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Journal

Communications Biology, 5(1)

ISSN

2399-3642

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Publication Date


















2022

DOI

10.1038/s42003-022-04267-y

Peer reviewed

A meta-analysis of pre-pregnancy maternal body mass index and placental DNA methylation identifies 27 CpG sites with implications for mother-child health

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Higher maternal pre-pregnancy body mass index (ppBMI) is associated with increased neonatal morbidity, as well as with pregnancy complications and metabolic outcomes in offspring later in life. The placenta is a key organ in fetal development and has been proposed to act as a mediator between the mother and different health outcomes in children. The overall aim of the present work is to investigate the association of ppBMI with epigenome-wide placental DNA methylation (DNAm) in 10 studies from the PACE consortium, amounting to 2631 mother-child pairs. We identify 27 CpG sites at which we observe placental DNAm variations of up to 2.0% per 10 ppBMI-unit. The CpGs that are differentially methylated in placenta do not overlap with CpGs identified in previous studies in cord blood DNAm related to ppBMI. Many of the identified CpGs are located in open sea regions, are often close to obesity-related genes such as *GPX1* and *LGR4* and altogether, are enriched in cancer and oxidative stress pathways. Our findings suggest that placental DNAm could be one of the mechanisms by which maternal obesity is associated with metabolic health outcomes in newborns and children, although further studies will be needed in order to corroborate these findings.

Higher maternal pre-pregnancy body mass index (ppBMI) is associated with aberrant fetal growth¹, macrosomia and increased neonatal morbidity and mortality², and also with pregnancy complications such as pre-eclampsia, gestational diabetes, gestational hypertension, pre-term delivery and cesarean section³. It has been shown that maternal adipokine and insulin signaling in the placenta could contribute to regulate both the vascular development of this organ and the nutrient transport to the fetus, and therefore impact fetal development³. Additionally, it has also been observed that maternal ppBMI is associated with other offspring health outcomes in later life, including increased risk for obesity in children⁴. Observational studies have suggested links between maternal obesity and long-term risk of coronary heart disease, stroke, type 2 diabetes and asthma in offspring⁵. Very high maternal ppBMI has also been associated with poorer cognitive performance in children and greater risk of neurodevelopmental disorders⁶, while there is also preliminary evidence in favor of potential implications in immune and infectious disease-related outcomes³. These associations could be mediated by epigenetic changes, including DNA methylation (DNAm), but the implication of an early epigenetic reprogramming *in utero* deserves further research⁷.

A previous study carried out within the Pregnancy and Childhood Epigenetics (PACE) consortium⁸ has shown that maternal ppBMI is widely associated with differences in cord blood DNAm in the newborn⁹. However, the authors observed that many of the significant epigenetic effects were modest (<0.2% methylation per BMI unit) and they did not detect enrichment for any particular biological pathway, leaving open questions regarding potential intra-uterine mechanisms that could be affecting the epigenetic profile of the newborn⁹. In this context, while the epigenetic alterations in cord and peripheral blood have been thoroughly investigated^{9,10}, the potential impact of maternal ppBMI in placental DNAm remains poorly explored. As far as we know, the most recent studies have performed methylation profiling with methylation arrays or reduced representation bisulfite sequencing in up to 300 term placentas of obese and non-obese mothers^{11,12}. Although interesting, these studies have yielded a limited number of significant results, probably because of their relatively small sample size.

A recently published meta-analysis with 1700 placental samples by the PACE consortium¹³ has discovered a placental DNAm signature of maternal smoking during pregnancy that is quite different from what has been observed in cord blood¹⁴. Differentially methylated CpGs related to smoking in pregnancy fall within active regions of the placental epigenome, and nearby genes are involved in the response to environmental stressors, regulation of inflammatory activity, and growth factor signaling. The placenta is a transient organ at the maternal-fetal interface, with endocrine and substrate-transport functions, that is sensitive to pregnancy environmental influences – exogenous or endogenous. Maternal pre-pregnancy obesity is often characterized by an adverse metabolic milieu that may alter placental function by increasing oxidative stress, vascular endothelium thickening, and inflammatory lesions in placental tissues¹¹. Altogether, these facts encourage the investigation of the placenta as a putative mediator of maternal obesity and health outcomes in the offspring, specifically through the modification of the placental DNAm landscape.

In this context, the overall aim of the current analyses was to investigate the association of maternal ppBMI with epigenome-wide placental DNAm in 10 studies from the PACE consortium amounting to 2631 mother-child pairs. We also conducted functional enrichment analyses and comparison of our results with maternal ppBMI-associated cord blood DNAm alterations previously reported by PACE.

Table 1 Effective sample size of the participating cohorts.

Cohort	Country	N	Array
AQUA	Australia	95	450 K
EARLI	USA	54	450 K
EDEN	France	664	450 K
HEBC	USA	186	450 K
Gen3G	Canada	448	EPIC
GENEIDA	Spain	103	450 K
INMA	Spain	168	450 K
ITU	Finland	352	EPIC
NHBCS	USA	311	450 K
RICHs	USA	250	450 K
TOTAL		2631	

Results

Study population. Eleven North-American, Australian, and European studies ($N = 2631$) contributed to the epigenome-wide association study (EWAS) to determine the associations of maternal ppBMI with placental DNAm (Table 1), including Asking Questions about Alcohol in pregnancy (AQUA)¹⁵, Early Autism Risk Longitudinal Investigation (EARLI)¹⁶, Study on the pre- and early postnatal determinants of child health and development (EDEN)¹⁷, Genetics of Glucose regulation in Gestation and Growth (Gen3G)¹⁸, Genetics, Early Life Environmental Exposures and Infant Development in Andalusia (GENEIDA)¹⁹, Harvard Epigenetic Birth cohort (HEBC)²⁰, Environment and Childhood Project (INMA)²¹, The Intrauterine Sampling in Early Pregnancy Study (ITU)²², Markers of Autism Risk in Babies-Learning Early Signs (MARBLES)²³, New Hampshire Birth Cohort Study (NHBCS)²⁴, and Rhode Island Child Health Study (RICHs)²⁵. MARBLES was excluded at a later stage as the sample size was too small and results were inconsistent with the other cohorts (Supplementary Fig. 1).

Maternal ppBMI (kg/m^2) was generally self-reported. In those cases where ppBMI was not available, BMI in early pregnancy (1st trimester) was used. For simplicity, we will refer to both of them as maternal ppBMI. In all the analyses performed, we used ppBMI as a continuous variable. The cohort-specific average maternal ppBMI ranged from 22.9 in EDEN (France) to 27.6 in EARLI (USA) (standard deviation-SD = 1.60). In general, 548 (20.8%) and 369 (14.0%) mothers reported overweight (>25 BMI) and obesity (>30 BMI), respectively, while 115 (4.37%) appeared to be underweight (<18.5 BMI). Mean age of the mothers was 30.7 years (SD = 2.9). The distributions of other covariates by cohort are provided in Supplementary Data 1.

Genome-wide DNAm meta-analyses. Each cohort analyst conducted two different EWAS, modeling DNAm beta-values at a maximum number of 419,460 CpG sites in relation to maternal ppBMI using robust linear regressions, with and without adjustment for putative cellular components. Cell composition was estimated using the reference-free deconvolution algorithm RefFreeCellMix²⁶. The CpGs included in the analyses were among those shared by the two most common Illumina Infinium Beadchip arrays, 450 K and EPIC, since both arrays were used to assess the DNAm levels of the samples, as shown in Table 1. All models were adjusted for maternal age, parity, maternal education and maternal smoking. Genomic inflation factors from the cohort-specific models (ranging from $\lambda = 0.692$ to 1.472) and a summary of the results can be seen in Supplementary Data 2. Finally, after quality control of the results, we conducted an inverse variance-weighted fixed-effects meta-analysis using the software GWAMA²⁷. The inflation factors from the meta-analyses ($\lambda = 1.230$ and 1.220 for the cell type-adjusted and

-unadjusted models, respectively) (Fig. 1) revealed potential residual confounding and moderate inflation of test statistics.

After applying the Bonferroni correction for multiple-testing (meta-analysis nominal p value $<1.2e-07$), we obtained 27 and 42 CpGs at which maternal ppBMI was significantly associated with placental DNAm in the models adjusted and unadjusted for cell type proportions, respectively. Full results for both models are provided in Supplementary Data 3 and 4, respectively. Higher maternal ppBMI was associated with lower placental DNAm in 24/27 differentially methylated CpGs identified in the cell type-adjusted model, while in the unadjusted model, 33/42 hits showed positive associations (higher maternal ppBMI associated with increased placental DNAm at the identified CpGs). However, beta-coefficients of CpGs that were differentially methylated in one model were positively correlated to the beta-coefficients of the same position in the other (Supplementary Fig. 2). Finally, the heterogeneity of associations across cohorts was lower for the model adjusted for cell type proportions compared to the unadjusted model (26/27 vs. 34/42 CpGs presented Cochran's Q-test p -values > 0.01) and thus, we continued with the results from the fully adjusted model for all downstream analyses.

Among the 27 differentially methylated CpGs identified in our cell type-adjusted EWAS (Table 2, Fig. 2A, B), a few individual CpGs are worthy of mention. The most notable association was observed at cg08219219, located in the eighth exon of *EPHX3*, with the lowest p -value in the meta-analysis (Bonferroni-corrected, meta-analysis p -value = $2.12e-05$) and a beta-coefficient of $-1.12e-03$, meaning that a 10-unit difference in maternal ppBMI is associated with a 1.1% lower DNAm at this specific CpG site. This association was consistent across all cohorts (Cochran's Q-test p -value = 0.12) (Fig. 3a). The largest beta-coefficient was observed in cg14704941, in the first intron of *CSRP3*, with a positive beta-coefficient of $1.96e-03$, corresponding to a 2% higher placental DNAm per 10-unit ppBMI (Bonferroni-corrected p -value = $2.24e-04$ and Cochran's Q-test p -value = 0.09) (Fig. 3b). In turn, the largest negative beta-coefficient was found in cg04724807 (more than 57 Kb upstream of *SYT16*) with 1.8% lower DNAm per 10-unit ppBMI (Bonferroni-corrected p -value = $3.83e-04$ and Cochran's Q-test p -value = 0.097) (Fig. 3c). The following CpGs reached the Bonferroni significance threshold and were not identified as highly heterogeneous across cohorts: cg00423969 and cg14163484, 1.5 kb upstream of the *FER1L5* promoter, as well as cg26433445, cg15933729 and

cg08539067, close to *CMIP*, *LGR4* and *GPX1*, respectively (Fig. 3d–h). The remaining Bonferroni-significant hits and their heterogeneity across cohorts are shown in Supplementary Fig. 3.

Gene-set and regulatory enrichment analyses. To gain insight into the biological processes that may be captured by placental DNAm associated with maternal ppBMI, we performed gene-set and regulatory enrichment analyses. To this end, first, we annotated CpGs to genes and regulatory elements as explained in the Material and Methods section. Then, we conducted gene-set enrichments for the 26 unique genes annotated to the 27 maternal ppBMI-sensitive CpGs with ConsensusPathDB²⁸ using KEGG, Reactome, Wikipathways and Biocarta reference databases. Two gene-set pathways were significantly enriched (q value < 0.05), namely small cell lung cancer and oxidative stress-induced signaling pathway (Supplementary Data 5). This was also true when we reduced the background from default to only the genes that are represented in the Illumina 450 K array ($\sim 21,231$). We also tested whether the genes annotated to maternal ppBMI-associated CpGs were enriched for regulatory regions of specific transcription factors (TFs). Most notably, our ppBMI-associated CpGs were enriched for genes regulated by *ZNF217* (adjusted p value = 0.02).

We then examined whether the 27 maternal ppBMI-associated CpGs were enriched for CpG island locations, placenta-specific imprinting regions or parent-of-origin-specific germline differentially methylated regions²⁹, regulatory features from the placenta-specific 15-chromatin state annotations from ROADMAP^{30,31}, or placenta-specific partially methylated domains³² that contain placenta-specific repressed genes. We did not find any significant enrichment except for CpG island location and features: the maternal ppBMI-associated CpGs were depleted in CpG islands ($\chi^2 = -2.927$, p value = $8.4e-04$) and highly enriched in open sea regions ($\chi^2 = 2.742$, p value = $1.3e-03$) (Supplementary Fig. 4).

Proximity to genetic variants relevant for birth outcomes. We wanted to determine whether the maternal ppBMI-associated CpGs that we identified here were localized near genetic variants that have been associated with birth outcomes in previously published genome-wide association studies (GWAS). Thus, we investigated whether ppBMI-associated CpGs were within ± 0.5 Mb (1 Mb window) of single nucleotide polymorphisms (SNPs) that have been associated with birth weight (BW,

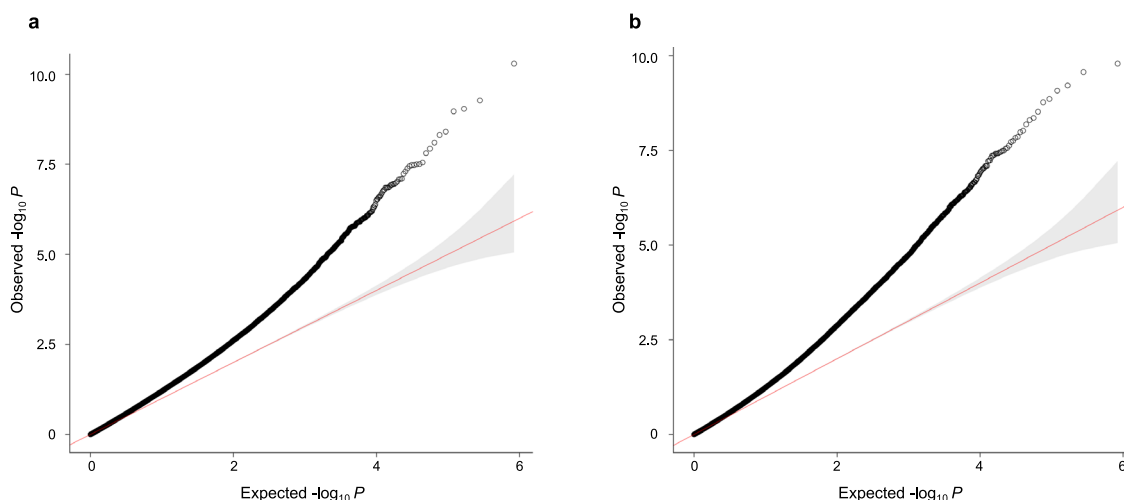


Fig. 1 QQ-plots of the meta-analyses of the association between maternal ppBMI and placental DNAm. Analyses were carried out **a** adjusting for putative cellular heterogeneity or **b** without adjusting for putative cellular heterogeneity ($N = 2631$ placental DNA samples). The red line represents the normal distribution and 95% confidence interval (gray shading).

Table 2 Meta-analysis results from the model adjusted for maternal age, parity, maternal education, maternal smoking and putative cellular heterogeneity, showing the 27 Bonferroni significant CpGs associated with maternal ppBMI.

CpG	Results from the meta-analysis				Meta-analysis heterogeneity				Location Annotations			Distance to closest Gene	Relation to Island	
	Beta	SE	95% CI	P-value	Adj. P-value	Direction	HetISq	HetDf	HetPVal	Chr	Pos			Closest Gene
cg00423969	-0.00092	0.00015	-0.0012;-0.0006	1.07E-09	4.49E-04	-----+-----	0.474	10	0.047	2	97359879	FERIL5	0	Open Sea
cg00510149	-0.00134	0.00025	-0.0018;-0.0008	9.56E-08	4.01E-02	--+-----	0.531	10	0.023	11	10674966	IRAG1	0	Open Sea
cg02286857	-0.00134	0.00025	-0.0018;-0.0008	1.11E-07	4.66E-02	-----+-----	0.602	9	0.010	2	47297177	TTC7A	0	Open Sea
cg03603866	0.00129	0.00023	0.0008;0.0017	1.16E-08	4.87E-03	+++++?+	0.167	9	0.294	13	107027625	LINC00460	1284	Open Sea
cg04724807	-0.00179	0.00029	-0.0024;-0.0012	9.12E-10	3.83E-04	++-----	0.391	10	0.097	14	62396305	SYT16	57496	Open Sea
cg05590755	-0.00080	0.00015	-0.0011;-0.0005	1.19E-07	4.99E-02	--+-----	0.439	10	0.066	1	23855149	E2F2	0	North Shore
cg05965490	-0.00132	0.00025	-0.0018;-0.0008	1.16E-07	4.87E-02	---+-----	0.398	10	0.092	8	30264627	RBPMS	0	Open Sea
cg08129759	-0.00077	0.00014	-0.0010;-0.0005	8.09E-08	3.39E-02	-----+-----	0.303	10	0.166	1	202091944	GPR37L1	83	Open Sea
cg08219219	-0.00112	0.00017	-0.0014;-0.0008	5.05E-11	2.12E-05	-----+-----	0.361	10	0.119	19	15337971	EPHX3	0	South Shelf
cg08539067	-0.00049	0.00009	-0.0007;-0.0003	2.84E-08	1.19E-02	--+-----+-----	0.491	10	0.039	3	49395985	GPX1	193	South Shore
cg09126859	-0.00106	0.00019	-0.0014;-0.0007	3.17E-08	1.33E-02	---+-----	0.415	10	0.081	12	52244063	FIGL2	18361	South Shore
cg09167414	-0.00152	0.00028	-0.0021;-0.0010	3.31E-08	1.39E-02	-----+-----	0.000	10	0.511	1	16076206	TMEM82	1913	South Shelf
cg12613632	-0.00088	0.00016	-0.0012;-0.0006	3.33E-08	1.40E-02	-----+-----	0.165	10	0.292	1	95385935	CNN3	0	Open Sea
cg14051770	-0.00162	0.00029	-0.0022;-0.0010	3.58E-08	1.50E-02	-----+-----	0.000	10	0.833	7	76054572	ZP3	0	Open Sea
cg14143441	-0.00174	0.00031	-0.0023;-0.0011	1.55E-08	6.50E-03	---+-----	0.513	10	0.030	8	134387493	NDRG1	77945	Open Sea
cg14163484	-0.00085	0.00015	-0.0012;-0.0005	4.13E-08	1.73E-02	-----+-----	0.224	8	0.251	2	97359926	FERIL5	0	Open Sea
cg14244402	-0.00170	0.00032	-0.0023;-0.0011	1.10E-07	4.61E-02	-----+-----	0.122	10	0.330	9	130681102	ST6GALNAC4	1796	South Shore
cg1704941	0.00196	0.00032	0.0013;0.0026	5.33E-10	2.24E-04	+++++?+	0.432	8	0.090	11	19224659	CSRP3	0	Open Sea
cg15258080	-0.00129	0.00022	-0.0017;-0.0008	7.91E-09	3.32E-03	-----+-----	0.419	10	0.078	10	71091204	HK1	0	Open Sea
cg15933729	-0.00079	0.00014	-0.0011;-0.0005	4.84E-09	2.03E-03	---+-----	0.182	10	0.276	11	27504612	LGR4	10277	Open Sea
cg16310415	0.00172	0.00032	0.0011;0.0023	4.99E-08	2.09E-02	+++++?+	0.550	9	0.023	8	25898539	PPP2R2A	0	North Shore
cg16724070	-0.00163	0.00030	-0.0022;-0.0010	8.22E-08	3.45E-02	---+-----+-----	0.605	9	0.009	1	183133736	LAMC1	19008	Open Sea

Table 2 (continued)

CpG	Results from the meta-analysis				Meta-analysis heterogeneity				Location Annotations					
	Beta	SE	95% CI	Adj. P-value	P-value	Direction	HetISq	HetDf	HetPVal	Chr	Pos	Closest Gene	Distance to closest Gene	Relation to Island
cg20042798	-0.00052	0.00010	-0.0007; -0.0003	7.95E-08	3.33E-02	-----	0.000	10	0.827	3	13036713	<i>IQSEC1</i>	0	South Shore
cg22673972	-0.00085	0.00015	-0.0011; -0.0005	3.12E-08	1.31E-02	-----	0.000	10	0.949	3	14415346	<i>SLC6A6</i>	28728	Open Sea
cg23696550	-0.00131	0.00024	-0.0018; -0.0008	5.80E-08	2.43E-02	-----+	0.136	10	0.318	14	24732386	<i>TGM1</i>	0	Open Sea
cg24893073	-0.00061	0.00012	-0.0008; -0.0004	1.04E-07	4.36E-02	-----	0.000	10	0.845	17	7742126	<i>KDM6B</i>	1107	N_Shelf
cg26433445	-0.00086	0.00015	-0.0011; -0.0006	3.92E-09	1.64E-03	-----	0.571	10	0.013	16	81764289	<i>CMIP</i>	18921	Open Sea

Beta: effect size; SE: Standard error; 95%CI: 95% confidence interval; Adjusted P-value: Bonferroni corrected p-value; HetISq: Heterogeneity test statistic¹²; HetDf: Heterogeneity degrees of freedom; HetPVal: Heterogeneity test p-value; Direction: direction of the effect in each cohort, ordered in alphabetical order (-: positive; +: negative; and ?; CpG not present in the cohort); CpGs that were not annotated with a gene name in the Illumina 450 K annotation file have been annotated with their closest gene.

$N = 310$), birth length ($N = 5$), head circumference ($N = 3$), gestational age (GA, $N = 6$) and BW + GA ($N = 6$)^{33–38}. Of the total 330 birth outcome SNPs in autosomal chromosomes, 10 BW-associated SNPs were within 0.5 Mb of CpGs that were associated with maternal ppBMI. Therefore, more than a third of the 27 ppBMI-associated CpG sites were within 0.5 Mb of BW SNPs, including cg00423969 and cg14163484 (*FERIL5*), cg00510149 (*IRAG1*), cg02286857 (*TTC7A*), cg15258080 (*HK1*), cg22673972 (*SLC6A6*) and cg24893073 (*KDM6B*) (Supplementary Data 6).

Comparison with maternal ppBMI-associated CpGs in cord blood DNAm. We assessed whether the DNAm signatures of maternal ppBMI in the placenta were consistent with associations in cord blood previously reported by the PACE consortium⁹. We did not find any overlapping CpGs associated with maternal ppBMI between the two tissues. However, we reported three maternal ppBMI-associated CpGs in the placenta that were less than 0.5 Mb upstream or downstream from CpGs that had been associated with maternal ppBMI in cord blood: two out of the 3 *loci* identified showed consistent effect directions of the association with maternal ppBMI in both tissues (Supplementary Data 7).

Discussion

As far as we know, this is the largest EWAS meta-analysis conducted to date on placental DNAm. We have analyzed a total of 2631 mother-child pairs from 10 different PACE cohorts from Europe, America and Australia. We have identified 27 CpGs associated to maternal ppBMI, some of which showed up to 2% lower DNAm per 10-unit higher BMI. Although such a difference in BMI is unlikely in an individual woman in the context of pregnancy interventions, we consider that it could represent the difference between women in the normal range of BMI and women with BMI in the obesity category.

The most significant association was observed for cg08219219, located in the eighth exon of *EPHX3*, for which a ppBMI difference of 10 units is associated with a 1.1% lower placental DNAm. It has been shown that soluble epoxide hydrolases such as *EPHX3* have higher activity in obese mice³⁹. Additionally, it has been suggested that this family of hydrolases could act as therapeutic targets for metabolic and cardiovascular abnormalities related to obesity⁴⁰. We highlighted two other significant hits showing the largest positive and negative beta-coefficients. cg14704941, in the first intron of *CSRP3*, presented with 2% higher placental DNAm per 10-unit ppBMI. *CRSP3* knockout mice develop dilated cardiomyopathy with hypertrophy and heart failure after birth⁴¹. The beta-coefficient of cg04724807, located upstream of *SYT16*, represented about 1.8% lower placental DNAm per 10-unit greater ppBMI. *SYT16* is over-expressed in pancreatic islet cells upon high glucose challenge and is thought to play a role in insulin secretion⁴². As previously stated, maternal obesity has been described to be associated with obesity, diabetes and cardiometabolic conditions in offspring later in life^{4,5}. The fact that our EWAS identified CpGs near these metabolically relevant genes highlights the plausibility that they may play a role in the link between maternal obesity and future health outcomes in children.

Among our significant signals, we also found two CpG sites, cg00423969 and cg14163484, 1.5 kb upstream of the *FERIL5* promoter, presenting lower placental DNAm levels associated with higher maternal ppBMI. Remarkably, *FERIL5* encodes a dysferlin- and myoferlin-related protein, which has been predicted to have a role in vesicle trafficking and muscle membrane fusion events⁴³. Both vesicle trafficking and membrane fusion are crucial events in placental development, since they allow the formation of the syncytiotrophoblast, an uninterrupted and

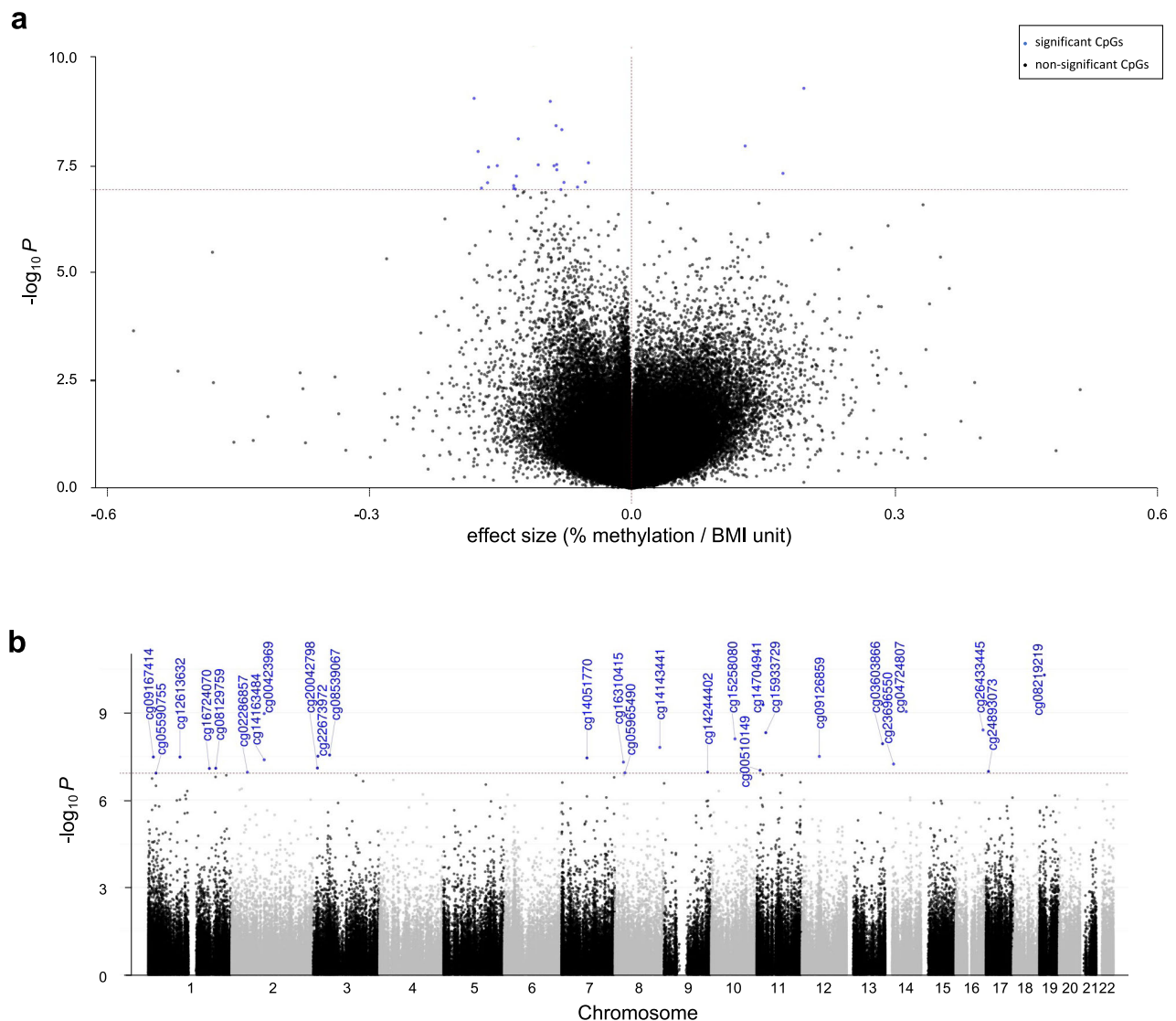


Fig. 2 Association between maternal ppBMI and placental DNAm ($N = 2631$ placental DNA samples), after adjusting for maternal age, parity, maternal education, maternal smoking and putative cellular heterogeneity. Association results are displayed as **a** volcano plot, where the X-axis shows the effect sizes (ranging between 0 and 1) in DNAm and **b** Manhattan plot, where the X-axis represents the genomic location of each CpG. In both panels blue dots indicate significantly associated CpGs (meta-analysis p -value $< 1.2e-07$).

multi-nucleated mass that covers the placental *villi* and enables the interplay with the mother⁴⁴. In addition, *CMIP* and *GPX1*, two of the genes annotated to maternal ppBMI-associated CpGs, may present relevant biological roles in pregnancy. For example, different CpGs surrounding *CMIP* have been associated with pre-eclampsia in a placental DNAm study⁴⁵. *GPX1* is an antioxidant gene and its mRNA levels are lower in the placenta of obese mothers compared to normal-weight mothers⁴⁶. Finally, in the context of obesity, *LGR4*, another gene identified in the present study, bears an activating variant that contributes to abdominal visceral fat accumulation and therefore, to central obesity⁴⁷, suggesting that both genetic and epigenetic regulation at this locus may have a role in obesity-related phenotypes.

Regarding enrichment analyses, one of the most interesting findings is that several significant CpGs are located close to cancer-related genes. It has been recently described that the placenta is organized as a big mass of tumoral clones, with rapid cell divisions that enable selection for good cells that will eventually form the baby. Additionally, cancer and the

syncytiotrophoblast of the placenta are both invasive tissues with many biological parallels^{48,49}. Indeed, it is not surprising that factors that are relevant to the placenta, such as maternal obesity, could affect genes that are relevant to cancer. The other pathway that was enriched for altered genes is oxidative stress. It is well known that excessive fat mass accumulation is linked to oxidative stress. Moreover, peroxisomal fatty acid oxidation seems to be enhanced in the placenta of obese women, while mitochondrial activity is impaired, with a greater lipid storage and an altered transfer of lipids to the fetus⁵⁰. Altogether, there is growing evidence suggesting that obesity-induced oxidative stress is a central factor involved in the risk for adverse outcomes in pregnancy^{51,52}.

Another interesting finding that deserves further investigation is the observation that differentially methylated CpGs are enriched for ZNF217 binding sites. This TF is epigenetically altered in placental cells under hypoxia⁵³, and it has been suggested that maternal obesity during pregnancy causes placental hypoxia⁵⁴. However, whether this TF can drive the methylation machinery to selected regions of the genome and cause epigenetic changes

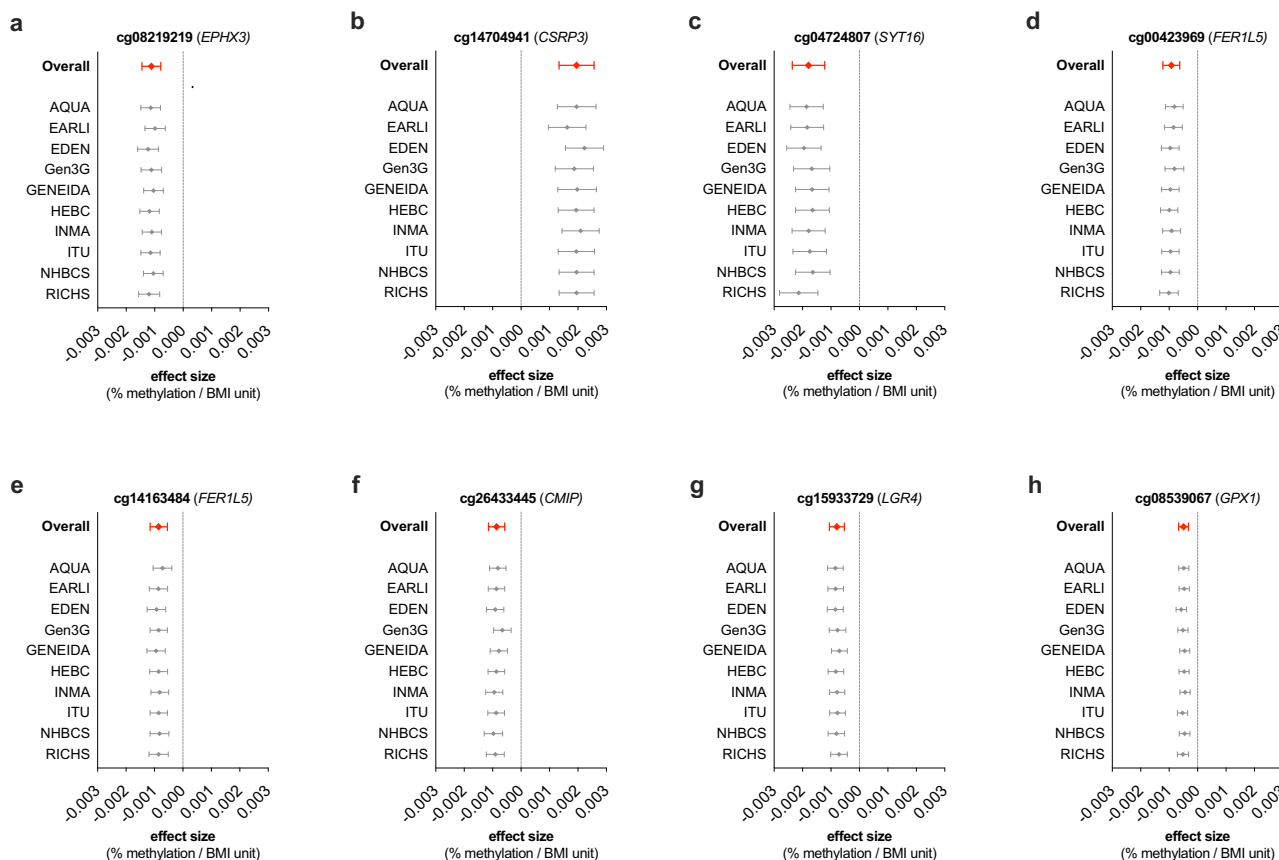


Fig. 3 Forest plots of the leave-one-out analysis showing the fixed effects meta-analysis estimates of association between maternal ppBMI and placental DNAm. Association of **a** cg08219219, **b** cg14704941, **c** cg04724807, **d** cg00423969, **e** cg14163484, **f** cg26433445, **g** cg15933729, **h** cg08539067 with maternal ppBMI. In all panels, cohort names indicate the cohort excluded in each row, and error bars represent the 95% confidence interval of the effect size. Numerical source data for the figure are available in file Supplementary Data 9.

has not yet been explored. Similarly, the overlap between our CpGs and BW-associated regions suggests that both fetal genetic and placental epigenetic factors may contribute to the regulation of fetal growth, but this requires further research.

Our study has notable strengths but also several limitations. As previously mentioned, we have been able to coordinate a large number of cohorts and thus, to obtain an important sample size. Additionally, we have the experience of previous works, in which robust pipelines had already been implemented for EWAS, and we have run the quality control and meta-analysis in two independent institutions. Finally, none of the maternal ppBMI-associated CpGs from the current study are among the problematic probes with absolute methylation differences greater than 10% between Illumina 450 K and EPIC arrays that we identified in a previous study⁵⁵.

Regarding limitations, we did not have access to individual data addressing whether each ppBMI measurement was self-reported or taken at the end of the first trimester of pregnancy. Therefore, we cannot use this variable as a covariate nor compare between measurement types. We are very aware that self-reported ppBMI may not be the most accurate measurement for our variable of interest, as may also be the case of the measurement of BMI at the end of the first trimester. However, self-reporting of ppBMI has been shown to be reliable and highly correlated to measured BMI at 12 weeks of gestation ($r = 0.96$; p -value < 0.0001)⁵⁶. Second, the unavailability of genotype data in some of the participating cohorts did not allow to add genotype principal components to our models and there might be residual confounding by population structure that we did not account for. Third, most of the

cohorts were composed by a majority of individuals from European descent, which limits generalizability of our findings to other populations.

On the other hand, we are aware that RefFreeCellMix, the R package employed for adjustment of cell mixtures, is a principal component analysis-type correction method, and therefore presents the risk of over-correcting the results, especially in dense signal scenarios like the Illumina Beadchips, due to the capture of the signal by some of the top components of the estimation⁵⁷. This, together with the fact that Bonferroni-correction is very strict and that our approach does not take into account the correlation between nearby CpGs, may have let some discoveries out of the focus. However, we have preferred to be strict and report only the most robust results.

In summary, here we present the largest EWAS of maternal ppBMI in association with placental DNAm performed to date. We identify 27 CpG sites at which we observe placental DNAm variations of 0.5–2.0% by 10-unit maternal ppBMI difference. Additionally, our DNAm findings seem to be placenta-specific, showing minimal overlap with a previous meta-analysis in cord-blood DNAm in relation to maternal ppBMI. The differentially methylated CpGs are mainly located in open sea regions, with a complete depletion from CpG islands, and enriched in cancer and oxidative stress-related pathways. These observations, together with the fact that maternal ppBMI is associated with placental DNAm at CpGs located close to obesity-related genes, leads us to hypothesize that placental DNAm could be one of the mechanisms by which maternal obesity is associated with aberrant fetal growth and maybe, other metabolic health outcomes in offspring

later in life. However, we cannot rule out that the changes observed could be markers of exposure to high ppBMI and therefore, our findings will need to be supplemented by functional studies or causal inference analyses to better understand if they truly have a role in pregnancy complications or long-term metabolic outcomes.

Methods

Participating cohorts. Cohorts that are members of the PACE consortium, had existing DNAm data from placental tissue obtained with the Illumina 450 K or EPIC BeadChips, and had maternal BMI information prior to the beginning of pregnancy were invited to participate in the present study. The ten cohorts that contributed to the meta-analysis were AQUA, EARLI, EDEN, Gen3G, GENEIDA, HEBC, INMA, ITU, NHBCS and RICHs. All cohorts obtained ethics approval and informed consent from participants prior to data collection through their Institutional Ethics Boards. Exclusion criteria for this study were: non-singleton births, pre-eclampsia, and DNAm data not derived from the fetal facing side of the placenta. All participants included in this meta-analysis were of European ancestry. Detailed methods for each cohort are provided in Supplementary Note 1.

DNAm data quality control and normalization. All DNAm data processing and analyses were conducted in R 3.3.2⁵⁸, with the exception of the meta-analyses, which were performed with the GWAMA software (<https://genomics.ut.ee/en/tools/gwama>)²⁷. DNAm from the fetal-facing side of the placenta was assessed with the Illumina 450 K or EPIC arrays. See Supplementary Note 1 for extra details on placenta collection, DNA extraction and DNAm acquisition in each cohort. In general, samples were randomized across the different arrays to avoid group differences derived from batch effects. Quality control of DNAm was standardized across all cohorts. Low-quality samples (showing a shifted beta-value distribution) were filtered out and probes with detection p -values >0.01 were excluded (for cohort-specific probe lists see Supplementary Data 8). DNAm beta-values were normalized with functional normalization⁵⁹ and beta-mixture quantile normalization (BMIQ)⁶⁰ was applied to correct for probe type bias. Cohorts examined their data for batch effects by depicting box-plots that divided the samples into different groups according to suspicious variables, and applied ComBat when applicable; all but one cohort (GENEIDA) identified batch effects and used ComBat to remove this source of variation. Probes that were exclusive for the EPIC array, hybridized to the X/Y chromosomes, cross-hybridizing probes and probes with SNPs at the CpG site, extension site, or within 10 bp of the extension site with an average minor allele frequency >0.01 were filtered out⁶¹. Overall, 419,460 probes were available in the ten participating cohorts to assess placental DNAm. Methylation beta-values were modeled in robust linear regressions considering maternal ppBMI as a continuous variable. Finally, DNAm extreme outliers (<25 th percentile - $3 \times \text{IQR}$ or >75 th percentile + $3 \times \text{IQR}$ across all the samples) were trimmed.

Estimates of putative cellular heterogeneity. Putative cellular heterogeneity was estimated from DNAm data using a reference-free cell-mixture deconvolution method (RefFreeCellMix)²⁶. The number of components varied between cohorts and ranged from 2 to 6, maybe because different sampling protocols result in differential heterogeneity across cohorts, or since the approach is data driven, those components could be capturing other major sources of variation in the array data, such as residual technical artifacts. Models for differential DNAm were corrected for the number of surrogate variables minus one to reduce multi-collinearity.

Genome-wide differential DNAm analyses. Within each cohort, robust linear regression from the MASS package⁶² in R was run to account for potential heteroskedasticity and to test the associations between normalized placental DNAm beta-values at each CpG and maternal ppBMI. Models were adjusted for maternal age, parity, maternal education and maternal smoking during pregnancy. Cohorts ran models both with and without adjustment for RefFreeCellMix cell type proportions. Covariate data are described in more detail in Supplementary Note 1.

Meta-analyses. We performed inverse variance-weighted fixed effects meta-analyses using GWAMA²⁷. The meta-analysis was performed independently by two groups to ensure consistent results and identical results were reproduced. We used the Bonferroni adjustment to correct for multiple testing. Secondary analyses were only performed on CpGs that passed the Bonferroni correction, particularly in the RefFreeCellMix-adjusted model. It is worth mentioning that both the meta-analysis and shadow meta-analysis teams performed a general quality control separately prior to the meta-analysis itself, showing consistent results. In summary, we checked that conflictive probes had been removed and drew the cohort-specific qq-plots, as well as the correlation between sample sizes and significant hits across cohorts. Finally, we checked the genomic inflation of the whole meta-analysis and plotted forest plots of the significant hits after leaving one cohort out at a time, to see whether any of the cohorts was guiding the associations. The MARBLES cohort

was excluded due to its small sample size and to the fact that forest plots showed inconsistencies compared to the rest of the cohorts (Supplementary Fig. 1).

Functional and regulatory enrichment analyses. We annotated CpGs to their closest genes and to CpG islands with annotations from the Illumina Human Methylation 450 K annotation file, and with several regulatory features using publicly available data: placental 15-chromatin states³⁰ released from the ROADMAP Epigenomics Mapping Consortium³¹ (ChromHMM v1.10), placental germline differentially methylated regions²⁹ and placental partially methylated domains³².

Over-representation analyses for gene-sets or pathways were performed at the gene level with ConsensusPathDB²⁸ using KEGG, Reactome, Wikipathways and Biocarta as reference databases. ConsensusPathDB performs a hypergeometric test, with a default background equal to the number of ConsensusPathDB entities that are annotated with an ID of the type the user has provided, and participate in at least one pathway. Finally, the program corrects multiple-testing with FDR. Enrichment for TFs was assessed at the gene level with EnrichR using ENCODE and ChEA consensus TFs from ChIP-X database. EnrichR results were ranked using the combined score (p -value computed using Fisher's exact test combined with the z -score of the deviation from the expected rank)⁶³.

Overlap of ppBMI-sensitive CpG sites and birth outcome SNPs. We assessed the genomic proximity between CpGs identified by our maternal ppBMI placental DNAm EWAS (Bonferroni significant in the cell-type adjusted model) and SNPs previously associated with BW, birth length, head circumference and GA³⁰⁻³⁵. Briefly, we verified the genomic proximity between SNPs from the largest GWAS performed to date on the above-mentioned birth outcomes and our identified CpGs by using the Genomic Ranges package⁶⁴ in R, within 1 Mb windows (± 0.5 Mb) surrounding each of the 367 autosomal SNPs.

Comparison of ppBMI-associated CpGs in placenta and in cord blood. We examined whether maternal ppBMI-associated CpGs in placenta were the same as those previously reported in cord blood⁹. As no overlap was found between the hits that passed the Bonferroni correction in each study, we searched for CpGs from the cord blood study present 0.5 Mb upstream or downstream of each of the maternal ppBMI-associated CpGs in placenta (1 Mb windows), by using the GenomicRanges R package, with the aim of finding genomic regions where DNAm was related to ppBMI in the two different tissues.

Statistics and reproducibility. DNAm data from up to 419,460 CpG obtained with the Illumina 450 K or EPIC BeadChips in 2631 placental samples of the fetal side were normalized with functional normalization and BMIQ, and corrected for batch effects with ComBat if applicable. Afterwards, DNAm was correlated with maternal ppBMI in each of the participating cohorts using robust linear regressions with the MASS package in R 3.3.2. The code to perform the full analysis and the details are publicly available as described in the Code Availability section. The cohort-specific results were then meta-analyzed using the inverse variance-weighted fixed effects method in the GWAMA software, simultaneously in two independent laboratories. Results were fully consistent. Statistical significance was set at a meta-analysis nominal p -value = 1.2×10^{-7} , after Bonferroni correction for multiple-testing. Only CpGs below this threshold were taken into account in the downstream analyses. Generally, the statistical methods used in each of the downstream analyses were the ones suggested by the developers of each of the analytic tool implemented. More details in each of the analytical steps included in this study can be obtained in the specific section in Methods, Supplementary Note 1 and the code, as described in the Code Availability section.

Reporting summary. Further information on research design is available in the Nature Portfolio Reporting Summary linked to this article.

Data availability

The full genome-wide results of the meta-analysis are presented in Supplementary Data 3 and 4 and have been uploaded to the EWAS Catalogue, available in Zenodo with identifier <https://doi.org/10.5281/zenodo.7314534>⁶⁵. The individual level data used are not publicly available for several reasons. First, participants were not explicitly informed about this in the informed consent. Second, there are some studies that suggest that DNA methylation data has enough information to identify participants. Third, each PACE cohort follows different internal regulations in regards to public access of the data. Individual level data can still be shared with external researchers after signature of a data transfer agreement (DTA) with each of the participant cohorts, listed in Supplementary Note 1. More information is available in the PACE consortium website (<https://www.niehs.nih.gov/research/atniehs/labs/epi/pi/genetics/pace/index.cfm>). Source data underlying Fig. 3 is presented in Supplementary Data 9.

Code availability

Scripts to reproduce the analysis have been deposited in a public GitHub repository and are available in Zenodo with identifier <https://doi.org/10.5281/zenodo.7313966>.

Received: 5 October 2021; Accepted: 16 November 2022;

Published online: 30 November 2022

References

- Maffei, C. & Morandi, A. Effect of Maternal Obesity on Foetal Growth and Metabolic Health of the Offspring. *Obes. Facts*. **10**, 112–117 (2017).
- Marchi, J. et al. Risks associated with obesity in pregnancy, for the mother and baby: a systematic review of reviews. *Obes. Rev.* **16**, 621–638 (2015).
- Howell, K. R. & Powell, T. L. Effects of maternal obesity on placental function and fetal development. *Reproduction* **153**, R97–R108 (2017).
- Mamun, A. A., Mannan, M. & Doi, S. A. Gestational weight gain in relation to offspring obesity over the life course: a systematic review and bias-adjusted meta-analysis. *Obes. Rev.* **15**, 338–347 (2014).
- Godfrey, K. M. et al. Influence of maternal obesity on the long-term health of offspring. *Lancet Diabetes Endocrinol.* **5**, 53–64 (2017).
- Rivera, H. M., Christiansen, K. J. & Sullivan, E. L. The role of maternal obesity in the risk of neuropsychiatric disorders. *Front. Neurosci.* **9**, 194 (2015).
- Desai, M., Jellyman, J. K. & Ross, M. G. Epigenomics, gestational programming and risk of metabolic syndrome. *Int. J. Obes. (Lond.)* **39**, 633–641 (2015).
- Felix, J. F. et al. Cohort profile: Pregnancy and childhood epigenetics (PACE) consortium. *Int. J. Epidemiol.* **47**, 22–23u (2018).
- Sharp, G. C. et al. Maternal BMI at the start of pregnancy and offspring epigenome-wide DNA methylation: findings from the pregnancy and childhood epigenetics (PACE) consortium. *Hum. Mol. Genet.* **26**, 4067–4085 (2017).
- Sharp, G. C. et al. Maternal pre-pregnancy BMI and gestational weight gain, offspring DNA methylation and later offspring adiposity: findings from the Avon Longitudinal Study of Parents and Children. *Int. J. Epidemiol.* **44**, 1288–1304 (2015).
- Shrestha, D. et al. Placental DNA methylation changes associated with maternal prepregnancy BMI and gestational weight gain. *Int. J. Obes. (Lond.)* **44**, 1406–1416 (2020).
- Thakali, K. M., Zhong, Y., Cleves, M., Andres, A. & Shankar, K. Associations between maternal body mass index and diet composition with placental DNA methylation at term. *Placenta* **93**, 74–82 (2020).
- Everson, T. M. et al. Placental DNA methylation signatures of maternal smoking during pregnancy and potential impacts on fetal growth. *Nat. Commun.* **12**, 5095 (2021).
- Joubert, B. R. et al. DNA Methylation in Newborns and Maternal Smoking in Pregnancy: Genome-wide Consortium Meta-analysis. *Am. J. Hum. Genet.* **98**, 680–696 (2016).
- Muggli, E. et al. Study protocol: Asking QUESIONS about Alcohol in pregnancy (AQUA): A longitudinal cohort study of fetal effects of low to moderate alcohol exposure. *BMC Pregnancy Childbirth* **14**, 302 (2014).
- Newschaffer, C. J. et al. Infant siblings and the investigation of autism risk factors. *J. Neurodev. Disord.* **4**, 7 (2012).
- Heude, B. et al. Cohort Profile: The EDEN mother-child cohort on the prenatal and early postnatal determinants of child health and development. *Int. J. Epidemiol.* **45**, 353–363 (2016).
- Guillemette, L. et al. Genetics of Glucose regulation in Gestation and Growth (Gen3G): a prospective prebirth cohort of mother-child pairs in Sherbrooke, Canada. *BMJ Open* **6**, e010031 (2016).
- Aguilar-Lacasaña, S. et al. Methylene tetrahydrofolate Reductase (MTHFR) Gene Polymorphism and Infant's Anthropometry at Birth. *Nutrients* **13**, 831 (2021).
- Michels, K. B., Harris, H. R. & Barault, L. Birthweight, maternal weight trajectories and global DNA methylation of LINE-1 repetitive elements. *PLoS One* **6**, e25254 (2011).
- Guxens, M. et al. Cohort Profile: The INMA—Infancia y Medio Ambiente—(Environment and Childhood). *Proj. Int. J. Epidemiol.* **41**, 930–940 (2012).
- Dieckmann, L. et al. Characteristics of epigenetic aging across gestational and perinatal tissues. *Clin. Epigenetics* **13**, 97 (2021).
- Hertz-Picciotto, I. et al. A Prospective Study of Environmental Exposures and Early Biomarkers in Autism Spectrum Disorder: Design, Protocols, and Preliminary Data from the MARBLES Study. *Environ. Health Perspect.* **126**, 117004 (2018).
- Gilbert-diamond, D., Emond, J. A., Baker, E. R., Korricks, S. A. & Karagas, M. R. Relation between in Utero Arsenic Exposure and Birth Outcomes in a Cohort of Mothers and Their Newborns from New Hampshire. *Environ. Health Perspect.* **124**, 1299–1307 (2016).
- Appleton, A. A. et al. Prenatal Programming of Infant Neurobehaviour in a Healthy Population. *Paediatr. Perinat. Epidemiol.* **30**, 367–375 (2016).
- Houseman, E. A. et al. Reference-free deconvolution of DNA methylation data and mediation by cell composition effects. *BMC Bioinforma.* **17**, 259 (2016).
- Magi, R. & Morris, A. P. GWAMA: software for genome-wide association meta-analysis. *BMC Bioinforma.* **11**, 288 (2010).
- Kamburov, A. et al. ConsensusPathDB: Toward a more complete picture of cell biology. *Nucleic Acids Res.* **39**, 712–717 (2011).
- Hamada, H. et al. Allele-Specific Methylome and Transcriptome Analysis Reveals Widespread Imprinting in the Human Placenta. *Am. J. Hum. Genet.* **99**, 1045–1058 (2016).
- Ernst, J. & Kellis, M. ChromHMM: automating chromatin-state discovery and characterization. *Nat. Methods* **9**, 215–216 (2012).
- Roadmap Epigenomics Consortium. et al. Integrative analysis of 111 reference human epigenomes. *Nature* **518**, 317–329 (2015).
- Schroeder, D. I. et al. The human placenta methylome. *Proc. Natl Acad. Sci.* **110**, 6037–6042 (2013).
- Horikoshi, M. et al. Genome-wide associations for birth weight and correlations with adult disease. *Nature* **538**, 248–252 (2016).
- Beaumont, R. N. et al. Genome-wide association study of offspring birth weight in 86-577 women identifies five novel loci and highlights maternal genetic effects that are independent of fetal genetics. *Hum. Mol. Genet.* **27**, 742–775 (2018).
- van der Valk, R. J. P. et al. A novel common variant in DCST2 is associated with length in early life and height in adulthood. *Hum. Mol. Genet.* **24**, 1155–1168 (2015).
- Taal, H. et al. Common variants at 12q15 and 12q24 are associated with infant head circumference. *Nat. Genet.* **44**, 532–538 (2012).
- Zhang, G. et al. Genetic Associations with Gestational Duration and Spontaneous Preterm Birth. *N. Engl. J. Med.* **377**, 1156–1167 (2017).
- Warrington, N. M. et al. Maternal and fetal genetic effects on birth weight and their relevance to cardio-metabolic risk factors. *Nat. Genet.* **51**, 804–814 (2019).
- Decker, M. et al. EH3 (ABHD9): the first member of a new epoxide hydrolase family with high activity for fatty acid epoxides. *J. Lipid Res.* **53**, 2038–2045 (2012).
- De Taeye, B. M. et al. Expression and regulation of soluble epoxide hydrolase in adipose tissue. *Obesity* **18**, 489–498 (2010).
- Sun, L. et al. CRISPR/Cas9 mediated establishment of a human CSR3P compound heterozygous knockout hESC line to model cardiomyopathy and heart failure. *Stem Cell Res.* **49**, 102077 (2020).
- Hall, E. et al. The effects of high glucose exposure on global gene expression and DNA methylation in human pancreatic islets. *Mol. Cell. Endocrinol.* **472**, 57–67 (2018).
- Usha Kalyani, R. et al. Fer1L5, a Dysferlin Homologue Present in Vesicles and Involved in C2C12 Myoblast Fusion and Membrane Repair. *Biology* **9**, 386 (2020).
- Knöfler, M. et al. Human placenta and trophoblast development: key molecular mechanisms and model systems. *Cell. Mol. Life Sci.* **76**, 3479–3496 (2019).
- Almomani, S. N. et al. Identification and validation of DNA methylation changes in pre-eclampsia. *Placenta* **110**, 16–23 (2021).
- Diceglio, C. et al. Placental Antioxidant Defenses and Autophagy-Related Genes in Maternal Obesity and Gestational Diabetes Mellitus. *Nutrients* **13**, 1303 (2021).
- Zou, Y. et al. Association of a gain-of-function variant in LGR4 with central obesity. *Obes. (Silver Spring)* **25**, 252–260 (2017).
- Coorens, T. H. H. et al. Inherent mosaicism and extensive mutation of human placentas. *Nature* **592**, 80–85 (2021).
- Costanzo, V., Bardelli, A., Siena, S. & Abrignani, S. Exploring the links between cancer and placenta development. *Open Biol.* **8**, 180081 (2018).
- Calabuig-Navarro, V. et al. Effect of Maternal Obesity on Placental Lipid Metabolism. *Endocrinology* **158**, 2543–2555 (2017).
- Alcala, M. et al. Antioxidants and Oxidative Stress: Focus in Obese Pregnancies. *Front. Physiol.* **9**, 1569 (2018).
- Malti, N. et al. Oxidative stress and maternal obesity: feto-placental unit interaction. *Placenta* **35**, 411–416 (2014).
- Yuen, R. K., Chen, B., Blair, J. D., Robinson, W. P. & Nelson, D. M. Hypoxia alters the epigenetic profile in cultured human placental trophoblasts. *Epigenetics* **8**, 192–202 (2013).
- Fernandez-Twinn, D. S. et al. Exercise rescues obese mothers' insulin sensitivity, placental hypoxia and male offspring insulin sensitivity. *Sci. Rep.* **7**, 44650 (2017).
- Fernandez-Jimenez, N. et al. Comparison of Illumina 450 K and EPIC arrays in placental DNA methylation. *Epigenetics* **14**, 1177–1182 (2019).
- Casas, M. et al. Maternal pre-pregnancy overweight and obesity, and child neuropsychological development: two Southern European birth cohort studies. *Int. J. Epidemiol.* **42**, 506–517 (2013).
- Chen, J. et al. Fast and robust adjustment of cell mixtures in epigenome-wide association studies with SmartSVA. *BMC Genomics* **18**, 413 (2017).
- R Core Team. R: A language and environment for statistical computing. R Foundation for Statistical Computing, Vienna, Austria. URL <https://www.R-project.org/> (2021).

59. Fortin, J.-P. et al. Functional normalization of 450k methylation array data improves replication in large cancer studies. *Genome Biol.* **15**, 503 (2014).
60. Teschendorff, A. E. et al. A beta-mixture quantile normalization method for correcting probe design bias in Illumina Infinium 450 k DNA methylation data. *Bioinformatics* **29**, 189–196 (2013).
61. Chen, J. et al. CpGFilter: model-based CpG probe filtering with replicates for epigenome-wide association studies. *Bioinformatics* **32**, 469–471 (2016).
62. Venables, W. N. & Ripley, B. D. *Modern Applied Statistics with S.* (Springer, 2002).
63. Chen, E. Y. et al. Enrichr: interactive and collaborative HTML5 gene list enrichment analysis tool. *BMC Bioinformatics* **14**, (2013).
64. Lawrence, M. et al. Software for Computing and Annotating Genomic Ranges. *PLOS Computational Biol.* **9**, e1003118 (2013).
65. Fernandez-Jimenez, N. et al. A meta-analysis of pre-pregnancy maternal body mass index and placental DNA methylation identifies 27 CpG sites with implications for mother-child health. [Data set]. *Zenodo*. <https://doi.org/10.5281/zenodo.7314534> (2022).
66. Cilleros-Portet, A. ariadnacilleros/EWAS-maternal-BMI-vs-placental-methylation: v1.0 (v1.0). *Zenodo*. <https://doi.org/10.5281/zenodo.7313966> (2022).

Acknowledgements

We would like to thank the Pregnancy and Childhood Epigenetics (PACE) consortium, as well as all the families that participated in these studies for their generous contribution. This work was partially funded by GVSAN2018111086 from the Basque Department of Health and PI18/01142 from ISCIII - Spanish Ministry of Science and Innovation - cofunded by the ERDF “A way to make Europe” to JRB and LSM, respectively; and by the Joint Programming Initiative – A Healthy Diet for a Healthy Life (JPI HDHL) (NutriPROGRAM). ACP was supported by grant GVSAN2019111085 from the Basque Department of Health to NFJ. Detailed acknowledgements and funding for each participating cohort are described in Supplementary Note 1.

Author contributions

N.F.-J., M.B., M.F.H. & J.R.B. conceived and designed the study. Study-specific analyses were completed by J. Loke. (AQUA), K.M.B. & J.F.D. (EARLI & MARBLES), A.F. & B.H. (EDEN), R.F. (Gen3G), P.C.-S. & J.M.-M. (GENEIDA), A.M.B. (HEBC), N.F.-J. (INMA), T.K. (ITU), F.-Y.T. (NHBCS) and C.L. (RICHES). N.F.-J. & R.F. meta-analyzed the results. N.F.-J. & A.C.-P. performed the follow-up analyses. N.F.-J., R.F., A.C.-P., M.F.H. & J.R.B. interpreted the results. N.F.-J., R.F., A.C.-P., M.F.H. & J.R.B. wrote the first draft of the manuscript. M.F.H. & J.R.B. contributed equally to guiding the direction of the project. All authors (N.F.-J., R.F., A.C.-P., J.L., P.P., T.K., F.-Y.T., C.L., A.M.B., M. Lozano, J.-M.-M., Y.J.L., K.M.B., Y.Z., A.F., S.S., T.M.E., J.C., K.B.M., T.B., P.C.-S., J.H., M.D.F., J.M.L., J.T., D.C., M.F.F., A.G.-M., J.M.C., B.G.-A., R.J.S., J.F.D., E.M., M. Lacasaña,

M.V., C.J.M., M.R.K., K.R., L.B., B.H., L.S.M., M.B., M.F.H. & J.R.B.) made substantial contributions to the acquisition, analysis, or interpretation of data, and read and critically revised the manuscript.

Competing interests

The authors declare no competing interests.

Additional information

Supplementary information The online version contains supplementary material available at <https://doi.org/10.1038/s42003-022-04267-y>.

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Peer review information *Communications Biology* thanks Tina Bianco-Miotto, Chiea Chuen Khor, Markos Tesfaye and the other, anonymous, reviewer(s) for their contribution to the peer review of this work. Primary Handling Editors: Eirini Trompouki, Anam Akhtar, Christina Karlsson Rosenthal and George Inglis. Peer reviewer reports are available.

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