 participants and 32,054 FEV1 measurements (Methods S1 in File S1 for further details). A novel region on chromosome 11 had a genome-wide significant association (P < 5 $\times 10^{-8}$ ) with the rate of change in FEV<sub>1</sub> (Table 4). The most statistically significant finding at this locus was for rs507211, an intronic SNP located in ME3 (Figures 2A and 2B). Six other SNPs, which are in linkage disequilibrium (LD) with rs507211 and are located in *ME3*, were identified at  $P < 1 \times 10^{-6}$  (Table S3 in File S1). The rs507211 A allele, with a frequency of 25%, was associated with an attenuation of the rate of decline in  $FEV_1$  by 2.09 mL/year in comparison to the G allele ( $P = 2.18 \times 10^{-8}$ ). Besides the ME3 locus, 17 SNPs from four other chromosomal regions had P values between 5  $\times$  10<sup>-8</sup> and 1  $\times$  10<sup>-5</sup> for associations with the rate of change in FEV1 (Tables 4 and Table S3 in File S1).

# Additional analyses

Corroborative evidence was sought for the sentinel SNP at each of the 14 loci associated at  $P < 1 \times 10^{-5}$  (from both the 14-study and five-study meta-analyses) in 1,494 adults from the AGES-Reykjavík population-based cohort study (Table S4 in File S1). A *P* value of 0.004, representing the Bonferroni correction for 14 tests at the  $\alpha = 0.05$  level, was selected *a priori* as the threshold for statistical significance. No SNPs achieved this threshold. The lowest *P* value was for rs740577 in *CACNG4* (*P* = 0.08), which showed consistent effect direction and magnitude with the original meta-analysis.

These same 14 SNPs were further examined in LHS, a clinical cohort study of 4,048 smokers with mild COPD for evidence of consistent association between healthy and diseased individuals.[17] None of the 14 SNPs were associated with the rate of change in FEV<sub>1</sub> in LHS at P < 0.004 (Table S4 in File S1).

Previous meta-analyses in the CHARGE and SpiroMeta consortia identified 26 novel loci associated with cross-sectional FEV<sub>1</sub> and/or FEV<sub>1</sub>/FVC at genome-wide significance.[9-11] We examined the sentinel SNPs from these loci in the meta-analysis of the 14 cohort studies for association with the rate of change in FEV<sub>1</sub> (Table S5 in File S1). Given the *a priori* association with cross-sectional lung function, a *P* value threshold of 0.05 was used. Sentinel SNPs in *PID1*, *HHIP*, *GPR126*, and *CFDP1* showed association with the rate of change in FEV<sub>1</sub> (0.005  $\leq P \leq 0.048$ ).

#### Gene expression analyses

Three genes (*IL16*, *STARD5*, and *TMC3*) at the novel chromosome 15 locus and *ME3* at the novel chromosome 11 locus were selected for follow-up mRNA expression profiling in human lung tissue, and primary cultures of human bronchial epithelial and airway smooth muscle cells, together with control tissues (peripheral blood mononuclear cells and brain). Transcripts of *STARD5* and *ME3* were found in all lung-derived tissues, transcripts of *IL16* were found in lung tissue and smooth muscle

cells, but not in epithelial cells, and *TMC3* was not expressed in any of the lung-derived tissues (Table S6 in File S1).

Using the public LGRC data repository, we found that the expression profiles of *IL16*, *STARD5*, and *ME3* in human lung samples showed statistically significant differences (P < 0.05) between COPD patients and controls (Figure S3 in File S1). Lower levels of *IL16* (P = 0.004) were observed in COPD patients compared with controls, whereas higher levels of *STARD5* ( $P = 3.22 \times 10^{-9}$ ) and *ME3* (P = 0.044) were observed in COPD patients compared with controls. Data on *TMC3* expression were not available.

We performed additional follow-up analysis of the sentinel SNPs at the two novel loci using an eQTL database of lymphoblastoid cell lines (Table S8 in File S1). Trans-eQTL associations were observed between rs4077833 at the *IL16/STARD5/TMC3* locus and a nuclear receptor, *NR112* (chromosome 3;  $P = 6.84 \times 10^{-4}$ ) and between rs507211 at the *ME3* locus and *KIAA1109* (chromosome 4;  $P = 5.20 \times 10^{-4}$ ), which is part of a gene cluster (*KIAA1109-TENR-IL2-IL21*) that encodes two interleukins (IL2 and IL21).[18]

# Discussion

Although the genetic contribution to cross-sectional lung function phenotypes has been addressed by large-scale GWAS, much less information is available for longitudinal lung function phenotypes. To identify novel loci that specifically affect lung function change over time, we performed a large-scale GWAS of the rate of change in FEV1 in 27,249 participants from 14 population-based cohort studies. We identified a novel locus (IL16/STARD5/TMC3) on chromosome 15 with suggestive evidence for association with the rate of change in  $FEV_1$ . Given the greater precision to estimate longitudinal trends with more measurements, a meta-analysis of the five cohort studies with  $\geq 3$ FEV1 measurements per participant was performed, and it identified a second novel locus (ME3) on chromosome 11 at genome-wide statistical significance. For both loci, the minor allele was protective, and the magnitude of the association with the rate of change in  $FEV_1$  was similar to that of being an intermittent or former smoker versus a never-smoker.

The sentinel SNP at the novel chromosome 15 locus is located in TMC3, although two neighboring genes, IL16 and STARD5 both harbor SNPs that are in modest LD with the sentinel SNP (Figure 1A). TMC3, a member of the transmembrane channel-like gene family, likely functions as an ion channel, transporter, or modifier,[19] and has been associated with deafness and skin cancer.[20,21] IL16 is a pleiotropic immunomodulatory cytokine that acts as a chemoattractant for CD4<sup>+</sup> cells and contributes to their recruitment and activation in response to inflammation.[22] Notably, asthma was the first disease where increased IL16 expression was observed. [23] Subsequent studies confirmed that in the non-diseased state IL16 is almost exclusively expressed by T lymphocytes in lymphatic tissue, whereas in asthmatic patients IL16 is also synthesized by airway epithelial cells to inhibit airway inflammation.[24-26] A promoter polymorphism (T-295C) in IL16 was associated with asthma in a Caucasian population in England, [27] although this finding was not confirmed in an Australian study.[28] STARD5 belongs to the steroidogenic acute

Table 4. Association   per participant (n =	10,476).	t statistically significant	t SNPs with the rate of change in FEV $_1$ (r	nL/year) in the me	eta-analysis of the five	: cohort stu	dies with	i ≥3 FEV <sub>1</sub> measurements
SNP	Chr	Position	Closest Gene(s)	Coded Allele	Frequency	B*	SE	<i>P</i> Value
rs10209501	2	28536881	FOSL2/PLB1	А	0.33	1.6	0.4	$7.09 \times 10^{-6}$
rs12692550	2	159958017	BAZ2B	Т	0.18	-2.0	0.4	$2.02 \times 10^{-6}$
rs1729588	ſ	110790025	FLJ25363/MIR4445	A	0.30	1.6	0.4	$8.38 \times 10^{-6}$

Data reported are the meta-analysis results of the SNP-by-time interaction term from the GWAS mixed effects model. A positive B-coefficient indicates an attenuation of FEV, decline and a negative B-coefficient an acceleration of single-nucleotide polymorphism. standard error; SNP = chromosome; SE = Definition of abbreviations: Chr

∢

 $4.15 \times 10^{-6}$  $2.18\,\times\,10^{-8}$ 

0.3 0.4

2.1

9.1 <u>د</u>

0.30 0.47 0.25

∢ ⊢

19863644 86054387

10 1

rs10764053

s507211

C10orf112

ME3

FEV, decline.

doi:10.1371/journal.pone.0100776.t004

regulatory lipid transfer domain protein superfamily, and is involved in the trafficking of cholesterol and other lipids between intracellular membranes.[29] Recent in vitro studies showed increased STARD5 expression and protein redistribution as a protective mechanism in response to induced endoplasmic reticulum (ER) stress and consequent over-accumulation of intracellular free cholesterol.[30] We confirmed the expression of STARD5 in all human lung tissues examined and of IL16 in human lung smooth muscle cells, but not epithelial cells, in line with previous observations. In contrast, no expression of TMC3 was detected in any of the tested human lung tissues. We also found significantly lower levels of IL16 in whole lung samples from COPD patients compared with controls, in contrast to its increased expression in asthma, and significantly higher levels of STARD5 in COPD patients compared with controls. Taken together, these results suggest IL16 as the most likely candidate accounting for the observed association, but further investigation is needed to elucidate underlying mechanisms.

The sentinel SNP at the novel chromosome 11 locus is located in ME3, whose protein product is a mitochondrial NADP(+)dependent malic enzyme that catalyzes the oxidative decarboxylation of malate to pyruvate using NADP+ as a cofactor.[31] Mitochondrial malic enzymes play a role in the energy metabolism in tumors, and are considered potential therapeutic targets in cancer.[32,33] We performed independent expression profiling of ME3 and confirmed its expression in all human lung tissues examined, and found significantly higher levels of ME3 in lung samples from COPD patients compared with controls. In addition, we looked up the sentinel SNP in ME3 in a recent GWAS of airway obstruction and found a P value of 0.049.[34] Taken together, these results support ME3 as a biologically plausible candidate in the regulation of lung function and pathogenesis of COPD.

The identification of trans-eQTL associations for the sentinel SNPs at both the IL16/STARD5/TMC3 and ME3 loci is interesting, and while the interpretation of trans-eQTL associations is ambiguous, [35] the regions these SNPs regulate merit further study.

Besides the GWAS meta-analyses, the assembly of 14 longitudinal cohort studies allowed us to meta-analyze the association of cumulative smoking patterns with the rate of change in  $FEV_1$  in the general population. The meta-analyzed estimate for the rate of decline in FEV1 in never smokers was 26.9 mL/year, and the annual decline was steeper in persistent, intermittent, and former smokers by 8.8, 2.6, and 2.3 mL/year, respectively. These findings provide a reference point for the effect of cigarette smoking on longitudinal lung function change in the general population.

There is phenotypic variation among the 14 cohort studies in aspects such as baseline age and cigarette smoking, and in factors that are of special importance to this longitudinal GWAS, such as the number of FEV1 measurements per participant and follow-up duration. Phenotypic heterogeneity represents a general challenge in genetic epidemiology, particularly in the investigation of longitudinal phenotypes. Thus, we performed a meta-analysis using the subset of cohort studies with  $\geq 3$  FEV<sub>1</sub> measurements per participant, given that longitudinal trajectories are best estimated over longer time periods and with more measurements. There was little overlap between the top loci identified in the two metaanalyses at  $P < 1 \times 10^{-5}$ , suggesting that phenotypic heterogeneity affected the association results. Future meta-studies of lung function decline should aim to increase sample size while maintaining high phenotypic comparability among participating studies. In addition, the trajectory of lung function change, especially over a long period of time, is known to be nonlinear,



B



Figure 2. Association of the chromosome 11 locus with the rate of change in FEV<sub>1</sub> in the meta-analysis of the five cohort studies with  $\geq$ 3 FEV<sub>1</sub> measurements per participant. A) Regional association plot, where the X-axis is Megabase (Mb) position, and the Y-axes are the negative log of the *P* value on the left and recombination rate on the right. The sentinel SNP is colored in purple and linkage disequilibrium to the sentinel SNP is depicted by degree of color according to the legend. B) Forest plot for rs507211, where the size of the square for each study represents its contributing weight to the meta-analysis.

doi:10.1371/journal.pone.0100776.g002

which may require the use of nonlinear time effects in the statistical model. In this study, given that over half of the included cohort studies have  $FEV_1$  measurements at only two time points, our consideration was limited to a linear time effect. Further, the outcome studied, the rate of change in lung function, represents one of many ways to describe lung function change. Additional studies of other aspects of lung function change, such as reduced growth and premature decline, would be of interest.

We sought corroborative evidence in a single cohort study of 1,494 participants. This sample size is much smaller and arguably insufficient compared with replications applied to previous studies of cross-sectional lung function phenotypes. Thus, despite the lack of corroboration for the two novel loci identified in the metaanalyses, results from the complementary gene expression analyses provide compelling evidence for biologically plausible roles of the implicated genes in the longitudinal change in lung function.

None of the 14 sentinel SNPs were associated with the rate of change in  $FEV_1$  in the COPD patient-based LHS cohort. Similarly, a previous population-based GWAS of lung function decline noted a high degree of heterogeneity in findings when analyses were stratified by presence/absence of asthma.[12] The observed discrepancy of association results suggests that the genetic determination of lung function decline may be different in healthy individuals compared with COPD patients, may contribute differentially in a pre-diseased vs. post-diseased state in which medications may influence the rates of decline, or that LHS was underpowered for confirming our findings.

In this study, statistical models included a comprehensive list of confounders that are commonly adjusted for when modeling lung function phenotypes. Given the study's meta-analysis design and the objective to carry out the same statistical model in all cohort studies, additional covariates that were not available in all cohort studies could not be included. In addition, the adjustment of certain confounders, such as smoking, is challenging in a longitudinal study, and although we accounted for the two most important aspects of smoking, cumulative pattern and dosage, residual confounding due to smoking cannot be excluded.

In summary, we performed GWAS of the longitudinal change in lung function and subsequent meta-analyses, using harmonized data from more than 27,000 participants of European ancestry to identify genetic loci influencing the rate of change in FEV<sub>1</sub>. We identified the novel *ME3* locus on chromosome 11 at genome-wide significance and found suggestive evidence for association at the novel *IL16/STARD5/TMC3* locus on chromosome 15. Additional expression analyses confirmed the expression of *ME3*, *IL16*, and *STARD5* in multiple lung tissues, and found differential expression profiles of these three genes in the lungs of COPD patients compared to non-COPD controls. These results support the involvement of these implicated genes in the longitudinal change in lung function in adults of European ancestry. Additional studies with larger sample size and in populations of other races/ ethnicities are warranted.

# Supporting Information

File S1 This is a single file that contains all supporting information for the paper. Briefly, File S1 contains the following items: Methods S1, which describes further details of the cohort studies and the statistical methodology; Table S1, Details of SNP genotyping, quality control (QC), imputation, and statistical analysis across the 14 cohort studies; Table S2, Regression results for single nucleotide polymorphisms associated with the rate of change in FEV<sub>1</sub> (mL/year) at  $\dot{P} < 1 \times 10^{-5}$  in the meta-analysis of 14 cohort studies (N = 27,249); Table S3, Regression results for single nucleotide polymorphisms associated with the rate of change in FEV<sub>1</sub> (mL/year) at  $P < 1 \times 10^{-5}$  in the meta-analysis of the five cohort studies with three or more FEV1 measurements per participant (N = 10,476); Table S4, Association of the 14 sentinel SNPs from the meta-analyses in the AGES-Reykjavík study (AGES) and the Lung Health Study (LHS) for the rate of change in FEV<sub>1</sub> (mL/year); Table S5, Association of previously reported loci in GWAS of cross-sectional lung function with the rate of change in FEV<sub>1</sub> (mL/year) in the meta-analysis of 14 cohort studies (N = 27,249); Table S6, mRNA expression profiling of the implicated genes at the two novel loci in human lung and control tissues; Table S7, Primers for mRNA expression profiling; Table S8, Summary of eOTL look-up for the most significant SNPs at the novel chromosome 11 and 15 loci; Figure S1, Manhattan and QQ plots for the meta-analysis of the rate of change in FEV<sub>1</sub> in 14 cohort studies; Figure S2, Manhattan and QQ plots for the meta-analysis of the rate of change in  $FEV_1$  in the five cohort studies with three or more FEV1 measurements per participant; Figure S3, mRNA expression profiling in human lung samples from 219 COPD patients and 137 controls for A) IL16, B) STARD5, and C) ME3, using publicly available microarray data from the Lung Genomics Research Consortium site (http://www. lung-genomics.org/). The y-axes reflect the probe intensities of each gene transcript in the binary logarithm form, with the red dots indicating the average probe intensities and the red bars indicating standard deviation. The P values were calculated using the two-sample t-test.

(DOCX)

# Checklist S1 PRISMA Checklist. (DOCX)

# **Author Contributions**

Wrote the paper: All authors. Drafted the manuscript: WT PAC. AGES Study concept, design: TBH VG LJL. ARIC Study concept, design: DJC NF DBH BRJ SJL ACM KEN. B58C Study concept and design: DPS. BHS Study concept, design: AJ B. Musk. CARDIA Study concept, design: M. Fornage AS. CHS Study concept, design: SRH SAG BMP. FHS Study concept, design: JD GTO JBW. Health ABC Study concept, design: PAC SBK WT. KORA Study concept, design: JH HS. LBC Study concept, design: IJD JMS. LHS Study concept, design: KCB NNH RAM. RS Study concept, design: GGB AH FR BHS AGU. SAPALDIA Study concept, design: MI NMP-H. SHIP Study concept, design: BK SG HV. SpiroMeta Study concept, design: IPH MDT. AGES Genotype data/QC: AVS. ARIC Phenotype data/QC: DJC. Genotype data/QC: ACM KEN. B58C Phenotype data/QC: DPS. Genotype data/QC: WLM. BHS Phenotype data/QC: AJ B. Musk. Genotype data/QC: AJ B. Musk LJP. CARDIA Phenotype data/QC: LJS AS ODW. Genotype data/QC: M. Fornage M. Foy XG. CHS Phenotype data/QC: BMP. Genotype data/QC: TL BMP JIR. FHS Phenotype data/QC: GTO. Genotype data/QC: GTO. Health ABC Phenotype data/QC: PAC SBK B. Mwibohm WT. Genotype data/QC: SBK YL KL. KORA Phenotype data/QC: JK SK HS. LBC Phenotype data/QC: IJD JMS. Genotype data/QC: GD. LHS Genotype

data/QC: KCB RAM IR. RS Phenotype data/QC: GGB L. Lahousse DWL BHS. Genotype data/QC:FR AGU. SAPALDIA Phenotype data/ QC: IC MI NMP-H. Genotype data/QC: IC AK MI NMP-H. SHIP Phenotype data/QC: BK SG HV. Genotype data/QC: BK SG AT HV. SpiroMeta Genotype data/QC: IPH MDT. PIVUS Phenotype data/QC: EI L. Lind. Genotype data/QC: EI L. Lind APM. AGES Data analysis: AVS. ARIC Data analysis: BRJ SJL. B58C Data analysis: DPS LVW. BHS

## References

- Rabe KF, Hurd S, Anzueto A, Barnes PJ, Buist SA, et al. (2007) Global strategy for the diagnosis, management, and prevention of chronic obstructive pulmonary disease: GOLD executive summary. Am J Respir Crit Care Med 176: 532–555.
- Schunemann HJ, Dorn J, Grant BJ, Winkelstein W Jr, Trevisan M (2000) Pulmonary function is a long-term predictor of mortality in the general population: 29-year follow-up of the Buffalo Health Study. Chest 118: 656–664.
- Young RP, Hopkins R, Eaton TE (2007) Forced expiratory volume in one second: not just a lung function test but a marker of premature death from all causes. Eur Respir I 30: 616-622.
- Fletcher C, Peto R (1977) The natural history of chronic airflow obstruction. Br Med J 1: 1645–1648.
- Halbert RJ, Natoli JL, Gano A, Badamgarav E, Buist AS, et al. (2006) Global burden of COPD: systematic review and meta-analysis. Eur Respir J 28: 523– 532.
- Eisner MD, Anthonisen N, Coultas D, Kuenzli N, Perez-Padilla R, et al. (2010) An official American Thoracic Society public policy statement: Novel risk factors and the global burden of chronic obstructive pulmonary disease. Am J Respir Crit Care Med 182: 693–718.
- Gottlieb DJ, Wilk JB, Harmon M, Evans JC, Joost O, et al. (2001) Heritability of longitudinal change in lung function. The Framingham study. Am J Respir Crit Care Med 164: 1655–1659.
- Finkel D, Pedersen NL, Reynolds CA, Berg S, de Faire U, et al. (2003) Genetic and environmental influences on decline in biobehavioral markers of aging. Behav Genet 33: 107–123.
- Hancock DB, Eijgelsheim M, Wilk JB, Gharib SA, Loehr LR, et al. (2009) Metaanalyses of genome-wide association studies identify multiple loci associated with pulmonary function. Nat Genet 42: 45–52.
- Repapi E, Sayers I, Wain LV, Burton PR, Johnson T, et al. (2009) Genome-wide association study identifies five loci associated with lung function. Nat Genet 42: 36–44.
- Soler Artigas M, Loth DW, Wain LV, Gharib SA, Obeidat M, et al. (2011) Genome-wide association and large-scale follow up identifies 16 new loci influencing lung function. Nat Genet 43: 1082–1090.
- Imboden M, Bouzigon E, Curjuric I, Ramasamy A, Kumar A, et al. (2012) Genome-wide association study of lung function decline in adults with and without asthma. The Journal of allergy and clinical immunology 129: 1218– 1228.
- Miller MR, Hankinson J, Brusasco V, Burgos F, Casaburi R, et al. (2005) Standardisation of spirometry. Eur Respir J 26: 319–338.
- Willer CJ, Li Y, Abecasis GR (2010) METAL: fast and efficient meta-analysis of genomewide association scans. Bioinformatics 26: 2190–2191.
- Hansel NN, Ruczinski I, Rafaels N, Sin DD, Daley D, et al. (2013) Genomewide study identifies two loci associated with lung function decline in mild to moderate COPD. Hum Genet 132: 79–90.
- Dixon AL, Liang L, Moffatt MF, Chen W, Heath S, et al. (2007) A genome-wide association study of global gene expression. Nat Genet 39: 1202–1207.
- Anthonisen NR, Connett JE, Kiley JP, Altose MD, Bailey WC, et al. (1994) Effects of smoking intervention and the use of an inhaled anticholinergic bronchodilator on the rate of decline of FEV1. The Lung Health Study. JAMA 272: 1497–1505.
- van Heel DA, Franke L, Hunt KA, Gwilliam R, Zhernakova A, et al. (2007) A genome-wide association study for celiac disease identifies risk variants in the region harboring IL2 and IL21. Nat Genet 39: 827–829.

Data analysis: MK LP. CARDIA Data analysis: M. Fornage M. Foy XG. CHS Data analysis: SAG SRH GL TL AV. FHS Data analysis: JD WG JBW. Health ABC Data analysis: PAC YL KL WT MTW. KORA Data analysis: EA. LBC Data analysis: MA GD. LHS Data analysis: KCB NNH RAM IR. RS Data analysis: L. Lahousse DWL. SAPALDIA Data analysis: MI. SHIP Data analysis: AT. SpiroMeta Data analysis: MSA IPH MDT. PIVUS Data analysis: TF.

- Kurima K, Yang Y, Sorber K, Griffith AJ (2003) Characterization of the transmembrane channel-like (TMC) gene family: functional clues from hearing loss and epidermodysplasia vertuciformis. Genomics 82: 300–308.
- Ramoz N, Rueda LA, Bouadjar B, Montoya LS, Orth G, et al. (2002) Mutations in two adjacent novel genes are associated with epidermodysplasia verruciformis. Nat Genet 32: 579–581.
- Vreugde S, Erven A, Kros CJ, Marcotti W, Fuchs H, et al. (2002) Beethoven, a mouse model for dominant, progressive hearing loss DFNA36. Nat Genet 30: 257–258.
- Cruikshank WW, Kornfeld H, Center DM (1998) Signaling and functional properties of interleukin-16. International reviews of immunology 16: 523–540.
- Bellini A, Yoshimura H, Vittori E, Marini M, Mattoli S (1993) Bronchial epithelial cells of patients with asthma release chemoattractant factors for T lymphocytes. The Journal of allergy and clinical immunology 92: 412–424.
- Cruikshank WW, Long A, Tarpy RE, Kornfeld H, Carroll MP, et al. (1995) Early identification of interleukin-16 (lymphocyte chemoattractant factor) and macrophage inflammatory protein 1 alpha (MIP1 alpha) in bronchoalveolar lavage fluid of antigen-challenged asthmatics. Am J Respir Cell Mol Biol 13: 738–747.
- Krug N, Cruikshank WW, Tschernig T, Erpenbeck VJ, Balke K, et al. (2000) Interleukin 16 and T-cell chemoattractant activity in bronchoalveolar lavage 24 hours after allergen challenge in asthma. Am J Respir Crit Care Med 162: 105– 111.
- Laberge S, Ernst P, Ghaffar O, Cruikshank WW, Kornfeld H, et al. (1997) Increased expression of interleukin-16 in bronchial mucosa of subjects with atopic asthma. Am J Respir Cell Mol Biol 17: 193–202.
- Burkart KM, Barton SJ, Holloway JW, Yang IA, Cakebread JA, et al. (2006) Association of asthma with a functional promoter polymorphism in the IL16 gene. The Journal of allergy and clinical immunology 117: 86–91.
- Åkesson LS, Duffy DL, Phelps SC, Thompson PJ, Kedda MA (2005) A polymorphism in the promoter region of the human interleukin-16 gene is not associated with asthma or atopy in an Australian population. Clinical and experimental allergy: journal of the British Society for Allergy and Clinical Immunology 35: 327–331.
- Rodriguez-Agudo D, Ren S, Hylemon PB, Redford K, Natarajan R, et al. (2005) Human StarD5, a cytosolic StAR-related lipid binding protein. Journal of lipid research 46: 1615–1623.
- Rodriguez-Agudo D, Calderon-Dominguez M, Medina MA, Ren S, Gil G, et al. (2012) ER stress increases StarD5 expression by stabilizing its mRNA and leads to relocalization of its protein from the nucleus to the membranes. Journal of lipid research 53: 2708–2715.
- Chang GG, Tong L (2003) Structure and function of malic enzymes, a new class of oxidative decarboxylases. Biochemistry 42: 12721–12733.
- Moreadith RW, Lehninger AL (1984) The pathways of glutamate and glutamine oxidation by tumor cell mitochondria. Role of mitochondrial NAD(P)+dependent malic enzyme. J Biol Chem 259: 6215–6221.
- Teller JK, Fahien LA, Davis JW (1992) Kinetics and regulation of hepatoma mitochondrial NAD(P) malic enzyme. J Biol Chem 267: 10423–10432.
- Wilk JB, Shrine NR, Loehr LR, Zhao JH, Manichaikul A, et al. (2012) Genome Wide Association Studies Identify CHRNA5/3 and HTR4 in the Development of Airflow Obstruction. Am J Respir Crit Care Med.
- Montgomery SB, Dermitzakis ET (2011) From expression QTLs to personalized transcriptomics. Nat Rev Genet 12: 277–282.