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Review

# Microglia and Cholesterol Handling: Implications for Alzheimer's Disease

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**Abstract:** Cholesterol is essential for brain function and structure, however altered cholesterol metabolism and transport are hallmarks of multiple neurodegenerative conditions, including Alzheimer's disease (AD). The well-established link between apolipoprotein E (APOE) genotype and increased AD risk highlights the importance of cholesterol and lipid transport in AD etiology. Whereas more is known about the regulation and dysregulation of cholesterol metabolism and transport in neurons and astrocytes, less is known about how microglia, the immune cells of the brain, handle cholesterol, and the subsequent implications for the ability of microglia to perform their essential functions. Evidence is emerging that a high-cholesterol environment, particularly in the context of defects in the ability to transport cholesterol (e.g., expression of the high-risk APOE4 isoform), can lead to chronic activation, increased inflammatory signaling, and reduced phagocytic capacity, which have been associated with AD pathology. In this narrative review we describe how cholesterol regulates microglia phenotype and function, and discuss what is known about the effects of statins on microglia, as well as highlighting areas of future research to advance knowledge that can lead to the development of novel therapies for the prevention and treatment of AD.

**Keywords:** microglia; cholesterol; Alzheimer's disease



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## 1. Introduction

Since Alzheimer's disease (AD) was first described by Alois Alzheimer over a century ago [1], it has been known that an aberrant accumulation of lipid "sacculles" as they were called then, essentially an enrichment in intracellular lipids, is characteristic of the brain in AD. Since then we have gained a much more sophisticated understanding of how lipid accumulation and aberrant lipid metabolism contribute to the etiology of AD. It is now clear, for example, that cholesterol enrichment in the plasma membranes of neurons is causally linked to production of the cytotoxic amyloid beta (A $\beta$ ) peptide [2] through a lipid raft-dependent mechanism. Cholesterol is an essential lipid component for various cellular structures and organelles, and its trafficking occurs via various distinct pathways which include endocytosis/phagocytosis, transport to the plasma membrane and repurposing of cholesterol by removal from the plasma membrane [3–17]. Disruption in cholesterol transport and trafficking can lead to aberrant cellular function. For example, in Niemann-Pick Type C disease, which involves an autosomal recessive mutation causing neurodegeneration, aberrant cholesterol metabolism is linked to A $\beta$  deposition similar to the pathology occurring in AD [18]. In mouse models hypercholesterolemia results in glial cell hyperactivation, accelerating amyloid pathology in the brain [19]. In zebrafish exposure to high cholesterol (4% weight/weight cholesterol) for 19 days resulted in higher brain mRNA expression of proinflammatory markers and elevated brain mRNA of cluster of differentiation molecule 11B, a microglia marker, in a type 2 diabetes model [20].

However, it is not yet clear whether unregulated cholesterol drives pathology or if cholesterol is simply worsening an already pathological process. Much more is known about the cholesterol-mediated regulation of cellular function in neurons and astrocytes in the

context of AD [21–23]; however not as much is known about how microglia, the immune cells of the brain, respond to high cholesterol environments. Microglia are quickly emerging as key players in the pathophysiology of AD [24–29]. Genome-wide association studies have consistently found that genes expressed predominantly or exclusively in microglia in the brain, such as triggering receptor expressed on myeloid cells 2 and myeloid cell surface antigen CD33, are associated with AD risk [30–34]. Disease-associated microglia (DAMs), or activated microglia found to be enriched in plaque areas of the AD brain [25,35–39], are directly linked to neuroinflammation, which has been shown to be a critical driver of pathology in AD [40]. Although a vast literature from the cardiovascular field points to cholesterol accumulation in macrophages, the peripheral cousins of microglia, in causing the formation of “foam cells” which are in turn causally linked to the process of atherosclerosis [41–43], not much is known about how microglia respond to excessive cholesterol accumulation.

Multi-omic analysis of postmortem brain samples from AD patients revealed that the concentrations of free cholesterol were elevated in microglial endocytic vesicles [44] suggesting that microglial cholesterol handling may be directly involved in AD pathology. Importantly, it has been shown that the ability of microglia to degrade A $\beta$  is dependent on their ability to efflux cholesterol [45]. Thus, there is already compelling evidence that cholesterol-mediated regulation of microglial function may be an important contributing factor in AD. Here, we review what is known about cholesterol-mediated regulation of microglia phenotype and function, discuss the impacts of statins, which inhibit the synthesis of cholesterol, on microglia function, and highlight areas of future research in the context of AD.

## 2. Cholesterol-Mediated Regulation of Microglia Phenotype

Microglia are classically characterized as immune cells predominantly found in a homeostatic state, only moving outside of that resting or homeostatic phenotype to respond acutely to the infiltration of foreign and harmful substances [46,47]. However, recent studies have revealed that microglia are in fact very dynamic, occupying a diverse set of states, with different functional phenotypes [48,49]. Microglia perform different roles in different environments, falling into four broad categories: injury-response microglia, proliferative-region-associated microglia (PAM), disease-associated microglia (DAM), and lipid droplet-accumulating microglia. Injury-response microglia can be induced by lysolecithin injection in mice [50], which leads to the upregulation of lipoprotein lipase and apolipoprotein E (APOE) among other effects [50]. Microglia that surround oligodendrocytes during the first week after birth are highly phagocytic, identified as PAM, and share a very similar profile at the transcriptional level with DAM, such as the increased expression of lipid metabolic genes [51]. Researchers have identified the transition into the DAM state by the downregulation of typical microglia markers (e.g., C-X3-C motif chemokine receptor 1 and adenosine diphosphate receptor P2Y12) and the activation of phagocytic and lipid metabolic genes; as well as having overlapping signatures with injury-response microglia and PAM [50,51]. Elements of the DAM profile are highly conserved from PAM to DAM in mouse models [36,51], behaving with similar phenotypic states as PAM in the zebrafish model [52], indicating that both processes are highly regulated. Even though the murine DAM gene profile from an AD-inducible model is detected in human DAM, the majority of those genes are dispersed across multiple subgroups of microglia instead of being specific to DAM in humans, highlighting species-specific differences in microglia phenotype [53].

Lipids are essential to the brain for both structure and function [54]; however, an overload of lipids can lead to catastrophic damage. When lipid molecules in a given area surpass a critical concentration they begin to aggregate, form micelles, and even form crystals (e.g., cholesterol crystals), which then act as detergents and structures that lyse and otherwise damage cells and their components [55]. Thus, lipid storage and disposal mechanisms have emerged to protect cells from these extreme events. Cells usually respond to excess cholesterol by inducing the cholesterol efflux machinery, including APOE [56]. Although beyond the scope of this review article, cholesterol metabolism and regulation have been described thoroughly elsewhere [57]. Briefly, the sterol regulatory element

binding protein family is involved in regulating genes involved in cholesterol synthesis, transport, and efflux. Induction of the sterol regulatory element binding protein transcriptional program in response to low or high concentrations of cholesterol detected in the endoplasmic reticulum membrane activates and deactivates, respectively, the transcription of genes involved in lipid biosynthesis and import vs. storage and efflux [58,59], including the expression of low-density lipoprotein receptor [58,60] for cholesterol import, and the adenosine triphosphate-binding cassette (ABC) family, in particular ABCA1, for cholesterol efflux [61–63]. However, when the capacity to utilize or efflux excess lipids has been surpassed all cells have the ability to form lipid droplets as a way to temporarily store the excess [64]. When the storage of lipids in lipid droplets becomes chronic and/or the number of lipid droplets starts to exceed the normal threshold, there can be effects on the ability of cells to perform their normal functions [65].

In the case of microglia, lipid droplet-accumulating microglia, or microglia that are characterized by lipid droplet accumulation, perform with a defective phagocytic phenotype [66]. Lipid droplet-accumulating microglia have an enhanced phagocytic uptake of lipid, exacerbating the lipid droplet accumulation burden, and promoting chronic and self-sustained microglial activation [67]. Sustained inflammation further pushes microglia into an activated state of hyperactivity, which creates a feedback that exacerbates neuroinflammation and damages blood–brain barrier integrity [68]. Whereas cholesterol excess leads to lipid droplet accumulation and chronic microglial activation, dysregulated cholesterol concentration in the opposite direction can also be problematic. For example, in mouse models with an interleukin-10 receptor knockout (specifically in astrocytes) there is prolonged neuroinflammatory response to peripheral lipopolysaccharide (LPS), with interleukin-10 receptor signaling deficits and a lack of cholesterol biosynthesis both leading to the inability to resolve microglial activation [69]. It is still unclear in which context injury-response microglia, PAM and DAM become aggressors vs. protectors [70], but there is ongoing research to understand the dynamics of change in microglia phenotype so that we can better understand how to intervene to modulate phenotype to prevent or decrease neurodegeneration [71].

### 3. Cholesterol-Mediated Regulation of Microglia Function

One of the main functions of microglia is to remove debris and other cytotoxic molecules in a constant effort to maintain a homeostatic environment [48,72]. When microglia fail to perform this essential function a number of downstream effects can occur and lead to disease development. For example, the failure to keep up with the clearance of A $\beta$  monomers contributes to the formation of A $\beta$  oligomers and eventually plaques, which are a hallmark of AD pathophysiology [73–75].

One of the metabolic pathways, in addition to cholesterol efflux and storage, that can be activated in the presence of cholesterol is the generation of a variety of cholesterol metabolites, which in turn can either act as regulators of downstream pathways or have direct deleterious effects. Oxysterols are generated in animals, including humans, by enzymatic means as well as non-enzymatic means. Cholesterol can be oxidized to 25-hydroxycholesterol by cholesterol 25-hydroxylase, and to 27-hydroxycholesterol by sterol 27-hydroxylase [76]. While 25-hydroxycholesterol can also be generated by non-enzymatic oxidation by reactive oxygen species (ROS), 7-ketocholesterol is generated exclusively through non-enzymatic means, for example via oxidation by ROS [76]. Oxidized cholesterol is fundamental for generating a pro-inflammatory environment for microglia [77]. In rodent microglia, cholesterol oxides confer cytotoxic effects by potentiating the effects of LPS and nitric oxide production, promoting programmed cell death [78]. Specifically, 25-hydroxycholesterol was observed to induce the highest mRNA levels of nitric oxide synthase in combination with LPS in these cells [78]. Similarly, 27-hydroxycholesterol *in vitro* treatment of rodent microglia cell lines induced accumulation of ROS and the subsequent activation of the pro-inflammatory interleukin-6/signal transducer and activator of transcription 3 signaling pathway [79]. In turn, in the presence of increased ROS the proportion of sterols of non-enzymatic origin increases, and promotes a chronic DAM state [80]. Studies have also shown that 25-hydroxycholesterol can

increase the area of the lipid bilayer as well as affecting the orientation of lipids within the membrane [81], increasing membrane permeability [82–84], which has a direct influence on cell death activation [85]. It has also been reported that 27-hydroxycholesterol can induce cellular senescence in microglia through oxidative damage [79,86,87], and that 7-ketocholesterol promotes cellular death by altering biogenesis and peroxisomal activity through oxidative stress [88,89]. Investigators report 7-ketocholesterol released during chronic inflammation indirectly induces neuronal damage mediated by activated microglial cells [90]. It is not yet clear how the relative and absolute concentrations of all of these cholesterol species impact overall microglial function and phenotype.

There is evidence that when microglia are unable to maintain proper cholesterol metabolism and lipid droplets accumulate a pro-inflammatory lipidomic profile emerges [91]. A number of approaches to reduce this cholesterol-induced microglia dysfunction have been investigated. The liver X receptor, which is induced by oxysterols, agonistically promotes an anti-inflammatory environment in the central nervous system (CNS) of rodent models and their primary microglia [92]. Liver X receptor-mediated suppression of inflammation and lipid recycling has also been shown to mitigate disease severity at the microglial level in rodent models [93]. These reports in rodent models suggest that reducing cholesterol via liver X receptor activation could be an approach for clearing the burden from microglia and restoring their functionality.

An additional way in which cholesterol can negatively impact microglia function is through mechanisms involving membrane proteins [94]. The enrichment of cholesterol in plasma membranes potentiates the formation of lipid rafts, which increases the physical proximity of raft-associated proteins. An example of how this can be detrimental when excessive is the case of overactivation via LPS. In a high-cholesterol membrane environment, monomers of Toll-like receptors are in close proximity to each other, enabling the formation of Toll-like receptor dimers, which in turn leads to pro-inflammatory signaling in response to activation by LPS [95]. In murine models chronic LPS activation leads to increased A $\beta$  deposition [96,97].

A high cholesterol diet (3% cholesterol) has been shown to induce a pro-inflammatory profile in rodent microglia models by activating the inflammasome; anti-inflammatory cytokines are also secreted as part of the response, but the result is ultimately a damaged blood-brain-barrier [98]. Furthermore, the literature indicates that cholesterol load causes chronic inflammation in microglia [99]. A high fat, high cholesterol diet (21% fat), administered for 18 weeks increased the presence of interleukin-6 in the microglia and plasma of wild-type and APOE<sup>-/-</sup> mice [100]. The APOE4 isoform of APOE has been consistently associated with an increased risk for AD in genome-wide association studies [101]. The ApoE4 protein encoded by the APOE4 gene has been shown to have a significantly reduced capacity to induce cholesterol efflux from a variety of cell types compared to APOE3 [102]. In a human microglia cell line expressing APOE4 it was observed that excess cholesterol leads to higher levels of inflammation [103], highlighting that a reduced capacity to efflux cholesterol, particularly in an environment of excess cholesterol, is associated with microglial activation. Together, these findings suggest that a high cholesterol environment, particularly in genetically susceptible individuals with a reduced capacity to transport and efflux cholesterol (e.g., APOE4 carriers) leads to chronic microglia inflammation and activation, reducing the ability of microglia to respond to additional stressors.

In the CNS, increasing cholesterol leads to reduced phagocytosis by phagocytes [104], and conversely, depleting cholesterol with methyl- $\beta$ -cyclodextrin increases phagocytosis. Depleting cholesterol using methyl- $\beta$ -cyclodextrin enhanced phagocytic activity in primary rat microglia when treated with cholesterol and LPS [105]. Alternatively, one group reported that the accumulation of esterified cholesterol in microglia as a result of the dysfunction of the transmembrane structure triggering receptor expressed on myeloid cells 2, a receptor for lipidated ApoE and other lipids [106,107], did not evoke changes in their phagocytic capacities [108]; suggesting that the concern should not lie solely on the amount of cholesterol microglia are exposed to, but their capacity to traffic cholesterol accordingly. Moreover, microglia cultured from ABCA1 <sup>-B/-B</sup> mice, exhibit augmented

LPS-induced secretion of tumor necrosis factor  $\alpha$  (TNF- $\alpha$ ) and decreased phagocytic activity hand in hand with decreased ABCA1/APOE expression, which are involved in cellular cholesterol efflux [56,109,110]. Loss-of-function of ABCA7, which also impairs the ability of microglia to efflux cholesterol, accelerated enzymatic activity on the amyloid precursor protein, impaired microglial A $\beta$  clearance and impaired the ability of microglia to perform phagocytosis, contributing to the development of AD [111,112]. In a mouse AD model using a knockout of the protein translocator protein 18 kDa, a molecular sensor specific to glial cells in the brain, it was shown that there is increased A $\beta$  deposition in the brain and a decreased number of microglia undergoing phagocytosis compared to control mice [113], highlighting the importance of effective microglial phagocytosis in the prevention of AD.

Accumulating evidence is demonstrating that defects in cholesterol metabolism, particularly in the context of a variety of high cholesterol environments, are deleterious to microglia function. However, it is not yet clear how to reverse this defective phenotype and correct the underlying cholesterol transport defect in order to improve microglia function. It is also not yet clear whether the DAM phenotype can be reversed.

#### 4. Effects of Statins on Microglia

Statins are among the most highly prescribed drugs for the management of hypercholesterolemia (plasma total cholesterol >200 mg/dL) and cardiovascular disease risk. As a result, millions of patients have been prescribed this class of drugs, which inhibit cholesterol biosynthesis. Clinical studies have shown that patients treated with statins, or a combination of statins plus medications to control blood pressure, have a reduced risk for developing AD and dementia [114,115]. Furthermore, patients who were exposed to angiotensin-receptor blockers had lower A $\beta$  deposition in the brain measured with positron emission tomography [116,117]. Statins have been found to have generally anti-inflammatory properties [118], which may be part of their mechanism of action in the context of AD. It has been established that statins have the ability to cross the blood–brain barrier [119]. Thus, it is plausible that this class of drugs could have direct effects on a variety of cells within the CNS, including microglia. In experimental AD rodent models, the statin atorvastatin had a positive impact on cognitive performance and dampened the activated state of microglia, downregulating inflammatory signaling at the mRNA level [120], suggesting that the inhibition of cholesterol synthesis in the context of AD may have beneficial effects via the reduction in neuroinflammation associated with microglia. It has been observed that in microglia statins can lead to a number of effects, including effects on phagocytic activity, on cell activation, and inflammatory signaling.

Firstly, statins have been found to improve phagocytic activity in human microglia and comparable cell models; however, with some unresolved questions about the underlying mechanisms involved [121–124]. Statins increase phagocytic activity in microglia and glia-like cells in part by inhibiting the secretion of pro-inflammatory signals, including TNF- $\alpha$  [125,126]. However, some studies found that changes in phagocytic activity may be linked to both cholesterol-dependent and cholesterol-independent mechanisms, with both increases and decreases in pro-inflammatory cytokine expression reported [105,127]. Further research is needed to understand both the cholesterol-dependent and cholesterol-independent effects of different statin formulations on microglia phagocytic activity.

Chronic microglia activation and concurrent pro-inflammatory signaling can result in damage to neighboring cells [90] and in turn diminish the capacity of microglia to undergo normal function [96,97]. The impact of statins on inflammatory signaling has been explored in rat cellular models, where an induced LPS inflammatory response in microglia was dampened by reducing the expression of pro-inflammatory cytokines that include TNF- $\alpha$  [128]. Statins are directly involved in attenuating the effects of TNF- $\alpha$ , and the release of associated factors and receptors in this signaling cascade [120,129]. Furthermore statins can also have an inhibitory effect on the production of superoxide free radical [123,130], protecting cells from oxidative damage, inducing the production of ionized calcium-binding adaptor molecule 1, regulating lipid metabolism and phagocytosis [131–133].

There are also several lines of investigation suggesting that statins can reduce microglial activation and as a result reduce the propensity for neurodegenerative disease [124,134]. Statins can inhibit microglial activation by blocking the activation of pro-inflammatory cascade pathways [25,130,135] and by promoting the enzymatic degradation of A $\beta$  [136]. Statins have also been found to attenuate the expression of matrixins in microglia, reducing pro-inflammatory signaling [130,137,138], inhibiting the expression of prostaglandin-endoperoxide synthase 2, and redirecting microglia away from an activated state and toward a homeostatic state [130,139]. Studies in human cell lines and in rat neonatal cortical microglia found that atorvastatin reduced the secretion of pro-inflammatory cytokine interleukin-6 [125]. However, simvastatin was not able to reduce pro-inflammatory cytokine interleukin-6 secretion and hindered cell viability in both human microglia cell lines and rat neonatal cortical microglia [125]. Thus, it is important to note that the effects of different statin drugs may be differential with respect to their specific effects on microglia.

An increase in APOE expression has been demonstrated to be a defining factor in the conversion of homeostatic microglia to DAM [36]. Several lines of evidence have now shown that statins may be important in regulating microglia function in part through their effects on APOE. Administration of statins reduced brain APOE mRNA levels, ApoE protein content and ApoE secretion, attenuating the effects of the high-cholesterol diet [140]. In vitro experiments have also shown that APOE expression can be increased in the CNS in response to cellular stress, and that simvastatin was able to reduce that expression [141]. In mice, inhibition of TNF- $\alpha$  signaling increased APOE mRNA and protein levels, whereas inflammatory signaling otherwise dampens APOE expression in microglia [142]. It is still unclear how these various anti-inflammatory mechanisms are activated by the different statin formulations, and the different hypotheses from different reports dependent on the model under observation [143]. However, the evidence thus far, as shown in Table 1, suggests that statins could be a viable approach, whether directly through cholesterol-mediated or indirectly through cholesterol-independent mechanisms, to modulate microglial activation and functionality.

**Table 1.** Summary of studies highlighting the effects of statins on microglia.

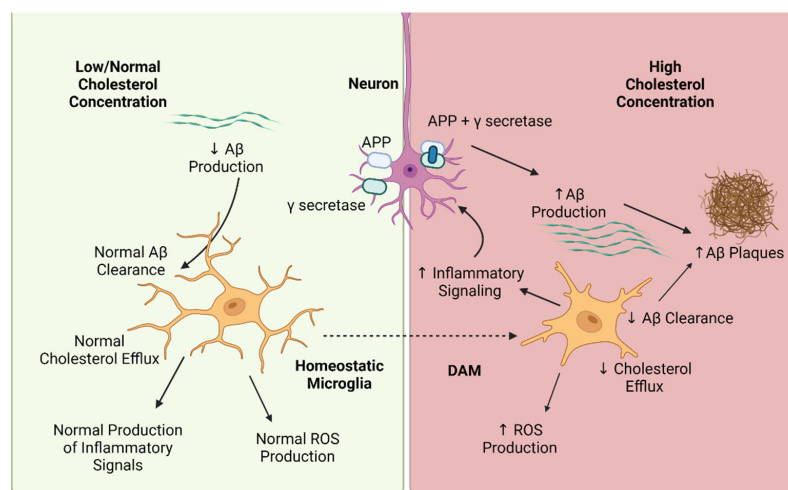
Study	Study Type	Model	Statin (Dose)	Outcome Measure	Results
Wang, 2018 [120]	In vivo	Sprague Dawley male rats (age 7 to 8 weeks), 250–300 g	Atorvastatin 5- and 10-mg/kg (chronic)	Number of Iba-1-positive microglia	Reduced number of Iba-1 positive microglia.
Ewen, 2013 [122]	In vivo	Sprague–Dawley male rats (12 week of age)	Atorvastatin 2, 5, and 10 mg/kg	TNF- $\alpha$ and IL-10 levels, and infiltration at site of injury	Atorvastatin decreased TNF- $\alpha$ and increased in IL-10 levels, and number of activated microglia.
Lindberg, 2005 [125]	In vitro and in vivo	CHME-3 human cell line; primary rat microglia	Atorvastatin 0.1, 1, 5, and 20 mM or Simvastatin 0.1, 1, 5, and 10 mM; Atorvastatin 1, 5, and 20 $\mu$ M	Microglial Secretion of IL-6	Atorvastatin reduce IL-6 secretion of stimulated human and rat microglia.
Townsend, 2004 [127]	In vitro and in vivo	BALB/c mice microglia	Lovastatin 10 $\mu$ M	IL-6, TNF- $\alpha$ and IL- $\beta$ 1 concentrations and phagocytosis activity	Lovastatin reduced IL-6, TNF- $\alpha$ and IL- $\beta$ 1 concentrations and attenuated impaired phagocytosis in primary mouse microglia.
Yongjun, 2013 [130]	In vitro	Primary human microglia	Atorvastatin 0.1 mM	MT1-MMP expression	Reduced microglia expression of MT1-MMP.
Kata, 2016 [131]	In vitro	Primary rat microglia	Rosuvastatin 1 $\mu$ M	Iba-1 immunoreactivity; phagocytosis activity; IL-10, IL-1b and TNF- $\alpha$ production.	In microglia challenged with LPS, rosuvastatin reduced IL-1 $\beta$ , TNF- $\alpha$ production and phagocytosis, IL-10 and Iba1 immunoreactivity was increased.
Chu, 2015 [135]	In vivo	Sprague–Dawley male rats	Atorvastatin 10 mg/kg/day	pNF $\kappa$ B immunostaining	Proinflammatory pNF $\kappa$ B proteins were decreased by atorvastatin in microglia, following surgery.

Table 1. Cont.

Study	Study Type	Model	Statin (Dose)	Outcome Measure	Results
Tamboli, 2010 [136]	In vivo	BV-2 mouse microglia	Lovastatin 5 $\mu$ M	A $\beta$ degradation, Western blot	Lovastatin enhanced the degradation of extracellular A $\beta$ by microglial cells.
Petanceska, 2003 [140]	In vivo	C57/BL6 mice and BV-2 cell line	Lovastatin 5 $\mu$ M and atorvastatin 5 $\mu$ M	ApoE Western blot	In mice, lovastatin reduced ApoE secretion. Atorvastatin reduced the levels of both cellular and secreted ApoE.

## 5. Conclusions

Exposure to excess cholesterol has been shown to drive several aspects of pathological microglia states, including increased inflammatory signaling and decreased phagocytic capacity, both of which have been implicated in AD pathophysiology. The overall picture that is emerging, as depicted in Figure 1, is that in a high cholesterol environment, high cholesterol concentrations in neuronal plasma membranes lead to higher production of A $\beta$  due to colocalization of amyloid precursor protein and  $\gamma$  secretase, which in turn leads to a higher burden of A $\beta$  that needs to be cleared by microglia. At the same time, high cholesterol concentrations impair the ability of microglia to clear A $\beta$ , and increase microglial inflammatory signaling and ROS production. All of this further drives the accumulation of A $\beta$  oligomers and eventually plaque formation, as well as creating a pro-inflammatory environment that contributes to neurodegeneration. The highly prescribed, generally safe, statin drugs may be a viable approach for regulating cholesterol and reducing neuroinflammation but further research on their specific effects on microglia are needed. Further research is also needed to understand how defective or suboptimal cholesterol metabolism impacts microglia phenotype and function so that novel targeted therapies to restore microglia to their functional, homeostatic state can be developed.



**Figure 1.** High cholesterol concentrations and disease-associated microglia (DAM). When cholesterol concentrations are low or normal the amount of amyloid- $\beta$  (A $\beta$ ) production is low, coupled with normal microglia A $\beta$  clearance, and a homeostatic microglia phenotype with normal levels of cholesterol efflux, normal production of inflammatory signals and reactive oxygen species (ROS). Conversely, exposure to high cholesterol concentrations leads to increased production of A $\beta$  by neurons due to colocalization of amyloid precursor protein (APP) and gamma secretase in the plasma membrane. Concurrently, high cholesterol concentrations lead to induction of the DAM phenotype, in which increased inflammatory signaling, increased production of ROS, and decreased cholesterol efflux hinder the ability of microglia to clear A $\beta$ , further increasing the concentration of A $\beta$  oligomers and driving plaque formation. Created with [BioRender.com](https://www.biorender.com).



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## Abbreviations

AD: Alzheimer's disease, A $\beta$ : amyloid beta, ABC: adenosine triphosphate-binding cassette, APOE: apolipoprotein E, CNS: central nervous system, DAM: disease-associated microglia, LPS: lipopolysaccharide, PAM: proliferative-region-associated microglia, ROS: reactive oxygen species, TNF- $\alpha$ : tumor necrosis factor  $\alpha$ .

## References

1. Hippius, H.; Neundörfer, G. The Discovery of Alzheimer's Disease. *Dialogues Clin. Neurosci.* **2003**, *5*, 101–108. [[CrossRef](#)] [[PubMed](#)]
2. Wang, H.; Kulas, J.A.; Wang, C.; Holtzman, D.M.; Ferris, H.A.; Hansen, S.B. Regulation of Beta-Amyloid Production in Neurons by Astrocyte-Derived Cholesterol. *Proc. Natl. Acad. Sci. USA* **2021**, *118*, e2102191118. [[CrossRef](#)] [[PubMed](#)]
3. Chang, T.Y.; Chang, C.C.Y.; Harned, T.C.; De La Torre, A.L.; Lee, J.; Huynh, T.N.; Gow, J.G. Blocking Cholesterol Storage to Treat Alzheimer's Disease. *Explor. Neuroprot. Ther.* **2021**, *1*, 173–184. [[CrossRef](#)] [[PubMed](#)]
4. Chang, T.-Y.; Yamauchi, Y.; Hasan, M.T.; Chang, C. Cellular Cholesterol Homeostasis and Alzheimer's Disease. *J. Lipid Res.* **2017**, *58*, 2239–2254. [[CrossRef](#)] [[PubMed](#)]
5. Storch, J.; Xu, Z. Niemann-Pick C2 (NPC2) and Intracellular Cholesterol Trafficking. *Biochim. Biophys. Acta* **2009**, *1791*, 671–678. [[CrossRef](#)]
6. Pfeiffer, S.R. NPC Intracellular Cholesterol Transporter 1 (NPC1)-Mediated Cholesterol Export from Lysosomes. *J. Biol. Chem.* **2019**, *294*, 1706–1709. [[CrossRef](#)]
7. Urano, Y.; Watanabe, H.; Murphy, S.R.; Shibuya, Y.; Geng, Y.; Peden, A.A.; Chang, C.C.Y.; Chang, T.Y. Transport of LDL-Derived Cholesterol from the NPC1 Compartment to the ER Involves the Trans-Golgi Network and the SNARE Protein Complex. *Proc. Natl. Acad. Sci. USA* **2008**, *105*, 16513–16518. [[CrossRef](#)]
8. Reverter, M.; Rentero, C.; de Muga, S.V.; Alvarez-Guaita, A.; Mulay, V.; Cairns, R.; Wood, P.; Monastyrskaya, K.; Pol, A.; Tebar, F.; et al. Cholesterol Transport from Late Endosomes to the Golgi Regulates T-SNARE Trafficking, Assembly, and Function. *Mol. Biol. Cell* **2011**, *22*, 4108–4123. [[CrossRef](#)]
9. Liscum, L.; Dahl, N.K. Intracellular Cholesterol Transport. *J. Lipid Res.* **1992**, *33*, 1239–1254. [[CrossRef](#)]
10. Yamauchi, Y.; Yokoyama, S.; Chang, T.-Y. ABCA1-Dependent Sterol Release: Sterol Molecule Specificity and Potential Membrane Domain for HDL Biogenesis. *J. Lipid Res.* **2016**, *57*, 77–88. [[CrossRef](#)]
11. Abi-Mosleh, L.; Infante, R.E.; Radhakrishnan, A.; Goldstein, J.L.; Brown, M.S. Cyclodextrin Overcomes Deficient Lysosome-to-Endoplasmic Reticulum Transport of Cholesterol in Niemann-Pick Type C Cells. *Proc. Natl. Acad. Sci. USA* **2009**, *106*, 19316–19321. [[CrossRef](#)] [[PubMed](#)]
12. Yamauchi, Y.; Iwamoto, N.; Rogers, M.A.; Abe-Dohmae, S.; Fujimoto, T.; Chang, C.C.Y.; Ishigami, M.; Kishimoto, T.; Kobayashi, T.; Ueda, K.; et al. Deficiency in the Lipid Exporter ABCA1 Impairs Retrograde Sterol Movement and Disrupts Sterol Sensing at the Endoplasmic Reticulum. *J. Biol. Chem.* **2015**, *290*, 23464–23477. [[CrossRef](#)] [[PubMed](#)]
13. Das, A.; Brown, M.S.; Anderson, D.D.; Goldstein, J.L.; Radhakrishnan, A. Three Pools of Plasma Membrane Cholesterol and Their Relation to Cholesterol Homeostasis. *eLife* **2014**, *3*, e02882. [[CrossRef](#)]
14. Mesmin, B.; Pipalia, N.H.; Lund, F.W.; Ramlall, T.F.; Sokolov, A.; Eliezer, D.; Maxfield, F.R. STARD4 Abundance Regulates Sterol Transport and Sensing. *Mol. Biol. Cell* **2011**, *22*, 4004–4015. [[CrossRef](#)]
15. Garbarino, J.; Pan, M.; Chin, H.F.; Lund, F.W.; Maxfield, F.R.; Breslow, J.L. STARD4 Knockdown in HepG2 Cells Disrupts Cholesterol Trafficking Associated with the Plasma Membrane, ER, and ERC. *J. Lipid Res.* **2012**, *53*, 2716–2725. [[CrossRef](#)] [[PubMed](#)]
16. Sandhu, J.; Li, S.; Fairall, L.; Pfisterer, S.G.; Gurnett, J.E.; Xiao, X.; Weston, T.A.; Vashi, D.; Ferrari, A.; Orozco, J.L.; et al. Aster Proteins Facilitate Nonvesicular Plasma Membrane to ER Cholesterol Transport in Mammalian Cells. *Cell* **2018**, *175*, 514–529.e20. [[CrossRef](#)]

17. Naito, T.; Ercan, B.; Krshnan, L.; Triebel, A.; Koh, D.H.Z.; Wei, F.-Y.; Tomizawa, K.; Torta, F.T.; Wenk, M.R.; Saheki, Y. Movement of Accessible Plasma Membrane Cholesterol by the GRAMD1 Lipid Transfer Protein Complex. *eLife* **2019**, *8*, e51401. [[CrossRef](#)] [[PubMed](#)]
18. Yamazaki, T.; Chang, T.-Y.; Haass, C.; Ihara, Y. Accumulation and Aggregation of Amyloid  $\beta$ -Protein in Late Endosomes of Niemann-Pick Type C Cells. *J. Biol. Chem.* **2001**, *276*, 4454–4460. [[CrossRef](#)]
19. Więckowska-Gacek, A.; Mietelska-Porowska, A.; Chutorański, D.; Wydrych, M.; Długosz, J.; Wojda, U. Western Diet Induces Impairment of Liver-Brain Axis Accelerating Neuroinflammation and Amyloid Pathology in Alzheimer's Disease. *Front. Aging Neurosci.* **2021**, *13*, 654509. [[CrossRef](#)]
20. Wang, J.; Li, Y.; Lai, K.; Zhong, Q.; Demin, K.A.; Kalueff, A.V.; Song, C. High-Glucose/High-Cholesterol Diet in Zebrafish Evokes Diabetic and Affective Pathogenesis: The Role of Peripheral and Central Inflammation, Microglia and Apoptosis. *Prog. Neuropsychopharmacol. Biol. Psychiatry* **2020**, *96*, 109752. [[CrossRef](#)]
21. Zhang, J.; Liu, Q. Cholesterol Metabolism and Homeostasis in the Brain. *Protein Cell* **2015**, *6*, 254–264. [[CrossRef](#)]
22. Liu, J.-P.; Tang, Y.; Zhou, S.; Toh, B.H.; McLean, C.; Li, H. Cholesterol Involvement in the Pathogenesis of Neurodegenerative Diseases. *Mol. Cell. Neurosci.* **2010**, *43*, 33–42. [[CrossRef](#)]
23. Puglielli, L.; Tanzi, R.E.; Kovacs, D.M. Alzheimer's Disease: The Cholesterol Connection. *Nat. Neurosci.* **2003**, *6*, 345–351. [[CrossRef](#)] [[PubMed](#)]
24. Leng, F.; Edison, P. Neuroinflammation and Microglial Activation in Alzheimer Disease: Where Do We Go from Here? *Nat. Rev. Neurol.* **2021**, *17*, 157–172. [[CrossRef](#)]
25. Hansen, D.V.; Hanson, J.E.; Sheng, M. Microglia in Alzheimer's Disease. *J. Cell Biol.* **2018**, *217*, 459–472. [[CrossRef](#)]
26. Mrak, R.E. Microglia in Alzheimer Brain: A Neuropathological Perspective. *Int. J. Alzheimers Dis.* **2012**, *2012*, 165021. [[CrossRef](#)]
27. Sarlus, H.; Heneka, M.T. Microglia in Alzheimer's Disease. *J. Clin. Investig.* **2017**, *127*, 3240–3249. [[CrossRef](#)] [[PubMed](#)]
28. Streit, W.J. Microglia and Alzheimer's Disease Pathogenesis. *J. Neurosci. Res.* **2004**, *77*, 1–8. [[CrossRef](#)]
29. Giulian, D. Microglia and the Immune Pathology of Alzheimer Disease. *Am. J. Hum. Genet.* **1999**, *65*, 13–18. [[CrossRef](#)] [[PubMed](#)]
30. Escott-Price, V.; Bellenguez, C.; Wang, L.-S.; Choi, S.-H.; Harold, D.; Jones, L.; Holmans, P.; Gerrish, A.; Vedernikov, A.; Richards, A.; et al. Gene-Wide Analysis Detects Two New Susceptibility Genes for Alzheimer's Disease. *PLoS ONE* **2014**, *9*, e94661. [[CrossRef](#)] [[PubMed](#)]
31. Jansen, I.E.; Savage, J.E.; Watanabe, K.; Bryois, J.; Williams, D.M.; Steinberg, S.; Sealock, J.; Karlsson, I.K.; Hägg, S.; Athanasiu, L.; et al. Genome-Wide Meta-Analysis Identifies New Loci and Functional Pathways Influencing Alzheimer's Disease Risk. *Nat. Genet.* **2019**, *51*, 404–413. [[CrossRef](#)] [[PubMed](#)]
32. Bellenguez, C.; Küçükali, F.; Jansen, I.E.; Kleindam, L.; Moreno-Grau, S.; Amin, N.; Naj, A.C.; Campos-Martin, R.; Grenier-Boley, B.; Andrade, V.; et al. New Insights into the Genetic Etiology of Alzheimer's Disease and Related Dementias. *Nat. Genet.* **2022**, *54*, 412–436. [[CrossRef](#)] [[PubMed](#)]
33. Bis, J.C.; Jian, X.; Kunkle, B.W.; Chen, Y.; Hamilton-Nelson, K.L.; Bush, W.S.; Salerno, W.J.; Lancour, D.; Ma, Y.; Renton, A.E.; et al. Whole Exome Sequencing Study Identifies Novel Rare and Common Alzheimer's-Associated Variants Involved in Immune Response and Transcriptional Regulation. *Mol. Psychiatry* **2020**, *25*, 1859–1875. [[CrossRef](#)]
34. Karch, C.M.; Goate, A.M. Alzheimer's Disease Risk Genes and Mechanisms of Disease Pathogenesis. *Biol. Psychiatry* **2015**, *77*, 43–51. [[CrossRef](#)] [[PubMed](#)]
35. Stelzmann, R.A.; Norman Schnitzlein, H.; Reed Murtagh, F. An English Translation of Alzheimer's 1907 Paper, "über Eine Eigenartige Erkrankung Der Hirnrinde". *Clin. Anat.* **1995**, *8*, 429–431. [[CrossRef](#)] [[PubMed](#)]
36. Keren-Shaul, H.; Spinrad, A.; Weiner, A.; Matcovitch-Natan, O.; Dvir-Szternfeld, R.; Ulland, T.K.; David, E.; Baruch, K.; Lara-Astaiso, D.; Toth, B.; et al. A Unique Microglia Type Associated with Restricting Development of Alzheimer's Disease. *Cell* **2017**, *169*, 1276–1290.e17. [[CrossRef](#)]
37. Graeber, M.B.; Kösel, S.; Egensperger, R.; Banati, R.B.; Müller, U.; Bise, K.; Hoff, P.; Möller, H.J.; Fujisawa, K.; Mehraein, P. Rediscovery of the Case Described by Alois Alzheimer in 1911: Historical, Histological and Molecular Genetic Analysis. *Neurogenetics* **1997**, *1*, 73–80. [[CrossRef](#)]
38. Perlmutter, L.S.; Scott, S.A.; Barrón, E.; Chui, H.C. MHC Class II-Positive Microglia in Human Brain: Association with Alzheimer Lesions: MHC Class II Microglia in Human Brain. *J. Neurosci. Res.* **1992**, *33*, 549–558. [[CrossRef](#)]
39. Rozemuller, J.M.; van der Valk, P.; Eikelenboom, P. Activated Microglia and Cerebral Amyloid Deposits in Alzheimer's Disease. *Res. Immunol.* **1992**, *143*, 646–649. [[CrossRef](#)]
40. Cai, Z.; Hussain, M.D.; Yan, L.-J. Microglia, Neuroinflammation, and Beta-Amyloid Protein in Alzheimer's Disease. *Int. J. Neurosci.* **2014**, *124*, 307–321. [[CrossRef](#)]
41. Yu, X.-H.; Fu, Y.-C.; Zhang, D.-W.; Yin, K.; Tang, C.-K. Foam Cells in Atherosclerosis. *Clin. Chim. Acta* **2013**, *424*, 245–252. [[CrossRef](#)]
42. Kruth, H.S. Macrophage Foam Cells and Atherosclerosis. *Front. Biosci.* **2001**, *6*, d429. [[CrossRef](#)] [[PubMed](#)]
43. Yuan, Y.; Li, P.; Ye, J. Lipid Homeostasis and the Formation of Macrophage-Derived Foam Cells in Atherosclerosis. *Protein Cell* **2012**, *3*, 173–181. [[CrossRef](#)]
44. Cohn, W.; Melnik, M.; Huang, C.; Teter, B.; Chandra, S.; Zhu, C.; McIntire, L.B.; John, V.; Gylys, K.H.; Bilousova, T. Multi-Omics Analysis of Microglial Extracellular Vesicles From Human Alzheimer's Disease Brain Tissue Reveals Disease-Associated Signatures. *Front. Pharmacol.* **2021**, *12*, 766082. [[CrossRef](#)] [[PubMed](#)]
45. Lee, C.Y.D.; Tse, W.; Smith, J.D.; Landreth, G.E. Apolipoprotein E Promotes  $\beta$ -Amyloid Trafficking and Degradation by Modulating Microglial Cholesterol Levels. *J. Biol. Chem.* **2012**, *287*, 2032–2044. [[CrossRef](#)]

46. Li, Q.; Barres, B.A. Microglia and Macrophages in Brain Homeostasis and Disease. *Nat. Rev. Immunol.* **2018**, *18*, 225–242. [[CrossRef](#)] [[PubMed](#)]
47. Schlegelmilch, T.; Henke, K.; Peri, F. Microglia in the Developing Brain: From Immunity to Behaviour. *Curr. Opin. Neurobiol.* **2011**, *21*, 5–10. [[CrossRef](#)]
48. Nimmerjahn, A.; Kirchhoff, F.; Helmchen, F. Resting Microglial Cells Are Highly Dynamic Surveillants of Brain Parenchyma in Vivo. *Science* **2005**, *308*, 1314–1318. [[CrossRef](#)] [[PubMed](#)]
49. Peri, F.; Nüsslein-Volhard, C. Live Imaging of Neuronal Degradation by Microglia Reveals a Role for V0-ATPase A1 in Phagosomal Fusion In Vivo. *Cell* **2008**, *133*, 916–927. [[CrossRef](#)]
50. Hammond, T.R.; Dufort, C.; Dissing-Olesen, L.; Giera, S.; Young, A.; Wysoker, A.; Walker, A.J.; Gergits, F.; Segel, M.; Nemes, J.; et al. Single-Cell RNA Sequencing of Microglia throughout the Mouse Lifespan and in the Injured Brain Reveals Complex Cell-State Changes. *Immunity* **2019**, *50*, 253–271.e6. [[CrossRef](#)]
51. Li, Q.; Cheng, Z.; Zhou, L.; Darmanis, S.; Neff, N.F.; Okamoto, J.; Gulati, G.; Bennett, M.L.; Sun, L.O.; Clarke, L.E.; et al. Developmental Heterogeneity of Microglia and Brain Myeloid Cells Revealed by Deep Single-Cell RNA Sequencing. *Neuron* **2019**, *101*, 207–223.e10. [[CrossRef](#)]
52. Wu, S.; Nguyen, L.T.M.; Pan, H.; Hassan, S.; Dai, Y.; Xu, J.; Wen, Z. Two Phenotypically and Functionally Distinct Microglial Populations in Adult Zebrafish. *Sci. Adv.* **2020**, *6*, eabd1160. [[CrossRef](#)]
53. Olah, M.; Menon, V.; Habib, N.; Taga, M.F.; Ma, Y.; Yung, C.J.; Cimpean, M.; Khairallah, A.; Coronas-Samano, G.; Sankowski, R.; et al. Single Cell RNA Sequencing of Human Microglia Uncovers a Subset Associated with Alzheimer’s Disease. *Nat. Commun.* **2020**, *11*, 6129. [[CrossRef](#)]
54. Dawson, G. Measuring Brain Lipids. *Biochim. Biophys. Acta BBA—Mol. Cell Biol. Lipids* **2015**, *1851*, 1026–1039. [[CrossRef](#)] [[PubMed](#)]
55. Cabral, D.J.; Small, D.M. Physical Chemistry of Bile. In *Comprehensive Physiology*; Terjung, R., Ed.; Wiley: Hoboken, NJ, USA, 1989; pp. 621–662.
56. von Eckardstein, A.; Nofer, J.-R.; Assmann, G. High Density Lipoproteins and Arteriosclerosis: Role of Cholesterol Efflux and Reverse Cholesterol Transport. *Arterioscler. Thromb. Vasc. Biol.* **2001**, *21*, 13–27. [[CrossRef](#)]
57. Feingold, K.R. Lipid and Lipoprotein Metabolism. *Endocrinol. Metab. Clin. N. Am.* **2022**, *51*, 437–458. [[CrossRef](#)] [[PubMed](#)]
58. Goldstein, J.L.; DeBose-Boyd, R.A.; Brown, M.S. Protein Sensors for Membrane Sterols. *Cell* **2006**, *124*, 35–46. [[CrossRef](#)] [[PubMed](#)]
59. Radhakrishnan, A.; Goldstein, J.L.; McDonald, J.G.; Brown, M.S. Switch-like Control of SREBP-2 Transport Triggered by Small Changes in ER Cholesterol: A Delicate Balance. *Cell Metab.* **2008**, *8*, 512–521. [[CrossRef](#)]
60. Yokoyama, C.; Wang, X.; Briggs, M.R.; Admon, A.; Wu, J.; Hua, X.; Goldstein, J.L.; Brown, M.S. SREBP-1, a Basic-Helix-Loop-Helix-Leucine Zipper Protein That Controls Transcription of the Low Density Lipoprotein Receptor Gene. *Cell* **1993**, *75*, 187–197. [[CrossRef](#)]
61. Fitzgerald, M.L.; Mendez, A.J.; Moore, K.J.; Andersson, L.P.; Panjeton, H.A.; Freeman, M.W. ATP-Binding Cassette Transporter A1 Contains an NH<sub>2</sub>-Terminal Signal Anchor Sequence That Translocates the Protein’s First Hydrophilic Domain to the Exoplasmic Space. *J. Biol. Chem.* **2001**, *276*, 15137–15145. [[CrossRef](#)] [[PubMed](#)]
62. Oram, J.F. HDL Apolipoproteins and ABCA1: Partners in the Removal of Excess Cellular Cholesterol. *Arterioscler. Thromb. Vasc. Biol.* **2003**, *23*, 720–727. [[CrossRef](#)] [[PubMed](#)]
63. Dean, M.; Hamon, Y.; Chimini, G. The Human ATP-Binding Cassette (ABC) Transporter Superfamily. *J. Lipid Res.* **2001**, *42*, 1007–1017. [[CrossRef](#)] [[PubMed](#)]
64. Fawcett, D.W. *An Atlas of Fine Structure: The Cell, Its Organelles, and Inclusions*; Saunders: Philadelphia, PA, USA, 1966.
65. Farese, R.V.; Walther, T.C. Lipid Droplets Finally Get a Little R-E-S-P-E-C-T. *Cell* **2009**, *139*, 855–860. [[CrossRef](#)]
66. Marschallinger, J.; Iram, T.; Zardeneta, M.; Lee, S.E.; Lehallier, B.; Haney, M.S.; Pluvinage, J.V.; Mathur, V.; Hahn, O.; Morgens, D.W.; et al. Lipid-Droplet-Accumulating Microglia Represent a Dysfunctional and Proinflammatory State in the Aging Brain. *Nat. Neurosci.* **2020**, *23*, 194–208. [[CrossRef](#)] [[PubMed](#)]
67. Colombo, A.; Dinkel, L.; Müller, S.A.; Sebastian Monasor, L.; Schifferer, M.; Cantuti-Castelvetri, L.; König, J.; Vidatic, L.; Bremova-Ertl, T.; Lieberman, A.P.; et al. Loss of NPC1 Enhances Phagocytic Uptake and Impairs Lipid Trafficking in Microglia. *Nat. Commun.* **2021**, *12*, 1158. [[CrossRef](#)] [[PubMed](#)]
68. Hanisch, U.-K.; Kettenmann, H. Microglia: Active Sensor and Versatile Effector Cells in the Normal and Pathologic Brain. *Nat. Neurosci.* **2007**, *10*, 1387–1394. [[CrossRef](#)]
69. O’Neil, S.M.; Hans, E.E.; Jiang, S.; Wangler, L.M.; Godbout, J.P. Astrocyte Immunosenescence and Deficits in Interleukin 10 Signaling in the Aged Brain Disrupt the Regulation of Microglia Following Innate Immune Activation. *Glia* **2022**, *70*, 913–934. [[CrossRef](#)] [[PubMed](#)]
70. Zareba, J.; Peri, F. Microglial ‘Fat Shaming’ in Development and Disease. *Curr. Opin. Cell Biol.* **2021**, *73*, 105–109. [[CrossRef](#)]
71. Badimon, A.; Strasburger, H.J.; Ayata, P.; Chen, X.; Nair, A.; Ikegami, A.; Hwang, P.; Chan, A.T.; Graves, S.M.; Uweru, J.O.; et al. Negative Feedback Control of Neuronal Activity by Microglia. *Nature* **2020**, *586*, 417–423. [[CrossRef](#)]
72. Colonna, M.; Butovsky, O. Microglia Function in the Central Nervous System During Health and Neurodegeneration. *Annu. Rev. Immunol.* **2017**, *35*, 441–468. [[CrossRef](#)]
73. Guglielmotto, M.; Monteleone, D.; Piras, A.; Valsecchi, V.; Tropiano, M.; Ariano, S.; Fornaro, M.; Vercelli, A.; Puyal, J.; Arancio, O.; et al. Aβ<sub>1-42</sub> Monomers or Oligomers Have Different Effects on Autophagy and Apoptosis. *Autophagy* **2014**, *10*, 1827–1843. [[CrossRef](#)] [[PubMed](#)]

74. Baerends, E.; Soud, K.; Folke, J.; Pedersen, A.-K.; Henmar, S.; Konrad, L.; Lycas, M.D.; Mori, Y.; Pakkenberg, B.; Woldbye, D.P.D.; et al. Modeling the Early Stages of Alzheimer's Disease by Administering Intracerebroventricular Injections of Human Native A $\beta$  Oligomers to Rats. *Acta Neuropathol. Commun.* **2022**, *10*, 113. [[CrossRef](#)] [[PubMed](#)]
75. Tamagno, E.; Bardini, P.; Guglielmotto, M.; Danni, O.; Tabaton, M. The Various Aggregation States of  $\beta$ -Amyloid 1–42 Mediate Different Effects on Oxidative Stress, Neurodegeneration, and BACE-1 Expression. *Free Radic. Biol. Med.* **2006**, *41*, 202–212. [[CrossRef](#)] [[PubMed](#)]
76. Brown, A.J.; Jessup, W. Oxysterols: Sources, Cellular Storage and Metabolism, and New Insights into Their Roles in Cholesterol Homeostasis. *Mol. Asp. Med.* **2009**, *30*, 111–122. [[CrossRef](#)] [[PubMed](#)]
77. Gamba, P.; Testa, G.; Gargiulo, S.; Staurengi, E.; Poli, G.; Leonarduzzi, G. Oxidized Cholesterol as the Driving Force behind the Development of Alzheimer's Disease. *Front. Aging Neurosci.* **2015**, *7*, 119. [[CrossRef](#)] [[PubMed](#)]
78. Chang, J.Y.; Chavis, J.A.; Liu, L.-Z.; Drew, P.D. Cholesterol Oxides Induce Programmed Cell Death in Microglial Cells. *Biochem. Biophys. Res. Commun.* **1998**, *249*, 817–821. [[CrossRef](#)] [[PubMed](#)]
79. Liu, J.; Liu, Y.; Chen, J.; Hu, C.; Teng, M.; Jiao, K.; Shen, Z.; Zhu, D.; Yue, J.; Li, Z.; et al. The ROS-Mediated Activation of IL-6/STAT3 Signaling Pathway Is Involved in the 27-Hydroxycholesterol-Induced Cellular Senescence in Nerve Cells. *Toxicol. Vitro.* **2017**, *45*, 10–18. [[CrossRef](#)]
80. Simpson, D.S.A.; Oliver, P.L. ROS Generation in Microglia: Understanding Oxidative Stress and Inflammation in Neurodegenerative Disease. *Antioxidants* **2020**, *9*, 743. [[CrossRef](#)]
81. Olsen, B.N.; Schlesinger, P.H.; Baker, N.A. Perturbations of Membrane Structure by Cholesterol and Cholesterol Derivatives Are Determined by Sterol Orientation. *J. Am. Chem. Soc.* **2009**, *131*, 4854–4865. [[CrossRef](#)]
82. Kauffman, J.M.; Westerman, P.W.; Carey, M.C. Fluorocholesterols, in Contrast to Hydroxycholesterols, Exhibit Interfacial Properties Similar to Cholesterol. *J. Lipid Res.* **2000**, *41*, 991–1003. [[CrossRef](#)]
83. Holmes, R.P.; Yoss, N.L. 25-Hydroxysterols Increase the Permeability of Liposomes to Ca<sup>2+</sup> and Other Cations. *Biochim. Biophys. Acta* **1984**, *770*, 15–21. [[CrossRef](#)] [[PubMed](#)]
84. Theunissen, J.J.; Jackson, R.L.; Kempen, H.J.; Demel, R.A. Membrane Properties of Oxysterols. Interfacial Orientation, Influence on Membrane Permeability and Redistribution between Membranes. *Biochim. Biophys. Acta* **1986**, *860*, 66–74. [[CrossRef](#)] [[PubMed](#)]
85. Appelqvist, H.; Wåster, P.; Kågedal, K.; Öllinger, K. The Lysosome: From Waste Bag to Potential Therapeutic Target. *J. Mol. Cell Biol.* **2013**, *5*, 214–226. [[CrossRef](#)] [[PubMed](#)]
86. Gosselet, F.; Saint-Pol, J.; Fenart, L. Effects of Oxysterols on the Blood–Brain Barrier: Implications for Alzheimer's Disease. *Biochem. Biophys. Res. Commun.* **2014**, *446*, 687–691. [[CrossRef](#)] [[PubMed](#)]
87. Gamba, P.; Leonarduzzi, G.; Tamagno, E.; Guglielmotto, M.; Testa, G.; Sottero, B.; Gargiulo, S.; Biasi, F.; Mauro, A.; Viña, J.; et al. Interaction between 24-Hydroxycholesterol, Oxidative Stress, and Amyloid- $\beta$  in Amplifying Neuronal Damage in Alzheimer's Disease: Three Partners in Crime: 24-Hydroxycholesterol Potentiates Amyloid-Beta Neurotoxicity. *Aging Cell* **2011**, *10*, 403–417. [[CrossRef](#)] [[PubMed](#)]
88. Trompier, D.; Vejux, A.; Zarrouk, A.; Gondcaille, C.; Geillon, F.; Nury, T.; Savary, S.; Lizard, G. Brain Peroxisomes. *Biochimie* **2014**, *98*, 102–110. [[CrossRef](#)]
89. Nury, T.; Yammine, A.; Menetrier, F.; Zarrouk, A.; Vejux, A.; Lizard, G. 7-Ketocholesterol- and 7 $\beta$ -Hydroxycholesterol-Induced Peroxisomal Disorders in Glial, Microglial and Neuronal Cells: Potential Role in Neurodegeneration: 7-Ketocholesterol and 7 $\beta$ -Hydroxycholesterol-Induced Peroxisomal Disorders and Neurodegeneration. In *Peroxisome Biology: Experimental Models, Peroxisomal Disorders and Neurological Diseases*; Lizard, G., Ed.; Advances in Experimental Medicine and Biology: London, UK; Springer International Publishing: Cham, Switzerland, 2020; Volume 1299, pp. 31–41.
90. Diestel, A.; Aktas, O.; Hackel, D.; Håke, I.; Meier, S.; Raine, C.S.; Nitsch, R.; Zipp, F.; Ullrich, O. Activation of Microglial Poly(ADP-Ribose)-Polymerase-1 by Cholesterol Breakdown Products during Neuroinflammation. *J. Exp. Med.* **2003**, *198*, 1729–1740. [[CrossRef](#)]
91. Loving, B.A.; Tang, M.; Neal, M.C.; Gorkhali, S.; Murphy, R.; Eckel, R.H.; Bruce, K.D. Lipoprotein Lipase Regulates Microglial Lipid Droplet Accumulation. *Cells* **2021**, *10*, 198. [[CrossRef](#)]
92. Pascual-García, M.; Rué, L.; León, T.; Julve, J.; Carbó, J.M.; Matalonga, J.; Auer, H.; Celada, A.; Escolà-Gil, J.C.; Steffensen, K.R.; et al. Reciprocal Negative Cross-Talk between Liver X Receptors (LXRs) and STAT1: Effects on IFN- $\gamma$ -Induced Inflammatory Responses and LXR-Dependent Gene Expression. *J. Immunol.* **2013**, *190*, 6520–6532. [[CrossRef](#)]
93. Berghoff, S.A.; Spieth, L.; Sun, T.; Hosang, L.; Schlaphoff, L.; Depp, C.; Düking, T.; Winchenbach, J.; Neuber, J.; Ewers, D.; et al. Microglia Facilitate Repair of Demyelinated Lesions via Post-Squalene Sterol Synthesis. *Nat. Neurosci.* **2021**, *24*, 47–60. [[CrossRef](#)]
94. Račková, L. Cholesterol Load of Microglia: Contribution of Membrane Architecture Changes to Neurotoxic Power? *Arch. Biochem. Biophys.* **2013**, *537*, 91–103. [[CrossRef](#)] [[PubMed](#)]
95. Ciesielska, A.; Matyjek, M.; Kwiatkowska, K. TLR4 and CD14 Trafficking and Its Influence on LPS-Induced pro-Inflammatory Signaling. *Cell. Mol. Life Sci.* **2021**, *78*, 1233–1261. [[CrossRef](#)] [[PubMed](#)]
96. Qiao, X.; Cummins, D.J.; Paul, S.M. Neuroinflammation-Induced Acceleration of Amyloid Deposition in the APP<sup>V717F</sup> Transgenic Mouse: Neuroinflammation-Induced Acceleration of Amyloid Deposition. *Eur. J. Neurosci.* **2001**, *14*, 474–482. [[CrossRef](#)] [[PubMed](#)]
97. Sheng, J. Lipopolysaccharide-Induced-Neuroinflammation Increases Intracellular Accumulation of Amyloid Precursor Protein and Amyloid  $\beta$  Peptide in APP<sup>swe</sup> Transgenic Mice. *Neurobiol. Dis.* **2003**, *14*, 133–145. [[CrossRef](#)]
98. Chen, Y.; Yin, M.; Cao, X.; Hu, G.; Xiao, M. Pro- and Anti-Inflammatory Effects of High Cholesterol Diet on Aged Brain. *Aging Dis.* **2018**, *9*, 374. [[CrossRef](#)]

99. Duong, M.T.; Nasrallah, I.M.; Wolk, D.A.; Chang, C.C.Y.; Chang, T.-Y. Cholesterol, Atherosclerosis, and APOE in Vascular Contributions to Cognitive Impairment and Dementia (VCID): Potential Mechanisms and Therapy. *Front. Aging Neurosci.* **2021**, *13*, 647990. [[CrossRef](#)]
100. Famer, D.; Wahlund, L.-O.; Crisby, M. Rosuvastatin Reduces Microglia in the Brain of Wild Type and ApoE Knockout Mice on a High Cholesterol Diet, Implications for Prevention of Stroke and AD. *Biochem. Biophys. Res. Commun.* **2010**, *402*, 367–372. [[CrossRef](#)]
101. Angelopoulou, E.; Paudel, Y.N.; Papageorgiou, S.G.; Piperi, C. APOE Genotype and Alzheimer’s Disease: The Influence of Lifestyle and Environmental Factors. *ACS Chem. Neurosci.* **2021**, *12*, 2749–2764. [[CrossRef](#)]
102. Minagawa, H.; Gong, J.-S.; Jung, C.-G.; Watanabe, A.; Lund-Katz, S.; Phillips, M.C.; Saito, H.; Michikawa, M. Mechanism Underlying Apolipoprotein E (ApoE) Isoform-Dependent Lipid Efflux from Neural Cells in Culture. *J. Neurosci. Res.* **2009**, *87*, 2498–2508. [[CrossRef](#)]
103. Iannucci, J.; Sen, A.; Grammas, P. Isoform-Specific Effects of Apolipoprotein E on Markers of Inflammation and Toxicity in Brain Glia and Neuronal Cells In Vitro. *Curr. Issues Mol. Biol.* **2021**, *43*, 215–225. [[CrossRef](#)]
104. Cantuti-Castelvetri, L.; Fitzner, D.; Bosch-Queralt, M.; Weil, M.-T.; Su, M.; Sen, P.; Ruhwedel, T.; Mitkovski, M.; Trendelenburg, G.; Lütjohann, D.; et al. Defective Cholesterol Clearance Limits Remyelination in the Aged Central Nervous System. *Science* **2018**, *359*, 684–688. [[CrossRef](#)] [[PubMed](#)]
105. Churchward, M.A.; Todd, K.G. Statin Treatment Affects Cytokine Release and Phagocytic Activity in Primary Cultured Microglia through Two Separable Mechanisms. *Mol. Brain* **2014**, *7*, 85. [[CrossRef](#)]
106. Yeh, F.L.; Wang, Y.; Tom, I.; Gonzalez, L.C.; Sheng, M. TREM2 Binds to Apolipoproteins, Including APOE and CLU/APOJ, and Thereby Facilitates Uptake of Amyloid-Beta by Microglia. *Neuron* **2016**, *91*, 328–340. [[CrossRef](#)] [[PubMed](#)]
107. Ulland, T.K.; Colonna, M. TREM2—A Key Player in Microglial Biology and Alzheimer Disease. *Nat. Rev. Neurol.* **2018**, *14*, 667–675. [[CrossRef](#)]
108. Nugent, A.A.; Lin, K.; van Lengerich, B.; Lianoglou, S.; Przybyla, L.; Davis, S.S.; Llapashtica, C.; Wang, J.; Kim, D.J.; Xia, D.; et al. TREM2 Regulates Microglial Cholesterol Metabolism upon Chronic Phagocytic Challenge. *Neuron* **2020**, *105*, 837–854.e9. [[CrossRef](#)] [[PubMed](#)]
109. Oram, J.F.; Heinecke, J.W. ATP-Binding Cassette Transporter A1: A Cell Cholesterol Exporter That Protects against Cardiovascular Disease. *Physiol. Rev.* **2005**, *85*, 1343–1372. [[CrossRef](#)] [[PubMed](#)]
110. Karasinska, J.M.; de Haan, W.; Franciosi, S.; Ruddle, P.; Fan, J.; Kruit, J.K.; Stukas, S.; Lütjohann, D.; Gutmann, D.H.; Wellington, C.L.; et al. ABCA1 Influences Neuroinflammation and Neuronal Death. *Neurobiol. Dis.* **2013**, *54*, 445–455. [[CrossRef](#)] [[PubMed](#)]
111. Tanaka, N.; Abe-Dohmae, S.; Iwamoto, N.; Fitzgerald, M.L.; Yokoyama, S. Helical Apolipoproteins of High-Density Lipoprotein Enhance Phagocytosis by Stabilizing ATP-Binding Cassette Transporter A7. *J. Lipid Res.* **2010**, *51*, 2591–2599. [[CrossRef](#)]
112. Aikawa, T.; Holm, M.-L.; Kanekiyo, T. ABCA7 and Pathogenic Pathways of Alzheimer’s Disease. *Brain Sci.* **2018**, *8*, 27. [[CrossRef](#)]
113. Dai, W.; Yao, R.-M.; Mi, T.-Y.; Zhang, L.-M.; Wu, H.; Cheng, J.-B.; Li, Y.-F. Cognition-Enhancing Effect of YL-IPA08, a Potent Ligand for the Translocator Protein (18 kDa) in the 5 × FAD Transgenic Mouse Model of Alzheimer’s Pathology. *J. Psychopharmacol.* **2022**, *36*, 1176–1187. [[CrossRef](#)]
114. Lee, J.-W.; Choi, E.-A.; Kim, Y.-S.; Kim, Y.; You, Y.H.-S.; Han, Y.-E.; Kim, H.-S.; Bae, Y.-J.; Kim, J.; Kang, H.-T. Statin Exposure and the Risk of Dementia in Individuals with Hypercholesterolaemia. *J. Intern. Med.* **2020**, *288*, 689–698. [[CrossRef](#)] [[PubMed](#)]
115. Barthold, D.; Joyce, G.; Diaz Brinton, R.; Wharton, W.; Kehoe, P.G.; Zissimopoulos, J. Association of Combination Statin and Antihypertensive Therapy with Reduced Alzheimer’s Disease and Related Dementia Risk. *PLoS ONE* **2020**, *15*, e0229541. [[CrossRef](#)] [[PubMed](#)]
116. Glodzik, L.; Santisteban, M.M. Blood-Brain Barrier Crossing Renin-Angiotensin System Drugs: Considerations for Dementia and Cognitive Decline. *Hypertension* **2021**, *78*, 644–646. [[CrossRef](#)] [[PubMed](#)]
117. Glodzik, L.; Rusinek, H.; Kamer, A.; Pirraglia, E.; Tsui, W.; Mosconi, L.; Li, Y.; McHugh, P.; Murray, J.; Williams, S.; et al. Effects of Vascular Risk Factors, Statins, and Antihypertensive Drugs on PiB Deposition in Cognitively Normal Subjects. *Alzheimers Dement. Diagn. Assess. Dis. Monit.* **2016**, *2*, 95–104. [[CrossRef](#)]
118. Dai, L.; Zou, L.; Meng, L.; Qiang, G.; Yan, M.; Zhang, Z. Cholesterol Metabolism in Neurodegenerative Diseases: Molecular Mechanisms and Therapeutic Targets. *Mol. Neurobiol.* **2021**, *58*, 2183–2201. [[CrossRef](#)]
119. Wood, W.G.; Eckert, G.P.; Igbavboa, U.; Müller, W.E. Statins and Neuroprotection: A Prescription to Move the Field Forward. *Ann. N. Y. Acad. Sci.* **2010**, *1199*, 69–76. [[CrossRef](#)]
120. Wang, S.; Zhang, X.; Zhai, L.; Sheng, X.; Zheng, W.; Chu, H.; Zhang, G. Atorvastatin Attenuates Cognitive Deficits and Neuroinflammation Induced by Aβ1–42 Involving Modulation of TLR4/TRAF6/NF-KB Pathway. *J. Mol. Neurosci.* **2018**, *64*, 363–373. [[CrossRef](#)]
121. van der Most, P.J.; Dolga, A.M.; Nijholt, I.M.; Luiten, P.G.M.; Eisel, U.L.M. Statins: Mechanisms of Neuroprotection. *Prog. Neurobiol.* **2009**, *88*, 64–75. [[CrossRef](#)]
122. Ewen, T.; Qiuting, L.; Chaogang, T.; Tao, T.; Jun, W.; Liming, T.; Guanghong, X. Neuroprotective Effect of Atorvastatin Involves Suppression of TNF-α and Upregulation of IL-10 in a Rat Model of Intracerebral Hemorrhage. *Cell Biochem. Biophys.* **2013**, *66*, 337–346. [[CrossRef](#)]
123. McFarland, A.J.; Davey, A.K.; Anoopkumar-Dukie, S. Statins Reduce Lipopolysaccharide-Induced Cytokine and Inflammatory Mediator Release in an In Vitro Model of Microglial-Like Cells. *Mediat. Inflamm.* **2017**, *2017*, 2582745. [[CrossRef](#)]

124. Bagheri, H.; Ghasemi, F.; Barreto, G.E.; Sathyapalan, T.; Jamialahmadi, T.; Sahebkar, A. The Effects of Statins on Microglial Cells to Protect against Neurodegenerative Disorders: A Mechanistic Review. *BioFactors* **2020**, *46*, 309–325. [[CrossRef](#)] [[PubMed](#)]
125. Lindberg, C.; Crisby, M.; Winblad, B.; Schultzberg, M. Effects of Statins on Microglia. *J. Neurosci. Res.* **2005**, *82*, 10–19. [[CrossRef](#)] [[PubMed](#)]
126. Clementino, A.R.; Marchi, C.; Pozzoli, M.; Bernini, F.; Zimetti, F.; Sonvico, F. Anti-Inflammatory Properties of Statin-Loaded Biodegradable Lecithin/Chitosan Nanoparticles: A Step Toward Nose-to-Brain Treatment of Neurodegenerative Diseases. *Front. Pharmacol.* **2021**, *12*, 716380. [[CrossRef](#)] [[PubMed](#)]
127. Townsend, K.P.; Shytle, D.R.; Bai, Y.; San, N.; Zeng, J.; Freeman, M.; Mori, T.; Fernandez, F.; Morgan, D.; Sanberg, P.; et al. Lovastatin Modulation of Microglial Activation via Suppression of Functional CD40 Expression. *J. Neurosci. Res.* **2004**, *78*, 167–176. [[CrossRef](#)] [[PubMed](#)]
128. Pahan, K.; Sheikh, F.G.; Namboodiri, A.M.; Singh, I. Lovastatin and Phenylacetate Inhibit the Induction of Nitric Oxide Synthase and Cytokines in Rat Primary Astrocytes, Microglia, and Macrophages. *J. Clin. Investig.* **1997**, *100*, 2671–2679. [[CrossRef](#)]
129. Takaesu, G.; Kishida, S.; Hiyama, A.; Yamaguchi, K.; Shibuya, H.; Irie, K.; Ninomiya-Tsuji, J.; Matsumoto, K. TAB2, a Novel Adaptor Protein, Mediates Activation of TAK1 MAPKKK by Linking TAK1 to TRAF6 in the IL-1 Signal Transduction Pathway. *Mol. Cell* **2000**, *5*, 649–658. [[CrossRef](#)]
130. Yongjun, Y.; Shuyun, H.; Lei, C.; Xiangrong, C.; Zhilin, Y.; Yiquan, K. Atorvastatin Suppresses Glioma Invasion and Migration by Reducing Microglial MT1-MMP Expression. *J. Neuroimmunol.* **2013**, *260*, 1–8. [[CrossRef](#)]
131. Kata, D.; Földesi, I.; Feher, L.Z.; Hackler, L.; Puskas, L.G.; Gulya, K. Rosuvastatin Enhances Anti-Inflammatory and Inhibits pro-Inflammatory Functions in Cultured Microglial Cells. *Neuroscience* **2016**, *314*, 47–63. [[CrossRef](#)]
132. Ohsawa, K.; Imai, Y.; Sasaki, Y.; Kohsaka, S. Microglia/Macrophage-Specific Protein Iba1 Binds to Fimbrin and Enhances Its Actin-Bundling Activity: Fimbrin as an Iba1-Interacting Molecule. *J. Neurochem.* **2004**, *88*, 844–856. [[CrossRef](#)]
133. Guirado, E.; Rajaram, M.V.S.; Chawla, A.; Daigle, J.; La Perle, K.M.D.; Arnett, E.; Turner, J.; Schlesinger, L.S. Deletion of PPAR $\gamma$  in Lung Macrophages Provides an Immunoprotective Response against M. Tuberculosis Infection in Mice. *Tuberculosis* **2018**, *111*, 170–177. [[CrossRef](#)]
134. Lyons, A.; Murphy, K.J.; Clarke, R.; Lynch, M.A. Atorvastatin Prevents Age-Related and Amyloid- $\beta$ -Induced Microglial Activation by Blocking Interferon- $\gamma$  Release from Natural Killer Cells in the Brain. *J. Neuroinflamm.* **2011**, *8*, 27. [[CrossRef](#)] [[PubMed](#)]
135. Chu, L.-W.; Chen, J.-Y.; Wu, P.-C.; Wu, B.-N. Atorvastatin Prevents Neuroinflammation in Chronic Constriction Injury Rats through Nuclear NF $\kappa$ B Downregulation in the Dorsal Root Ganglion and Spinal Cord. *ACS Chem. Neurosci.* **2015**, *6*, 889–898. [[CrossRef](#)] [[PubMed](#)]
136. Tamboli, I.Y.; Barth, E.; Christian, L.; Siepmann, M.; Kumar, S.; Singh, S.; Tolksdorf, K.; Heneka, M.T.; Lütjohann, D.; Wunderlich, P.; et al. Statins Promote the Degradation of Extracellular Amyloid  $\beta$ -Peptide by Microglia via Stimulation of Exosome-Associated Insulin-Degrading Enzyme (IDE) Secretion. *J. Biol. Chem.* **2010**, *285*, 37405–37414. [[CrossRef](#)]
137. Wee Yong, V.; Forsyth, P.A.; Bell, R.; Krekoski, C.A.; Edwards, D.R. Matrix Metalloproteinases and Diseases of the CNS. *Trends Neurosci.* **1998**, *21*, 75–80. [[CrossRef](#)] [[PubMed](#)]
138. Nagase, H.; Visse, R.; Murphy, G. Structure and Function of Matrix Metalloproteinases and TIMPs. *Cardiovasc. Res.* **2006**, *69*, 562–573. [[CrossRef](#)]
139. Temel, S.G.; Kahveci, Z. Cyclooxygenase-2 Expression in Astrocytes and Microglia in Human Oligodendroglioma and Astrocytoma. *J. Mol. Histol.* **2009**, *40*, 369–377. [[CrossRef](#)]
140. Petanceska, S.S.; DeRosa, S.; Sharma, A.; Diaz, N.; Duff, K.; Tint, S.G.; Refolo, L.M.; Pappolla, M. Changes in Apolipoprotein E Expression in Response to Dietary and Pharmacological Modulation of Cholesterol. *J. Mol. Neurosci.* **2003**, *20*, 395–406. [[CrossRef](#)]
141. Sodero, A.O.; Barrantes, F.J. Pleiotropic Effects of Statins on Brain Cells. *Biochim. Biophys. Acta BBA—Biomembr.* **2020**, *1862*, 183340. [[CrossRef](#)]
142. Pocivavsek, A.; Rebeck, G.W. Inhibition of C-Jun N-Terminal Kinase Increases ApoE Expression in Vitro and in Vivo. *Biochem. Biophys. Res. Commun.* **2009**, *387*, 516–520. [[CrossRef](#)]
143. Jamshidnejad-Tosaramandani, T.; Kashanian, S.; Al-Sabri, M.H.; Kročianová, D.; Clemensson, L.E.; Gentreau, M.; Schiöth, H.B. Statins and Cognition: Modifying Factors and Possible Underlying Mechanisms. *Front. Aging Neurosci.* **2022**, *14*, 968039. [[CrossRef](#)]